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## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(54) Title: REDIRECTION OF CELLULAR IMMUNITY BY RECEPTOR CHIMERAS

## (57) Abstract

Disclosed is a method of directing a cellular response in a mammal by expressing in a cell of the mammal a chimeric receptor which causes the cells to specifically recognize and destroy an infective agent, a cell infected with an infective agent, a tumor or cancerous cell, or an autoimmune-generated cell. Also disclosed are cells which express the chimeric receptors and DNA encoding the chimeric receptors.

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REDIRECTION OF CELLULAR IMMUNITY BY RECEPTOR CHIMERAS

Field of the Invention

The invention concerns functional T cell receptor,  
5 Fc receptor, or B cell receptor chimeras which are  
capable of redirecting immune system function. More  
particularly, it concerns the regulation of lymphocytes,  
macrophages, natural killer cells or granulocytes by the  
expression in said cells of chimeras which cause the  
10 cells to respond to targets recognized by the chimeras.  
The invention also concerns functional T cell receptor,  
Fc receptor, or B cell receptor chimeras which are  
capable of directing therapeutic cells to specifically  
recognize and destroy either cells infected with a  
15 specific infective agent, the infective agent itself, a  
tumor cell, or an autoimmune-generated cell. More  
particularly, the invention relates to the production of  
T cell receptor or Fc receptor chimeras capable of  
directing cytotoxic T lymphocytes to specifically  
20 recognize and lyse cells expressing HIV envelope  
proteins. The invention therefore provides a therapy for  
diseases such as AIDS (Acquired Immunodeficiency  
Syndrome) which are caused by the HIV virus.

Background of the Invention

25 T cell recognition of antigen through the T cell  
receptor is the basis of a range of immunological  
phenomena. The T cells direct what is called cell-  
mediated immunity. This involves the destruction by  
cells of the immune system of foreign tissues or infected  
30 cells. A variety of T cells exist, including "helper"  
and "suppressor" cells, which modulate the immune  
response, and cytotoxic (or "killer") cells, which can  
kill abnormal cells directly.

A T cell that recognizes and binds a unique  
35 antigen displayed on the surface of another cell becomes

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activated; it can then multiply, and if it is a cytotoxic cell, it can kill the bound cell.

Autoimmune disease is characterized by production of either antibodies that react with host tissue or 5 immune effector T cells that are autoreactive. In some instances, autoantibodies may arise by a normal T- and B-cell response activated by foreign substances or organisms that contain antigens that cross react with similar compounds in body tissues. Examples of 10 clinically relevant autoantibodies are antibodies against acetylcholine receptors in myasthenia gravis; and anti-DNA, anti-erythrocyte, and anti-platelet antibodies in systemic lupus erythematosus.

#### HIV and Immunopathogenesis

15 In 1984 HIV was shown to be the etiologic agent of AIDS. Since that time the definition of AIDS has been revised a number of times with regard to what criteria should be included in the diagnosis. However, despite the fluctuation in diagnostic parameters, the simple 20 common denominator of AIDS is the infection with HIV and subsequent development of persistent constitutional symptoms and AIDS defining diseases such as a secondary infections, neoplasms, and neurologic disease.

Harrison's Principles of Internal Medicine, 12th ed., 25 McGraw Hill (1991).

HIV is a human retrovirus of the lentivirus group. The four recognized human retroviruses belong to two distinct groups: the human T lymphotropic (or leukemia) retroviruses, HTLV-1 and HTLV-2, and the human 30 immunodeficiency viruses, HIV-1 and HIV-2. The former are transforming viruses whereas the latter are cytopathic viruses.

HIV-1 has been identified as the most common cause of AIDS throughout the world. Sequence homology between

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HIV-2 and HIV-1 is about 40% with HIV-2 being more closely related to some members of a group of simian immunodeficiency viruses (SIV). See Curran, J. et al., Science, 329:1357-1359 (1985); Weiss, R. et al., Nature, 5 324:572-575 (1986).

HIV has the usual retroviral genes (env, gag, and pol) as well as six extra genes involved in the replication and other biologic activities of the virus. As stated previously, the common denominator of AIDS is a 10 profound immunosuppression, predominantly of cell-mediated immunity. This immune suppression leads to a variety of opportunistic diseases, particularly certain infections and neoplasms.

The main cause of the immune defect in AIDS, has 15 been identified as a quantitative and qualitative deficiency in the subset of thymus-derived (T) lymphocytes, the T4 population. This subset of cells is defined phenotypically by the presence of the CD4 surface molecule, which has been demonstrated to be the cellular 20 receptor for HIV. Dalgleish et al., Nature, 312:763 (1984). Although the T4 cell is the major cell type infected with HIV, essentially any human cell that expresses the CD4 molecule on its surface is capable of binding to and being infected with HIV.

25 Traditionally, CD4<sup>+</sup> T cells have been assigned the role of helper/inducer, indicating their function in providing an activating signal to B cells, or inducing T lymphocytes bearing the reciprocal CD8 marker to become cytotoxic/suppressor cells. Reinherz and Schlossman, 30 Cell, 19:821-827 (1980); Goldstein et al., Immunol. Rev., 68:5-42, (1982).

HIV binds specifically and with high affinity, via a stretch of amino acids in the viral envelope (gp120), to a portion of the V1 region of the CD4 molecule located 35 near its N-terminus. Following binding, the virus fuses

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with the target cell membrane and is internalized. Once internalized it uses the enzyme reverse transcriptase to transcribe its genomic RNA to DNA, which is integrated into the cellular DNA where it exists for the life of the 5 cell as a "provirus."

The provirus may remain latent or be activated to transcribe mRNA and genomic RNA, leading to protein synthesis, assembly, new virion formation, and budding of virus from the cell surface. Although the precise 10 mechanism by which the virus induces cell death has not been established, it is felt that the major mechanism is massive viral budding from the cell surface, leading to disruption of the plasma membrane and resulting osmotic disequilibrium.

15 During the course of the infection, the host organism develops antibodies against viral proteins, including the major envelope glycoproteins gp120 and gp41. Despite this humoral immunity, the disease progresses, resulting in a lethal immunosuppression 20 characterized by multiple opportunistic infections, parasitemia, dementia and death. The failure of the host anti-viral antibodies to arrest the progression of the disease represents one of the most vexing and alarming aspects of the infection, and augurs poorly for 25 vaccination efforts based upon conventional approaches.

Two factors may play a role in the efficacy of the humoral response to immunodeficiency viruses. First, like other RNA viruses (and like retroviruses in particular), the immunodeficiency viruses show a high 30 mutation rate in response to host immune surveillance. Second, the envelope glycoproteins themselves are heavily glycosylated molecules presenting few epitopes suitable for high affinity antibody binding. The poorly antigenic target which the viral envelope presents, allows the host

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little opportunity for restricting viral infection by specific antibody production.

Cells infected by the HIV virus express the gp120 glycoprotein on their surface. Gp120 mediates fusion 5 events among CD4<sup>+</sup> cells via a reaction similar to that by which the virus enters the uninfected cells, leading to the formation of short-lived multinucleated giant cells. Syncytium formation is dependent on a direct interaction of the gp120 envelope glycoprotein with the CD4 protein.

10 Dalgleish et al., supra; Klatzman, D. et al., Nature, 312:763 (1984); McDougal, J.S. et al., Science, 231:382 (1986); Sodroski, J. et al., Nature, 322:470 (1986); Lifson, J.D. et al., Nature, 323:725 (1986); Sodroski, J. et al., Nature, 321:412 (1986).

15 Evidence that the CD4-gp120 binding is responsible for viral infection of cells bearing the CD4 antigen includes the finding that a specific complex is formed between gp120 and CD4. McDougal et al., supra. Other investigators have shown that the cell lines, which were 20 noninfective for HIV, were converted to infectable cell lines following transfection and expression of the human CD4 cDNA gene. Maddon et al., Cell, 46:333-348 (1986).

Therapeutic programs based on soluble CD4 as a passive agent to interfere with viral adsorption and 25 syncytium-mediated cellular transmission have been proposed and successfully demonstrated in vitro by a number of groups (Deen et al., Nature, 332:82-84 (1988); Fisher et al., Nature, 331:76-78 (1988); Hussey et al., Nature 331:78-81 (1988); Smith et al., Science, 238:1704-30 1707 (1987); Traunecker et al., Nature, 331:84-86 (1988)); and CD4 immunoglobulin fusion proteins with extended halflives and modest biological activity have subsequently been developed (Capon et al., Nature, 337:525-531 (1989); Traunecker et al. Nature, 339, 68-70 35 (1989); Byrn et al., Nature, 344:667-670 (1990);

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Zettl et al., DNA Cell Biol. 9:347-353 (1990)). Although CD4 immunotoxin conjugates or fusion proteins show potent cytotoxicity for infected cells in vitro (Chaudhary et al., Nature, 335:369-372 (1988); Till et al., Science, 242:1166-1168 (1988)), the latency of the immunodeficiency syndrome makes it unlikely that any single-treatment therapy will be effective in eliminating viral burden, and the antigenicity of foreign fusion proteins is likely to limit their acceptability in treatments requiring repetitive dosing. Trials with monkeys affected with SIV have shown that soluble CD4, if administered to animals without marked CD4 cytopenia, can reduce SIV titer and improve in vitro measures of myeloid potential (Watanabe et al., Nature, 337:267-270 (1989)). However a prompt viral reemergence was observed after treatment was discontinued, suggesting that lifelong administration might be necessary to prevent progressive immune system debilitation.

#### T Cell and Fc Receptors

Cell surface expression of the most abundant form of the T cell antigen receptor (TCR) requires the coexpression of at least 6 distinct polypeptide chains (Weiss et al., J. Exp. Med., 160:1284-1299 (1984); Orloffhashi et al., Nature, 316:606-609 (1985); Berkhout et al., J. Biol. Chem., 263:8528-8536 (1988); Sussman et al., Cell, 52:85-95 (1988)), the  $\alpha/\beta$  antigen binding chains, the three polypeptides of the CD3 complex, and  $\zeta$ . If any of the chains are absent, stable expression of the remaining members of the complex does not ensue.  $\zeta$  is the limiting polypeptide for surface expression of the complete complex (Sussman et al., Cell, 52:85-95 (1988)) and is thought to mediate at least a fraction of the cellular activation programs triggered by receptor recognition of ligand (Weissman et al., EMBO J.,

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8:3651-3656 (1989); Frank et al., Science, 249:174-177 (1990)). A 32kDa type I integral membrane homodimer,  $\zeta$  (zeta) has a 9 residue extracellular domain with no sites for N-linked glycan addition, and a 112 residue (mouse) 5 or 113 residue (human) intracellular domain (Weissman et al., Science, 238:1018-1020 (1988a); Weissman et al., Proc. Natl. Acad. Sci. USA, 85:9709-9713 (1988b)). An isoform of  $\zeta$  called  $\eta$  (eta) (Baniyash et al., J. Biol. Chem., 263:9874-9878 (1988); Orloff et al., J. Biol. Chem., 264:14812-14817 (1989)), which arises from an 10 alternate mRNA splicing pathway (Jin et al., Proc. Natl. Acad. Sci. USA, 87:3319-3233 (1990)), is present in reduced amounts in cells expressing the antigen receptor.  $\zeta$ - $\eta$  heterodimers are thought to mediate the formation of 15 inositol phosphates, as well as the receptor-initiated programmed cell death called apoptosis (Merćep et al., Science, 242:571-574 (1988); Merćep et al., Science, 246:1162-1165 (1989)).

Like  $\zeta$  and  $\eta$ , the Fc receptor-associated  $\gamma$  chain 20 is expressed in cell surface complexes with additional polypeptides, some of which mediate ligand recognition, and others of which have undefined function.  $\gamma$  (gamma) bears a homodimeric structure and overall organization very similar to that of  $\zeta$ , and is a component of both the 25 mast cell/basophil high affinity IgE receptor, Fc $\epsilon$ RI, which consists of at least three distinct polypeptide chains (Blank et al., Nature, 337:187-189 (1989); Ra et al., Nature, 241:752-754 (1989)), and one of the low affinity receptors for IgG, represented in mice by 30 Fc $\gamma$ RII $\alpha$  (RA et al., J. Biol. Chem. J. Biol. Chem., 264:15323-15327 (1989)), and in humans by the CD16 subtype expression by macrophages and natural killer cells, CD16<sub>TM</sub> (CD16 transmembrane) (Lanier et al., Nature, 342:803-805 (1989); Anderson et al., Proc. Natl. Acad. Sci. USA, 87:2274-2278 (1990)) and with a polypeptide of 35

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unidentified function (Anderson et al., Proc. Natl. Acad. Sci. USA, 87:2274-2278 (1990)). Recently it has been reported that  $\gamma$  is expressed by a mouse T cell line, CTL, in which it forms homodimers as well as  $\gamma-\zeta$  and  $\gamma-\eta$  heterodimers (Orloff et al., Nature, 347:189-191 (1990)).

The Fc receptors mediate phagocytosis of immune complexes, transcytosis, and antibody dependent cellular cytotoxicity (ADCC) (Ravetch and Kinet, Annu. Rev. Immunol. 9:457-492 (1991); Unkeless et al., Annu. Rev. Immunol. 6:251-281 (1988); and Mellman, Curr. Opin. Immunol. 1:16-25 (1988)). Recently it has been shown that one of the murine low affinity Fc receptor isoforms (FcR $\gamma$ IIIB1) mediates internalization of Ig-coated targets into clathrin coated pits, and that another low affinity receptor (Fc $\gamma$ IIIA) mediates ADCC through its association with one or more members of a small family of 'trigger molecules' (Miettinen et al., Cell 58:317-327 (1989); and Hunziker and Mellman, J. Cell Biol. 109:3291-3302 (1989)). These trigger molecules, T cell receptor (TCR)  $\zeta$  chain, TCR  $\eta$  chain, and Fc receptor  $\gamma$  chain, interact with ligand recognition domains of different immune system receptors and can autonomously initiate cellular effector programs, including cytolysis, following aggregation (Samelson et al., Cell 43:223-231 (1985); Weissman et al., Science 239:1018-1020 (1988); Jin et al., Proc. Natl. Acad. Sci. USA 87:3319-3323 (1990); Blank et al., Nature 337:187-189 (1989); Lanier et al., Nature 342:803-805 (1989); Kurosaki and Ravetch, Nature 342:805-807 (1989); Hibbs et al., Science 246:1608-1611 (1989); Anderson et al., Proc. Natl. Acad. Sci. USA 87:2274-2278 (1990); and Irving and Weiss, Cell 64: 891-901 (1991)).

In drawing parallels between the murine and human low affinity Fc receptor families, however, it has become clear that the human FcR $\gamma$ IIA and C isoforms have no

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murine counterpart. In part because of this, their function has yet to be defined.

Because humoral agents based on CD4 may have limited utility in vivo, the inventors began to explore 5 the possibility of augmenting cellular immunity to HIV. As a result they report the preparation of protein chimeras in which the extracellular domain of CD4 is fused to the transmembrane and/or intracellular domains of T cell receptor, IgG Fc receptor, or B cell receptor 10 signal transducing elements. Cytolytic T cells expressing chimeras which include an extracellular CD4 domain show potent MHC-independent destruction of cellular targets expressing HIV envelope proteins. An extremely important and novel component of this approach 15 has been the identification of single T cell receptor, or Fc receptor, and B cell receptor chains whose aggregation suffices to initiate the cellular response.

One particularly useful application of this approach has been the invention of chimeras between CD4 20 and  $\zeta$ ,  $\eta$ , or  $\gamma$  that direct cytolytic T lymphocytes to recognize and kill cells expressing HIV gp120.

#### Summary of the Invention

Although native T cell, B cell, and Fc receptors are or can be highly complicated multimeric structures 25 not lending themselves to convenient manipulation, the present invention demonstrates the feasibility of creating chimeras between the intracellular domain of any of a variety of molecules which are capable of fulfilling the task of target recognition. In particular, the 30 formation of chimeras consisting of the intracellular portion of T cell/Fc receptor zeta, eta, or gamma chains joined to the extracellular portion of a suitably engineered antibody molecule allows the target recognition-potential of an immune system cell to be

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specifically redirected to the antigen recognized by the extracellular antibody portion. Thus with an antibody portion capable of recognizing some determinant on the surface of a pathogen, immune system cells armed with the 5 chimera would respond to the presence of the pathogen with the effector program appropriate to their lineage, e.g., helper T lymphocytes would respond by cytotoxic activity against the target, and B lymphocytes would be activated to synthesize antibody. Macrophages and 10 granulocytes would carry out their effector programs, including cytokine release, phagocytosis, and reactive oxygen generation. Similarly, with an antibody portion capable of recognizing tumor cells, the immune system response to the tumor would be beneficially elevated. 15 With an antibody capable of recognizing immune cells having an inappropriate reactivity with self determinants, the autoreactive cells could be selectively targeted for destruction. Although these examples draw on the use of antibody chimeras as a convenient 20 expository tool, the invention is not limited in scope to antibody chimeras, and indeed, the use of specific nonantibody extracellular domains may have important advantages. For example with an extracellular portion that is the receptor for a virus, bacterium, or parasite, 25 cells armed with the chimeras would specifically target cells expressing the viral, bacterial or parasitic determinants. The advantage of this approach over the use of antibodies is that the native receptor for pathogen may have uniquely high selectivity or affinity 30 for the pathogen, allowing a greater degree of precision in the resulting immune response. Similarly, to delete immune system cells which inappropriately react with a self antigen, it may suffice to join the antigen (either as an intact protein, in the case of B cell depletion 35 therapies, or as MHC complex, in the case of T cell

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depletion therapies) to intracellular zeta, eta or gamma chains, and thereby affect the specific targeting of the cells inappropriately responding to self determinants.

Another use of the chimeras is the control of cell populations in vivo subsequent to other forms of genetic engineering. For example, the use of tumor infiltrating lymphocytes or natural killer cells to carry cytotoxic principles to the site of tumors has been proposed. The present invention provides a convenient means to regulate the numbers and activity of such lymphocytes and cells without removing them from the body of the patient for amplification in vitro. Thus, because the intracellular domains of the chimeric receptors mediate the proliferative responses of the cells, the coordination of the extracellular domains by a variety of aggregating stimuli specific for the extracellular domains (e.g., an antibody specific for the extracellular domain) will result in proliferation of the cells bearing the chimeras.

Although the specific embodiments of the present invention comprise chimeras between zeta, eta or gamma chains, or active fragments thereof (e.g., those discussed below), any receptor chain having a similar function to these molecules, e.g., in granulocytes or B lymphocytes, could be used for the purposes disclosed here. The distinguishing features of desirable immune cell trigger molecules comprise the ability to be expressed autonomously (i.e., as a single chain), the ability to be fused to an extracellular domain such that the resultant chimera is present on the surface of a therapeutic cell, and the ability to initiate cellular effector programs upon aggregation secondary to encounter with a target ligand.

At present the most convenient method for delivery of the chimeras to immune system cells is through some

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form of genetic therapy. However, r constituting immun system cells with chimeric receptors by mixture of the cells with suitably solubilized purified chimeric protein would also result in the formation of an engineered cell 5 population capable of responding to the targets recognized by the extracellular domain of the chimeras. Similar approaches have been used, for example, to introduce the intact HIV receptor, CD4, into erythrocytes for therapeutic purposes. In this case the engineered 10 cell population would not be capable of self renewal.

The present invention relates to functional simplified T cell receptor, B cell receptor, and Fc receptor chimeras which are capable of redirecting immune system function. More particularly, it relates to the 15 regulation of lymphocytes, macrophages, natural killer cells or granulocytes by the expression in said cells of chimeras which cause the cells to respond to targets recognized by the chimeras. The invention also relates to a method of directing cellular response to an 20 infective agent, a tumor or cancerous cell, or an autoimmune generated cell. The method for directing the cellular response in a mammal comprises administering an effective amount of therapeutic cells to said mammal, said cells being capable of recognizing and destroying 25 said infective agent, tumor, cancer cell or autoimmune generated cell.

In another embodiment, the method of directing cellular response to an infective agent comprises administering therapeutic cells capable of recognizing 30 and destroying said agent, wherein the agent is a specific virus, bacteria, protozoa, or fungi. Even more specifically, the method is directed against agents such as HIV and *Pneumocystis carinii*.

Specifically the invention provides for a method 35 of directing cellular response to an HIV infected cell.

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The method comprises administering to a patient an effective amount of cytotoxic T lymphocytes, said lymphocytes being capable of specifically recognizing and lysing cells infected with HIV.

5        Thus, in one embodiment, there is provided according to the invention a method for directing cellular response to HIV infected cells, comprising administering to a patient an effective amount of cytotoxic T lymphocytes which are capable of specifically 10 recognizing and lysing cells infected with HIV.

In yet another embodiment is provided the chimeric receptor proteins which direct the cytotoxic T lymphocytes to recognize and lyse the HIV infected cell. Yet another embodiment of the invention comprises host 15 cells transformed with a vector comprising the chimeric receptors.

In yet another embodiment, the present invention provides for an antibody against the chimeric receptors of the invention.

20        In order to obtain cytotoxic T lymphocytes which specifically bind and lyse cells infected with HIV, the present inventors therefore attempted, and herein receptor chimeras. These chimeric receptors are functionally active and possess the extraordinary ability 25 of being able to specifically bind and lyse cells expressing gp120.

It is an object of the present invention, then, to provide for a method of treatment for individuals infected with HIV. The present invention thus provides a 30 number of important advances in the therapy of AIDS.

These and other non-limiting embodiments of the present invention will be apparent to those of skill from the following detailed description of the invention.

In the following detailed description, reference 35 will be made to various methodologies known to those of

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skill in the art of molecular biology and immunology. Publications and other materials setting forth such known methodologies to which reference is made are incorporated herein by reference in their entireties as though set forth in full.

Standard reference works setting forth the general principles of recombinant DNA technology include Watson, J.D. et al., Molecular Biology of the Gene, Volumes I and II, the Benjamin/Cummings Publishing Company, Inc., publisher, Menlo Park, CA (1987); Darnell, J.E. et al., Molecular Cell Biology, Scientific American Books, Inc., Publisher, New York, N.Y. (1986); Lewin, B.M., Genes II, John Wiley & Sons, publishers, New York, N.Y. (1985); Old, R.W., et al., Principles of Gene Manipulation: An Introduction to Genetic Engineering, 2d edition, University of California Press, publisher, Berkeley, CA (1981); Maniatis, T., et al., Molecular Cloning: A Laboratory Manual, 2nd Ed. Cold Spring Harbor Laboratory, publisher, Cold Spring Harbor, NY (1989); and Current Protocols in Molecular Biology, Ausubel et al., Wiley Press, New York, NY (1989).

#### DEFINITIONS

By "cloning" is meant the use of in vitro recombination techniques to insert a particular gene or other DNA sequence into a vector molecule. In order to successfully clone a desired gene, it is necessary to employ methods for generating DNA fragments for joining the fragments to vector molecules, for introducing the composite DNA molecule into a host cell in which it can replicate, and for selecting the clone having the target gene from amongst the recipient host cells.

By "cDNA" is meant complementary or copy DNA produced from an RNA template by the action of RNA-dependent DNA polymerase (reverse transcriptase). Thus a

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"cDNA clone" means a duplex DNA sequence complementary to an RNA molecule of interest, carried in a cloning vector.

By "cDNA library" is meant a collection of recombinant DNA molecules containing cDNA inserts which 5 comprise DNA copies of mRNA being expressed by the cell at the time the cDNA library was made. Such a cDNA library may be prepared by methods known to those of skill, and described, for example, in Maniatis et al., Molecular Cloning: A Laboratory Manual, supra.

10 Generally, RNA is first isolated from the cells of an organism from whose genome it is desired to clone a particular gene. Preferred for the purpose of the present invention are mammalian, and particularly human, lymphocytic cell lines. A presently preferred vector for 15 this purpose is the vaccinia virus WR strain.

By "vector" is meant a DNA molecule, derived, e.g., from a plasmid, bacteriophage, or mammalian or insect virus, into which fragments of DNA may be inserted or cloned. A vector will contain one or more unique 20 restriction sites and may be capable of autonomous replication in a defined host or vehicle organism such that the cloned sequence is reproducible. Thus, by "DNA expression vector" is meant any autonomous element capable of directing the synthesis of a recombinant 25 peptide. Such DNA expression vectors include bacterial plasmids and phages and mammalian and insect plasmids and viruses.

By "substantially pure" is meant a compound, e.g., a protein, a polypeptide, or an antibody, that is 30 substantially free of the components that naturally accompany it. Generally, a compound is substantially pure when at least 60%, more preferably at least 75%, and most preferably at least 90% of the total material in a sample is the compound of interest. Purity can be 35 measured by any appropriate method, e.g., column

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chromatography, polyacrylamide gel electrophoresis, or HPLC analysis. In the context of a nucleic acid, "substantially pure" means a nucleic acid sequence, segment, or fragment that is free from the genes that 5 flank it in its naturally-occurring state (e.g., free from the sequences that flank the nucleic acid in its native genomic position). By "functional derivative" is meant the "fragments," "variants," "analogues," or "chemical derivatives" of a molecule. A "fragment" of a 10 molecule, such as any of the cDNA sequences of the present invention, is meant to refer to any nucleotide subset of the molecule. A "variant" of such molecule is meant to refer to a naturally occurring molecule substantially similar to either the entire molecule, or a 15 fragment thereof. An "analog" of a molecule is meant to refer to a non-natural molecule substantially similar to either the entire molecule or a fragment thereof. A molecule is said to be "substantially similar" to another molecule if the sequence of amino acids in both molecules 20 is substantially the same. Substantially similar amino acid molecules will possess a similar biological activity. Thus, provided that two molecules possess a similar activity, they are considered variants as that term is used herein even if one of the molecules contains 25 additional or fewer amino acid residues not found in the other, or if the sequence of amino acid residues is not identical. As used herein, a molecule is said to be a "chemical derivative" of another molecule when it contains additional chemical moieties not normally a part 30 of the molecule. Such moieties may improve the molecule's solubility, absorption, biological half life, etc. The moieties may alternatively decrease the toxicity of the molecule, eliminate or attenuate any undesirable side effect of the molecule, etc. Moieties 35 capable of mediating such effects are disclosed, for

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example, in Remington's Pharmaceutical Sciences, 16th et., Mack Publishing Co., Easton, Penn. (1980).

Similarly, a "functional derivative" of a receptor chimera of the present invention is meant to include

5 "fragments," "variants," or "analogues" of the gene, which may be "substantially similar" in nucleotide sequence, and which encode a molecule possessing similar activity to, for example, a T cell, B cell, or Fc receptor chimera.

10 Thus, as used herein, a T cell, B cells or Fc receptor chimera protein is also meant to include any functional derivative, fragments, variants, analogues, or chemical derivatives which may be substantially similar to the "wild-type" chimera and which possess similar 15 activity (i.e., most preferably, 90%, more preferably, 70%, preferably 40%, or at least 10% of the wild-type receptor chimera's activity). The activity of a functional chimeric receptor derivative includes specific binding (with its extracellular portion) to a targeted 20 agent or cell and resultant destruction (directed by its intracellular or transmembrane portion) of that agent or cell; such activity may be tested, e.g., using any of the assays described herein.

A DNA sequence encoding the T cell, B cell, or Fc 25 receptor chimera of the present invention, or its functional derivatives, may be recombined with vector DNA in accordance with conventional techniques, including blunt-ended or staggered-ended termini for ligation, restriction enzyme digestion to provide appropriate 30 termini, filling in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and ligation with appropriate ligases. Techniques for such manipulations are disclosed by Maniatis, T., et al., supra, and are well known in the 35 art.

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A nucleic acid molecule, such as DNA, is said to be "capable of expressing" a polypeptide if it contains nucleotide sequences which contain transcriptional and translational regulatory information and such sequences 5 are "operably linked" to nucleotide sequences which encode the polypeptide. An operable linkage is a linkage in which the regulatory DNA sequences and the DNA sequence sought to be expressed are connected in such a way as to permit gene expression. The precise nature of 10 the regulatory regions needed for gene expression may vary from organism to organism, but shall in general include a promoter region which, in prokaryotes, contains both the promoter (which directs the initiation of RNA transcription) as well as the DNA sequences which, when 15 transcribed into RNA, will signal the initiation of protein synthesis. Such regions will normally include those 5'-non-coding sequences involved with initiation of transcription and translation, such as the TATA box, capping sequence, CAAT sequence, and the like.

20 If desired, the non-coding region 3' to the gene sequence coding for the protein may be obtained by the above-described methods. This region may be retained for its transcriptional termination regulatory sequences, such as termination and polyadenylation. Thus, by 25 retaining the 3'-region naturally contiguous to the DNA sequence coding for the protein, the transcriptional termination signals may be provided. Where the transcriptional termination signals are not satisfactorily functional in the expression host cell, 30 then a 3' region functional in the host cell may be substituted.

Two DNA sequences (such as a promoter region sequence and a T cell receptor, a B cell receptor, or Fc receptor chimera encoding sequence) are said to be 35 operably linked if the nature of the linkage between the

two DNA sequences does not (1) result in the introduction of a frame-shift mutation, (2) interfere with the ability of the promoter region sequence to direct the transcription of the receptor chimera gene sequence, or 5 (3) interfere with the ability of the receptor chimera gene sequence to be transcribed by the promoter region sequence. A promoter region would be operably linked to a DNA sequence if the promoter were capable of effecting transcription of that DNA sequence. Thus, to express the 10 protein, transcriptional and translational signals recognized by an appropriate host are necessary.

The present invention encompasses the expression of a T cell receptor, B cell receptor, or Fc receptor chimera protein (or a functional derivative thereof) in 15 either prokaryotic or eukaryotic cells, although eukaryotic (and, particularly, human lymphocyte) expression is preferred.

Antibodies according to the present invention may be prepared by any of a variety of methods. For example, 20 cells expressing the receptor chimera protein, or a functional derivative thereof, can be administered to an animal in order to induce the production of sera containing polyclonal antibodies that are capable of binding the chimera.

25 In a preferred method, antibodies according to the present invention are monoclonal antibodies. Such monoclonal antibodies can be prepared using hybridoma technology (Kohler et al., Nature 256:495 (1975); Kohler et al., Eur. J. Immunol. 6:511 (1976); Kohler et al., 30 Eur. J. Immunol. 6:292 (1976); Hammerling et al., In: Monoclonal Antibodies and T-Cell Hybridomas, Elsevier, N.Y., pp. 563-684 (1981)). In general, such procedures involve immunizing an animal with the T cell receptor, B cell receptor, or Fc receptor chimera antigen. The 35 splenocytes of such animals are extracted and fused with

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a suitable myeloma cell line. Any suitable myeloma cell line may be employed in accordance with the present invention. After fusion, the resulting hybridoma cells are selectively maintained in HAT medium, and then cloned 5 by limiting dilution as described by Wands, J.R., et al. (Gastroenterology 80:225-232 (1981)). The hybridoma cells obtained through such a selection are then assayed to identify clones which secrete antibodies capable of binding the chimera.

10 Antibodies according to the present invention also may be polyclonal, or, preferably, region specific polyclonal antibodies.

Antibodies against the T cell receptor, B cell receptor, or Fc receptor chimera according to the present 15 invention may be used to monitor the amount of chimeric receptor (or chimeric receptor-bearing cells) in a patient. Such antibodies are well suited for use in standard immunodiagnostic assay known in the art, including such immunometric or "sandwich" assays as the 20 forward sandwich, reverse sandwich, and simultaneous sandwich assays. The antibodies may be used in any number of combinations as may be determined by those of skill without undue experimentation to effect immunoassays of acceptable specificity, sensitivity, and 25 accuracy.

Standard reference works setting forth general principles of immunology include Roitt, I., Essential Immunology, Sixth Ed., Blackwell Scientific Publications, Publisher, Oxford (1988); Kimball, J. W., Introduction to Immunology, Second Ed., Macmillan Publishing Co., Publisher, New York (1986); Roitt, I., et al., Immunology, Gower Medical Publishing Ltd., Publisher, London, (1985); Campbell, A. , "Monoclonal Antibody Technology," in, Burdon, R., et al., eds., Laboratory 30 Techniques in Biochemistry and Molecular Biology, Volume 35 Techniques in Biochemistry and Molecular Biology, Volume

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13, Elsevier, Publisher, Amsterdam (1984); Klein, J., Immunology: The Science of Self-Nonself Discrimination, John Wiley & Sons, Publisher, New York (1982); and Kennett, R., et al., eds., Monoclonal Antibodies, 5 Hybridoma: A New Dimension In Biological Analyses, Plenum Press, Publisher, New York (1980).

By "detecting" it is intended to include determining the presence or absence of a substance or quantifying the amount of a substance. The term thus 10 refers to the use of the materials, compositions, and methods of the present invention for qualitative and quantitative determinations.

The isolation of other hybridomas secreting monoclonal antibodies of the same specificity as those 15 described herein can be accomplished by the technique of anti-idiotypic screening (Potocnjak, et al., Science 215:1637 (1982)). Briefly, an anti-idiotypic antibody is an antibody which recognizes unique determinants present on the antibody produced by the clone of interest. The 20 anti-idiotypic antibody is prepared by immunizing an animal of the same strain used as the source of the monoclonal antibody with the monoclonal antibody of interest. The immunized animal will recognize and respond to the idotypic determinants of the immunizing 25 antibody by producing antibody to these idotypic determinants (anti-idiotypic antibody).

For replication, the hybrid cells may be cultivated both in vitro and in vivo. High in vivo production makes this the presently preferred method of 30 culture. Briefly, cells from the individual hybrid strains are injected intraperitoneally into pristane-primed BALB/c mice to produce ascites fluid containing high concentrations of the desired monoclonal antibodies. Monoclonal antibodies of isotype IgM or IgG may be 35 purified from cultured supernatants using column

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chromatography methods well known to those of skill in the art.

Antibodies according to the present invention are particularly suited for use in immunoassays wherein they 5 may be utilized in liquid phase or bound to a solid phase carrier. In addition, the antibodies in these immunoassays can be detectably labeled in various ways.

There are many different labels and methods of labeling known in the art. Examples of the types of 10 labels which can be used in the present invention include, but are not limited to, enzymes, radioisotopes, fluorescent compounds, chemiluminescent compounds, bioluminescent compounds and metal chelates. Those of ordinary skill in the art will know of other suitable 15 labels for binding to antibodies, or will be able to ascertain the same by the use of routine experimentation. Furthermore, the binding of these labels to antibodies can be accomplished using standard techniques commonly known to those of ordinary skill in the art.

20 One of the ways in which antibodies according to the present invention can be detectably labeled is by linking the antibody to an enzyme. This enzyme, in turn, when later exposed to its substrate, will react with the substrate in such a manner as to produce a chemical 25 moiety which can be detected as, for example, by spectrophotometric or fluorometric means. Examples of enzymes which can be used to detectably label antibodies include malate dehydrogenase, staphylococcal nuclease, delta-V-steroid isomerase, yeast alcohol dehydrogenase, 30 alpha-glycerophosphate dehydrogenase, triose phosphate isomerase, biotinavidin peroxidase, horseradish peroxidase, alkaline phosphatase, asparaginase, glucose oxidase,  $\beta$ -galactosidase, ribonuclease, urease, catalase, glucose-VI-phosphate dehydrogenase, glucoamylase and 35 acetylcholine esterase.

The presence of detectably labeled antibodies also can be detected by labeling the antibodies with a radioactive isotope which then can be determined by such means as the use of a gamma counter or a scintillation counter. Isotopes which are particularly useful for the purpose of the present invention are  $^3\text{H}$ ,  $^{125}\text{I}$ ,  $^{32}\text{P}$ ,  $^{35}\text{S}$ ,  $^{14}\text{C}$ ,  $^{51}\text{Cr}$ ,  $^{36}\text{Cl}$ ,  $^{57}\text{Co}$ ,  $^{58}\text{Co}$ ,  $^{59}\text{Fe}$  and  $^{75}\text{Se}$ .

It is also possible to detect the binding of detectably labeled antibodies by labeling the antibodies with a fluorescent compound. When a fluorescently labeled antibody is exposed to light of the proper wavelength, its presence then can be detected due to the fluorescence of the dye. Among the most commonly used fluorescent labeling compounds are fluorescein,  $^{152}\text{Eu}$  isothiocyanate, rhodamine, phycoerythrin, phycocyanin, allophycocyanin,  $\alpha$ -phthaldehyde and fluorescamine.

The antibodies of the invention also can be detectably labeled using fluorescent emitting metals such as  $^{152}\text{Eu}$ , or others of the lanthanide series. These metals can be attached to the antibody molecule using such metal chelating groups as diethylenteriaminepentaacetic acid (DTPA) or ethylenediaminetetraacetic acid (EDTA).

Antibodies also can be detectably labeled by coupling them to a chemiluminescent compound. The presence of the chemiluminescent-tagged antibody is then determined by detecting the presence of luminescence that arises during the course of the chemical reaction. Examples of particularly useful chemiluminescent labeling compounds are luminal, isoluminol, theromatic acridinium ester, imidazole, acridinium salts, oxalate ester, and dioxetane.

Likewise, a bioluminescent compound may be used to label the antibodies according to the present invention. Bioluminescence is a type of chemiluminescence found in

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biological systems in which a catalytic protein increases the efficiency of the chemiluminescent reaction. The presence of a bioluminescent antibody is determined by detecting the presence of luminescence. Important 5 bioluminescent compounds for purposes of labeling include luciferin, luciferase aequorin.

The antibodies and substantially purified antigen of the present invention are ideally suited for the preparation of a kit. Such a kit may comprise a carrier 10 means being compartmentalized to receive in close confinement therewith one or more container means such as vials, tubes and the like, each of said container means comprising the separate elements of the assay to be used.

The types of assays which can be incorporated in 15 kit form are many, and include, for example, competitive and non-competitive assays. Typical examples of assays which can utilize the antibodies of the invention are radioimmunoassays (RIA), enzyme immunoassays (EIA), enzyme-linked immunosorbent assays (ELISA), and 20 immunometric, or sandwich, immunoassays.

By the term "immunometric assay" or "sandwich immunoassay," it is meant to include simultaneous sandwich, forward sandwich and reverse sandwich immunoassays. These terms are well understood by those 25 skilled in the art. Those of skill will also appreciate that antibodies according to the present invention will be useful in other variations and forms of assays which are presently known or which may be developed in the future. These are intended to be included within the 30 scope of the present invention.

In the preferred mode for performing the assays it is important that certain "blockers" be present in the incubation medium (usually added with the labeled soluble antibody). The "blockers" are added to assure that non- 35 specific proteins, protease, or human antibodies to mouse

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immunoglobulins present in the experimental sample do not cr ss-link or destroy the antibodies on the solid phase support, or the radiolabeled indicator antibody, to yield false positive or false negative results. The selection 5 of "blockers" therefore adds substantially to the specificity of the assays described in the present invention.

It has been found that a number of nonrelevant (i.e., nonspecific) antibodies of the same class or 10 subclass (isotype) as those used in the assays (e.g., IgG<sub>1</sub>, IgG<sub>2a</sub>, IgM, etc.) can be used as "blockers." The concentration of the "blockers" (normally 1-100  $\mu$ g/ $\mu$ l) is important, in order to maintain the proper sensitivity yet inhibit any unwanted interference by mutually 15 occurring cross reactive proteins in human serum. In addition, the buffer system containing the "blockers" needs to be optimized. Preferred buffers are those based on weak organic acids, such as imidazole, HEPES, MOPS, TES, ADA, ACES, HEPES, PIPES, TRIS, and the like, at 20 physiological pH ranges. Somewhat less preferred buffers are inorganic buffers such as phosphate, borate or carbonate. Finally, known protease inhibitors should be added (normally at 0.01-10  $\mu$ g/ml) to the buffer which contains the "blockers."

25 There are many solid phase immunoadsorbents which have been employed and which can be used in the present invention. Well known immunoadsorbents include glass, polystyrene, polypropylene, dextran, nylon and other materials, in the form of tubes, beads, and microtiter 30 plates formed from or coated with such materials, and the like. The immobilized antibodies can be either covalently or physically bound to the solid phase immunoadsorbent, by techniques such as covalent bonding via an amide or ester linkage, or by absorption. Those 35 skilled in the art will know many other suitable solid

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phase immunoadsorbents and methods for immobilizing antibodies thereon, or will be able to ascertain such, using no more than routine experimentation.

For in vivo, in vitro, or in situ diagnosis,

5 labels such as radionuclides may be bound to antibodies according to the present invention either directly or by using an intermediary functional group. An intermediary group which is often used to bind radioisotopes which exist as metallic cations to antibodies is

10 diethylenetriaminepentaacetic acid (DTPA). Typical examples of metallic cations which are bound in this manner are:  $^{99m}\text{Tc}$ ,  $^{123}\text{I}$ ,  $^{111}\text{IN}$ ,  $^{131}\text{I}$ ,  $^{97}\text{Ru}$ ,  $^{67}\text{Cu}$ ,  $^{67}\text{Ga}$  and  $^{68}\text{Ga}$ . The antibodies of the invention can also be labeled with non-radioactive isotopes for purposes of diagnosis.

15 Elements which are particularly useful in this manner are  $^{157}\text{Gd}$ ,  $^{55}\text{Mn}$ ,  $^{162}\text{Dy}$ ,  $^{52}\text{Cr}$  and  $^{56}\text{Fe}$ .

The antigen of the invention may be isolated in substantially pure form employing antibodies according to the present invention. Thus, an embodiment of the 20 present invention provides for substantially pure T cell receptor, B cell receptor, or Fc receptor chimera, said antigen characterized in that it is recognized by and binds to antibodies according to the present invention. In another embodiment, the present invention provides a 25 method of isolating or purifying the receptor chimeric antigen, by forming a complex of said antigen with one or more antibodies directed against the receptor chimera.

The substantially pure T cell receptor, B cell receptor, or Fc receptor chimera antigens of the present 30 invention may in turn be used to detect or measure antibody to the chimera in a sample, such as serum or urine. Thus, one embodiment of the present invention comprises a method of detecting the presence or amount of antibody to receptor chimera antigen in a sample, 35 comprising contacting a sample containing an antibody to

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the chimeric antigen with detectably labeled receptor chimera, and detecting said label. It will be appreciated that immunoreactive fractions and immunoreactive analogues of the chimera also may be used.

5 By the term "immunoreactive fraction" is intended any portion of the chimeric antigen which demonstrates an equivalent immune response to an antibody directed against the receptor chimera. By the term "immunoreactive analogue" is intended a protein which 10 differs from the receptor chimera protein by one or more amino acids, but which demonstrates an equivalent immunoresponse to an antibody of the invention.

By "specifically recognizes and binds" is meant an antibody which recognizes and binds a chimeric receptor 15 polypeptide but which does not substantially recognize and bind other molecules in a sample, e.g., in a biological sample, which includes the receptor polypeptide.

By "autoimmune-generated cell" is meant cells 20 producing antibodies that react with host tissue or immune effector T cells that are autoreactive; such cells include antibodies against acetylcholine receptors (leading, e.g., to myasthenia gravis) or anti-DNA, anti-erythrocyte, and anti-placelet autoantibodies (leading, 25 e.g., to lupus erythematosus).

By "therapeutic cell" is meant a cell which has been transformed by a chimera of the invention so that it is capable of recognizing and destroying a specific infective agent, a cell infected by a specific agent, a 30 tumor or cancerous cell, or an autoimmune-generated cell; preferably such therapeutic cells are cells of the hematopoietic system.

By "extracellular" is meant having at least a portion of the molecule exposed at the cell surface. By 35 "intracellular" is meant having at least a portion of the

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molecule exposed to the therapeutic cell's cytoplasm. By "transmembrane" is meant having at least a portion of the molecule spanning the plasma membrane. An "extracellular portion", an "intracellular portion" and a "transmembrane portion", as used herein, may include flanking amino acid sequences which extend into adjoining cellular compartments.

By "oligomerize" is meant to complex with other proteins to form dimers, trimers, tetramers, or other 10 higher order oligomers. Such oligomers may be homo-oligomers or hetero-oligomers. An "oligomerizing portion" is that region of a molecule which directs complex (i.e., oligomer) formation.

By "cytolytic" is meant to be capable of 15 destroying a cell (e.g., a cell infected with a pathogen, a tumor or cancerous cell, or an autoimmune-generated) cell or to be capable of destroying an infective agent (e.g., a virus).

By "immunodeficiency virus" is meant a retrovirus 20 that, in wild-type form, is capable of infecting T4 cells of a primate host and possesses a viral morphogenesis and morphology characteristic of the lentivirus subfamily. The term includes, without limitation, all variants of HIV and SIV, including HIV-1, HIV-2, SIVmac, SIVagm, 25 SIVmnd, SIVsmm, SIVman, SIVmand, and SIVcpz.

By "MHC-independent" is meant that the cellular cytolytic response does not require the presence of an MHC class II antigen on the surface of the targeted cell.

By a "functional cytolytic signal-transducing 30 derivative" is meant a functional derivative (as defined above) which is capable of directing at least 10%, preferably 40%, more preferably 70%, or most preferably at least 90% of the biological activity of the wild type molecule. As used herein, a "functional cytolytic 35 signal-transducing derivative" may act by directly

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signaling the therapeutic cell to destroy a receptor-bound agent or cell (e.g., in the case of an intracellular chimeric receptor portion) or may act indirectly by promoting oligomerization with cytolytic 5 signal transducing proteins of the therapeutic cell (e.g., in the case of a transmembrane domain). Such derivatives may be tested for efficacy, e.g., using the in vitro assays described herein.

By a "functional HIV envelope-binding derivative" 10 is meant a functional derivative (as defined above) which is capable of binding any HIV envelope protein. Functional derivatives may be identified using, e.g., the in vitro assays described herein.

#### **THERAPEUTIC ADMINISTRATION**

15 The transformed cells of the present invention may be used for the therapy of a number of diseases. Current methods of administering such transformed cells involve adoptive immunotherapy or cell-transfer therapy. These methods allow the return of the transformed immune-system 20 cells to the bloodstream. Rosenberg, S.A., Scientific American, 62 (May 1990); Rosenberg et al., The New England Journal of Medicine, 323(9):570 (1990).

The pharmaceutical compositions of the invention may be administered to any animal which may experience 25 the beneficial effects of the compounds of the invention. Foremost among such animals are humans, although the invention is not intended to be so limited.

#### Detailed Description

The drawings will first be described.

30 Brief Description of the Drawings

**FIG. 1** Characterization of CD4 chimeras. Fig. 1A presents the amino acid sequence about the site of fusion

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betw en CD4 (residues 1-369) and the different  $\gamma$  ceptor chains. The underlined sequence shows the position of the amino acids encoded within the BamHI site used for fusion construction. The beginning of the transmembrane 5 domain is marked with a vertical bar. The  $\eta$  sequence is identical to the  $\zeta$  sequence at the amino terminus, but diverges at the carboxyl terminus (Jin et al., Proc. Natl. Acad. Sci. USA, 87:3319-3323 (1990)). Fig. 1B presents flow cytometric analysis of surface expression 10 of CD4, CD4: $\zeta$ , CD4: $\gamma$  and CD4: $\eta$  in CV1 cells. Cells were infected with virus expressing CD4 chimeras or CD16<sub>TM</sub>, incubated for 9 hours at 37°C, and stained with phycoerythrin-conjugated anti-CD4 MAb Leu3A. Fig. 1C presents immunoprecipitation of labeled CD4: $\zeta$ , CD4: $\gamma$ , or 15 native CD4 expressed in CV1 cells. Lanes were run with reducing (R) or without reducing (NR) agent. Molecular mass standards in kD are shown at left.

FIG. 2 Surface expression of CD16<sub>TM</sub> following coinfection of CD16<sub>TM</sub> alone (dense dots), or coinfecte 20 with virus expressing CD4: $\gamma$  (dashes) or CD4: $\zeta$  (solid line). Sparse dots, cells infected with CD4: $\zeta$  alone, stained with 3G8 (Fleit et al., Proc. Natl. Acad. Sci. USA, 79:3275-3279 (1982)) (anti-CD16 MAb).

FIG. 3 Mutant CD4: $\zeta$  chimeric receptors lacking  $\zeta$  25 Asp-15 do not support the coexpression of CD16<sub>TM</sub>. Fig. 3A is an autoradiogram of immunoprecipitated mutant chimeras electrophoresed either with reduction (R) or without reduction (NR). Fig. 3B details surface expression of CD16<sub>TM</sub> following coinfection by viruses expressing CD16<sub>TM</sub> 30 and the following  $\zeta$  chimeras: CD4: $\zeta$  (thick line), CD4: $\zeta$  C11G (solid line); CD4: $\zeta$  (dashed line); CD4: $\zeta$  C11G/D15G (dense dots); no coinfection (CD16<sub>TM</sub> alone, sparse dots). Cells were incubated with anti-CD16 MAb 3G8 and phycoerythrin- conjugated Fab'<sub>2</sub> goat antibodies to mouse 35 IgG. The level of expression of the  $\zeta$  chimeras was

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essentially identical for the different mutants analyzed, and coinfection of cells with viruses expressing CD16<sub>TM</sub> and  $\zeta$  chimeras did not appreciably alter surface expression of the chimeras (data not shown).

5       FIG. 4 Increased intracellular free calcium ion follows crosslinking of mutant  $\zeta$  chimeras in a T cell line. Jurkat E6 cells (Weiss et al., J. Immunol., 133:123-128 (1984)) were infected with recombinant vaccinia viruses and analyzed by flow cytometry. The 10 results shown are for the gated CD4<sup>+</sup> population, so that only cells expressing the relevant chimeric protein are analyzed. The mean ratio of violet to blue Indo-1 fluorescence reflects the intracellular free calcium concentration in the population as a whole and the 15 percentage of responding cells reflects the fraction of cells which exceed a predetermined threshold ratio (set so that 10% of untreated cells are positive). Fig. 4A and Fig. 4B show Jurkat cells expressing CD4: $\zeta$  (solid line) or CD16: $\zeta$  (dashed line) which were exposed to anti- 20 CD4 MAb Leu3a (phycoerythrin conjugate), followed by crosslinking with goat antibody to mouse IgG. The dotted line shows the response of uninfected cells to anti-CD3 MAb OKT3. Figs. 4C and 4D show Jurkat cells expressing CD4: $\zeta$ D15G (solid line); CD4: $\zeta$ C11G/D15G (dashes); or 25 CD4; $\zeta$ C11G (dots) which were treated and analyzed as in Figs. 4A and 4B.

FIG. 5 CD4: $\zeta$ , CD4: $\eta$ , and CD4: $\gamma$  receptors allow cytolytic T lymphocytes (CTL) to kill targets expressing HIV-1 gp120/41. Fig. 5A: solid circles, CTL expressing 30 CD4: $\zeta$  incubated with HeLa cells expressing gp120/41; open circles, CTL expressing CD4: $\zeta$  incubated with uninfected HeLa cells; solid squares, uninfected CTL incubated with HeLa cells expressing gp120/41; open squares, uninfected CTL incubated with uninfected HeLa cells. Fig. 5B: 35 solid circles, CTL expressing CD4: $\eta$  incubated with HeLa

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cells expressing gp120/41; open circles, CTL expressing CD4:γ, incubated with HeLa cells expressing gp120/41; open squares, CTL expressing the C11G/D15G double mutant CD4:γ chimera incubated with HeLa cells expressing gp120/41.

5 Fig. 5C: Flow cytometric analysis of CD4 expression by the CTL used in Fig. 5B. To correct the target to effector ratios the percent of cells expressing CD4 chimera was determined by subtracting the scaled negative (uninfected) population by histogram superposition; for 10 comparative purposes in this figure the uninfected cells were assigned an arbitrary threshold which gives roughly the same fraction positive for the other cell populations as would histogram subtraction.

**FIG. 6 Specificity of the CD4-directed cytolysis.**

15 Fig. 6A: solid circles, CTL expressing CD4:γ incubated with HeLa cells expressing CD16<sub>PI</sub>; open circles, CTL expressing CD4 incubated with HeLa cells expressing gp120; solid squares, CTL expressing CD16:γ incubated with HeLa cells expressing gp120/41; open squares, CTL 20 expressing CD16<sub>PI</sub> incubated with HeLa cells expressing gp120/41. Fig. 6B: solid circles, CTL expressing CD4:γ incubated with Raji (MHC class II<sup>+</sup>) cells; open circles, uninfected CTL cells incubated with RJ2.2.5 (MHC class II<sup>-</sup> Raji mutant) cells; solid squares, uninfected CTL 25 incubated with Raji (MHC class II<sup>+</sup>) cells; open squares, CTL expressing CD4:γ incubated with RJ2.2.5 (MHC class II<sup>-</sup>) cells. The ordinate scale is expanded.

**FIG. 7 Characterization of the CD16:γ chimeric receptor.** Fig. 7A is a schematic diagram of the CD16:γ 30 fusion protein. The extracellular portion of the phosphatidylinositol-linked form of monomeric CD16 was joined to dimeric γ just external to the transmembrane domain. The protein sequence at the fusion junction is shown at the bottom. Fig. 7B shows a flow cytometric 35 analysis of calcium mobilization following crosslinking

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of the CD16: $\zeta$  chimera in either a TCR positive or TCR positive or TCR negative cell line. The mean ratio of violet to blue fluorescence (a measure of relative calcium ion concentration) among cell populations treated 5 with antibodies at time 0 is shown. Solid squares, the response of Jurkat cells to anti-CD3 MAb OKT3; solid triangles, the response of CD16: $\zeta$  to anti-CD16 MAb 3G8 crosslinking in the REX33A TCR $^{-}$  mutant; open squares, the response to CD16: $\zeta$  crosslinking in the Jurkat TCR $^{-}$  mutant 10 line JRT3.T3.5; open triangles, the response to CD16: $\zeta$  crosslinking in Jurkat cells; crosses, the response to nonchimeric CD16 in Jurkat cells; and dots, the response to nonchimeric CD16 in the REX33A TCR $^{-}$  cell line.

FIG. 8 Deletion analysis of cytolytic potential.

15 Fig. 8A shows the locations of the  $\zeta$  deletion endpoints. Here as elsewhere mutations in  $\zeta$  are represented by the original residue-location-mutant residue convention, so that D66\*, for example, denotes replacement of Asp-66 by a termination codon. Fig. 8B shows cytolysis assay 20 results of undeleted CD16: $\zeta$  and salient  $\zeta$  deletions. Hybridoma cells expressing surface antibody to CD16 were loaded with  $^{51}\text{Cr}$  and incubated with increasing numbers of human cytolytic lymphocytes (CTL) infected with vaccinia recombinants expressing CD16: $\zeta$  chimeras. The percent of 25  $^{51}\text{Cr}$  released is plotted as a function of the effector (CTL) to target (hybridoma) cell ratio (e/t). Solid circles, cytolysis mediated by cells expressing CD16: $\zeta$  (mfi 18.7); solid squares, cytolysis mediated by cells expressing CD16: $\zeta$  Asp66\* (mfi 940.2); open squares, 30 cytolysis mediated by cells expressing CD16: $\zeta$  Glu60\* (mfi 16.0); open circles, cytolysis mediated by cells expressing CD16: $\zeta$  Tyr51\* (mfi 17.4); solid triangles, cytolysis mediated by cells expressing CD16: $\zeta$  Phe34\* (mfi 17.8); and open triangles, cytolysis mediated by cells 35 expressing nonchimeric CD16 (mfi 591). Although in this

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experiment the expression of CD16: $\zeta$ Asp66\* was not matched to that of the other fusion proteins, cytolysis by cells expressing CD16: $\zeta$  at equivalent levels in the same experiment gave results essentially identical to those shown by cells expressing CD16: $\zeta$ Asp66\* (not shown).

FIG. 9 Elimination of the potential for transmembrane interactions reveals a short  $\zeta$  segment capable of mediating cytolysis. Fig. 9A is a schematic diagram of the monomeric bipartite and tripartite chimeras. At the top is the CD16: $\zeta$  construct truncated at residue 65 and lacking transmembrane Cys and Asp residues. Below are the CD16:CD5: $\zeta$  and CD16:CD7: $\zeta$  constructs and related controls. The peptide sequences of the intracellular domains are shown below. Fig. 9B shows the cytolytic activity of monomeric chimera deletion mutants. The cytolytic activity of cells expressing CD16: $\zeta$  (solid circles; mfi 495) was compared to that of cells expressing CD16: $\zeta$ Asp66\* (solid squares; mfi 527) or the mutants CD16: $\zeta$ Cys11Gly/Asp15Gly/Asp66\*, (open squares; mfi 338) and CD16: $\zeta$ Cys11Gly/Asp15Gly/Glu60\* (filled triangles; mfi 259). Fig. 9C shows the cytolytic activity mediated by tripartite fusion proteins. Solid triangles, CD16: $\zeta$ Asp66\*; open squares, CD16:5: $\zeta$ (48-65); solid squares CD16:7: $\zeta$ (48-65); open triangles, CD16:7: $\zeta$ (48-59); open circles, CD16:5; solid circles, CD16:7. Fig. 9D shows calcium mobilization by mutant and tripartite chimeras in the TCR negative Jurkat JRT3.T3.5 mutant cell line. Open circles, response of cells expressing dimeric CD16: $\zeta$ Asp66\*; solid squares, response of cells expressing CD16: $\zeta$ Cys11Gly/Asp15Gly/Asp66\*; open squares, response of cells expressing CD16: $\zeta$ Cys11Gly/Asp15Gly/Glu60\*; solid triangles, response of cells expressing CD16:7: $\zeta$ (48-65); and open triangles, response of cells expressing CD16: $\zeta$ (48-59).

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**FIG. 10** Contribution of individual amino acids to the activity of the 18 residue cytolytic signal-transducing motif. Figs. 10A and 10B show cytolytic activity and Fig. 10C shows calcium ion mobilization mediated by chimeras bearing point mutations near the carboxyl terminal tyrosine (Y62). Figs. 10A and 10B represent data collected on cells expressing low and high amounts, respectively, of the CD16: $\zeta$  fusion proteins. Identical symbols are used for the calcium mobilization and cytolysis assays, and are shown in one letter code at right. Solid circles, cells expressing CD16: $\zeta$  (mfi in A, 21; B, 376); solid squares, cells expressing CD16:7: $\zeta$  (48-65) (mfi A, 31; B, 82); open squares, CD16:7: $\zeta$  (48-65)Glu60Gln (mfi A, 33; B, 92), crosses, CD16:7: $\zeta$  (48-65)Asp63Asn (mfi A, 30; B, 74); solid triangles, CD16:7: $\zeta$  (48-65)Tyr62Phe (mfi A, 24; B, 88); open circles, CD16:7: $\zeta$  (48-65)Glu61Gln (mfi A, 20; B, 62); and open triangles, CD16:7: $\zeta$  (48-65)Tyr62Ser (mfi B, 64). Figs. 10D and 10E show cytolytic activity and Fig. 10F shows calcium ion mobilization by chimeras bearing point mutations near the amino terminal tyrosine (Y51). Identical symbols are used for the calcium mobilization and cytolysis assays and are shown at right. Solid circles, cells expressing CD16: $\zeta$  (mfi in D, 21.2; in E, 672); solid squares, cells expressing CD16:7: $\zeta$  (48-65) (mfi D, 31.3; E, 179); solid triangles, CD16:7: $\zeta$  (48-65)Asn48Ser (mfi D, 22.4; E, 209); open squares, CD16:7: $\zeta$  (48-65)Leu50Ser (mfi D, 25.0; E, 142); and open triangles, CD16:7: $\zeta$  (48-65)Tyr51Phe (mfi D, 32.3; E, 294).

**FIG. 11** Alignment of internal repeats of  $\zeta$  and comparison of their ability to support cytolysis. Fig. 11A is a schematic diagram of chimeras formed by dividing the  $\zeta$  intracellular domain into thirds and appending them to the transmembrane domain of a CD16:7 chimera. The sequences of the intracellular domains are shown below,

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with shared residues boxed, and related residues denoted by asterisks. Fig. 11B shows the cytolytic potency of the three  $\zeta$  subdomains. Solid circles, cells expressing CD16: $\zeta$  (mfi 476); solid squares, CD16:7: $\zeta$ (33-65) (mfi 68); open squares, CD16:7: $\zeta$ (71-104) (mfi 114); and solid triangles, CD16:7: $\zeta$ (104-138) (mfi 104).

FIG. 12 is a schematic diagram of the CD16:FcR $\gamma$ II chimeras.

FIG. 13 Calcium mobilization following crosslinking of CD4:FcR $\gamma$ II and CD16:FcR $\gamma$ II chimeras. Fig. 13A shows the ratio of violet to blue fluorescence emitted by cells loaded with the calcium sensitive fluorophore Indo-1 shown as a function of time following crosslinking of the CD16 extracellular domain with antibodies. Fig. 13B shows a similar analysis of the increase in ratio of violet to blue fluorescence of cells bearing CD4:FcR $\gamma$ II chimeras, following crosslinking with antibodies.

FIG. 14 Cytolysis assay of CD4:FcR $\gamma$ II and CD16:FcR $\gamma$ II chimeras. Fig. 14A shows the percent of  $^{51}\text{Cr}$  released from anti-CD16 hybridoma (target) cells when the cells are exposed to increasing numbers of cytotoxic T lymphocytes expressing CD16:FcR $\gamma$ II chimeras (effector cells). Fig. 14B shows a similar analysis of cytotoxicity mediated by CD4:FcR $\gamma$ II chimeras against target cells expressing HIV envelope glycoproteins.

FIG. 15 Identification of residues in the FcR $\gamma$ II A tail which are important for cytolysis. Fig. 15A is a schematic diagram of the deletion constructs. Figs. 15B and 15C shows calcium mobilization and cytolysis by carboxyl-terminal deletion variants of CD16:FcR $\gamma$ II A. Figs. 15D and 15E show calcium mobilization and cytolysis by tripartite chimeras bearing progressively less of the amino terminus of the intracellular tail of CD16:FcR $\gamma$ II A.

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FIG. 16 (SEQ ID NO: 24) shows the amino acid sequence of the CD3 delta receptor protein; the boxed sequence represents a preferred cytolytic signal transducing portion.

5 FIG. 17 (SEQ ID NO: 25) shows the amino acid sequence of the T3 gamma receptor protein; the boxed sequence represents a preferred cytolytic signal transducing portion.

10 FIG. 18 (SEQ ID NO: 26) shows the amino acid sequence of the mbl receptor protein; the boxed sequence represents a preferred cytolytic signal transducing portion.

15 FIG. 19 (SEQ ID NO: 27) shows the amino acid sequence of the B29 receptor protein; the boxed sequence represents a preferred cytolytic signal transducing portion.

#### EXAMPLE I

##### Construction of Human IgG1:Receptor Chimeras

Human IgG1 heavy chain sequences were prepared by 20 joining sequences in the C<sub>H</sub>3 domain to a cDNA fragment derived from the 3' end of the transmembrane form of the antibody mRNA. The 3' end fragment was obtained by polymerase chain reaction using a tonsil cDNA library as substrate, and oligonucleotides having the sequences:

25 CGC GGG GTG ACC GTG CCC TCC AGC AGC TTG GGC (SEQ ID NO: 7) and

CGC GGG GAT CCG TCG TCC AGA GCC CGT CCA GCT CCC CGT CCT GGG CCT CA (SEQ ID NO: 8), corresponding to the 5' and 3' ends of the desired DNA 30 fragments respectively. The 5' oligo is complementary to a site in the C<sub>H</sub>1 domain of human IgG1, and the 3' oligo is complementary to a site just 5' of the sequences encoding the membrane spanning domain. The PCR product was digested with BstXI and BamHI and ligated between

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BstXI and BamHI sites of a semisynthetic IgG1 antibody  
bearing variable and constant regions. Following  
the insertion of the BstXI to BamHI fragment, the  
amplified portions of the construct were replaced up to  
5 the SmaI site in C<sub>H</sub>3 by restriction fragment interchange,  
so that only the portion between the SmaI site and the 3'  
oligo was derived from the PCR reaction.

To create a human IgG1:γ chimeric receptor, the  
heavy chain gene ending in a BamHI site was joined to the  
10 BamHI site of the γ chimera described below, so that the  
antibody sequences formed the extracellular portion.  
Flow cytometry of COS cells transfected with a plasmid  
encoding the chimera showed high level expression of  
antibody determinants when an expression plasmid encoding  
15 a light chain cDNA was cotransfected, and modest  
expression of antibody determinants when the light chain  
expression plasmid was absent.

Similar chimeras including human IgG1 fused to η  
or γ (see below), or any signal-transducing portion of a  
20 T cell receptor or Fc receptor protein may be constructed  
generally as described above using standard techniques of  
molecular biology.

To create a single transcription unit which would  
allow both heavy and light chains to be expressed from a  
25 single promoter, a plasmid encoding a bicistronic mRNA  
was created from heavy and light chain coding sequences,  
and the 5' untranslated portion of the mRNA encoding the  
78kD glucose regulated protein, otherwise known as grp78,  
or BiP. grp78 sequences were obtained by PCR of human  
30 genomic DNA using primers having the sequences:

CGC GGG CGG CCG CGA CGC CGG CCA AGA CAG CAC (SEQ  
ID NO: 9) and

CGC GTT GAC GAG CAG CCA GTT GGG CAG CAG CAG (SEQ  
ID NO: 10)

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at the 5' and 3' ends respectively. Polymerase chain reactions with these oligos were performed in the presence of 10% dimethyl sulfoxide. The fragment obtained by PCR was digested with NotI and HincII and 5 inserted between NotI and HpaI sites downstream from human IgG1 coding sequences. Sequences encoding a human IgG kappa light chain cDNA were then inserted downstream from the grp78 leader, using the HincII site and another site in the vector. The expression plasmid resulting 10 from these manipulations consisted of the semisynthetic heavy chain gene, followed by the grp78 leader sequences, followed by the kappa light chain cDNA sequences, followed by polyadenylation signals derived from an SV40 DNA fragment. Transfection of COS cells with the 15 expression plasmid gave markedly improved expression of heavy chain determinants, compared to transfection of plasmid encoding heavy chain determinants alone.

To create a bicistronic gene comprising a heavy chain/receptor chimera and a light chain, the upstream 20 heavy chain sequences can be replaced by any chimeric heavy chain/receptor gene described herein.

#### EXAMPLE II

##### Construction of CD4 Receptor Chimeras

Human  $\zeta$  (Weissman et al., Proc. Natl. Acad. Sci. 25 USA, 85:9709-9713 (1988b)) and  $\gamma$  (Küster et al., J. Biol. Chem., 265:6448-6452 (1990)) cDNAs were isolated by polymerase chain reaction from libraries prepared from the HPB-ALL tumor cell line (Aruffo et al., Proc. Natl. Acad. Sci. USA, 84:8573-8577 (1987b)) and from human 30 natural killer cells, while  $\eta$  cDNA (Jin et al., Proc. Natl. Acad. Sci. USA, 87:3319-3323 (1990)) was isolated from a murine thymocyte library.  $\zeta$ ,  $\eta$  and  $\gamma$  cDNAs were joined to the extracellular domain of an engineered form of CD4 possessing a BamHI site just upstream of the

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membrane spanning domain (Aruffo et al., Proc. Natl. Acad. Sci. USA, 84:8573-8577 (1987b); Zettlmeissl et al., DNA Cell Biol., 9:347-353 (1990)) which was joined to the BamHI site naturally present in the  $\zeta$  and  $\eta$  cDNAs at a 5 similar location a few residues upstream of the membrane spanning domain (SEQ ID NOS: 1, 3, 4 and 6). To form the fusion protein with  $\gamma$  a BamHI site was engineered into the sequence at the same approximate location (Fig. 1; SEQ ID NO: 2 and 5). The gene fusions were introduced 10 into a vaccinia virus expression plasmid bearing the E. coli gpt gene as a selectable marker (M. Amiot and B.S., unpublished), and inserted into the genome of the vaccinia WR strain by homologous recombination and selection for growth in mycophenolic acid (Falkner et 15 al., J. Virol., 62:1849-1854 (1988); Boyle et al., Gene, 65:123-128 (1988)). Flow cytometric analysis showed that the vaccinia recombinants direct the abundant production of CD4: $\zeta$  and CD4: $\gamma$  fusion proteins at the cell surface, whereas the expression of CD4: $\eta$  is substantially weaker 20 (Fig. 1). The latter finding is consistent with a recent report that transfection of an  $\eta$  cDNA expression plasmid into a murine hybridoma cell line gave substantially less expression than transfection of a comparable  $\zeta$  expression plasmid (Clayton et al., J. Exp. Med., 172:1243-1253 25 (1990)). Immunoprecipitation of cells infected with the vaccinia recombinants revealed that the fusion proteins form covalent dimers, unlike the naturally occurring CD4 antigen (Fig. 1). The molecular masses of the monomeric CD4: $\zeta$  and CD4: $\gamma$  fusion proteins and native CD4 were found 30 to be 63, 55 and 53 kD respectively. The larger masses of the fusion proteins are approximately consistent with the greater length of the intracellular portion, which exceeds that of native CD4 by 75 (CD4: $\zeta$ ) or 5 (CD4: $\gamma$ ) residues.

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#### EXAMPLE III

##### CD4 Chimeras Can Associate With Other Receptor Chains

Cell surface expression of the macrophage/natural killer cell form of human Fc $\gamma$ RIII (CD16<sub>TM</sub>) on

5 transfectants is facilitated by cotransfection with murine (Kurosaki et al., Nature, 342:805-807 (1989)) or human (Hibbs et al., Science, 246:1608-1611 (1989))  $\gamma$ , as well as by human  $\zeta$  (Lanier et al., Nature, 342:803-805 (1989)).

10 Consistent with these reports, expression of the chimeras also allowed surface expression of CD16<sub>TM</sub>) when delivered to the target cell either by cotransfection or by coinfection with recombinant vaccinia viruses (Fig.

2). The promotion of (CD16<sub>TM</sub>) surface expression by  $\zeta$  was 15 more pronounced than promotion by  $\gamma$  (Fig. 2) in the cell lines examined, whereas native CD4 (data not shown) did not enhance CD16<sub>TM</sub> surface expression.

#### EXAMPLE IV

##### Asp $\zeta$ Mutants Do Not Coassociate with Fc Receptor

20 To create chimeras which would not associate with existing antigen or Fc receptors, mutant  $\zeta$  fusion proteins which lacked either the intramembranous Asp or intramembranous Cys residue or both were prepared. Flow cytometry showed that the intensity of cell surface

25 expression by the different mutant chimeras was not appreciably different from the unmutated precursor (data not shown) and immunoprecipitation experiments showed that total expression by the chimeras was similar (Fig. 3). As expected, the mutant chimeras lacking the

30 transmembrane cysteine residue were found not to form disulfide linked dimers (Fig. 3). The two mutant chimeras lacking Asp were incapable of supporting the surface expression of CD16<sup>TM</sup>, whereas the monomeric chimeras lacking Cys but bearing Asp allowed CD16<sub>TM</sub> to be

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coexpressed, but at lower efficiency than the parental dimer (Fig. 3).

EXAMPLE V

**Mutant Receptors Retain the Ability to Initiate a Calcium 5 Response**

To determine whether crosslinking of the fusion proteins would allow the accumulation of free intracellular calcium in a manner similar to that known to occur with the T cell antigen receptor, cells of the 10 human T cell leukemia line, Jurkat E6 (ATCC Accession Number TIB 152, American Type Culture Collection, Rockville, MD), were infected with the vaccinia recombinants and the relative cytoplasmic calcium concentration following crosslinking of the extracellular 15 domain with antibodies was measured. Flow cytometric measurements were performed with cells loaded with the calcium sensitive dye Indo-1 (Grynkiewicz et al., J. Biol. Chem., 260:3340-3450 (1985); Rabinovitch et al., J. Immunol., 137:952-961 (1986)). Figure 4 shows the 20 results of calcium flux experiments with cells infected with CD4: $\zeta$  and the Asp $^{-}$  and Cys $^{-}$  mutants of  $\zeta$ . Crosslinking of the chimeras, reproducibly increased intracellular calcium. CD4: $\eta$  and CD4: $\gamma$  similarly allowed accumulation intracellular calcium in infected cells 25 (data not shown). Jurkat cells express low levels of CD4 on the cell surface, however, crosslinking of the native CD4 in the presence or absence of CD16: $\zeta$  (C.R. and B.S. unpublished) (Fig. 4 and data not shown) does not alter intracellular calcium levels.

30

EXAMPLE VI

**CD4: $\zeta$ ,  $\eta$ , and  $\gamma$  Chimeras Mediate Cytolysis of Targets Expressing HIV gp120/41**

To determine whether the chimeric receptors would trigger cytolytic effector programs, a model

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target: effector system based on CD4 recognition of the HIV env lop gp120/gp41 complex was created. HeLa cells were infected with recombinant vaccinia viruses expressing gp120/gp41 (Chakrabarti et al., Nature, 5 320:535-537 (1986); Earl et al., J. Virol., 64:2448-2451 (1990)) and labeled with  $^{51}\text{Cr}$ . The labeled cells were incubated with cells from a human allospecific (CD8 $^{+}$ , CD4 $^{-}$ ) cytotoxic T lymphocyte line which had been infected with vaccinia recombinants expressing the CD4: $\zeta$ , CD4: $\eta$ , 10 or CD4: $\gamma$  chimeras, or the CD4: $\zeta$ Cys11Gly:Asp15Gly double mutant chimera. Fig. 5 shows that HeLa cells expressing gp120/41 were specifically lysed by cytotoxic T lymphocytes (CTL) expressing CD4 chimeras. Uninfected HeLa cells were not targeted by CTL armed with CD4: $\zeta$  15 chimeras, and HeLa cells expressing gp120/41 were not recognized by uninfected CTL. To compare the efficacy of the various chimeras, the effector to target ratios were corrected for the fraction of CTL expressing CD4 chimeras, and for the fraction of HeLa cells expressing 20 gp120/41, as measured by flow cytometry. Fig. 5C shows a cytometric analysis of CD4 expression by the CTL used in the cytolysis experiment shown in Figs. 4A and 4B. Although the mean density of surface CD4: $\zeta$  greatly exceeded the mean density of CD4: $\eta$ , the cytolytic 25 efficiencies of cells expressing either form were similar. Correcting for the fraction of targets expressing gp120, the efficiency of cytolysis mediated by CD4: $\zeta$  and CD4: $\eta$  proteins are comparable to the best efficiencies reported for specific T cell receptor 30 target:effector pairs (the mean effector to target ratio for 50% release by T cells expressing CD4: $\zeta$  was  $1.9 \pm 0.99$ , n=10). The CD4: $\gamma$  fusion was less active, as was the CD4: $\zeta$  fusion lacking the transmembrane Asp and Cys residues. However in both cases significant cytolysis 35 was observed (Fig. 5).

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To control for the possibility that vaccinia infection might promote artefactual recognition by CTL, similar cytolysis experiments were performed with target cells infected with vaccinia recombinants expressing the 5 phosphatidylinositol linked form of CD16 (CD16<sub>PI</sub>) and labeled with <sup>51</sup>Cr, and with CTL infected with control recombinants expressing either CD16<sub>PI</sub> or CD16:5. Fig. 6A shows that T cells expressing non-CD4 chimeras do not recognize native HeLa cells or HeLa cells expressing 10 gp120/41, and similarly that T cells expressing CD4 chimeras do not recognize HeLa cells expressing other vaccinia-encoded surface proteins. In addition, CTLs expressing non-chimeric CD4 do not significantly lyse HeLa cells expressing gp120/41 (Fig. 6A).

15

#### EXAMPLE VII

##### MHC Class II-Bearing Cells Are Not Targeted by the Chimeras

CD4 is thought to interact with a nonpolymorphic sequence expressed by MHC class II antigen (Gay et al., 20 Nature, 328:626-629 (1987); Sleckman et al., Nature, 328:351-353 (1987)). Although a specific interaction between CD4 and class II antigen has never been documented with purified proteins, under certain conditions adhesion between cells expressing CD4 and 25 cells expressing class II molecules can be demonstrated (Doyle et al., Nature, 330:256-259 (1987); Clayton et al., J. Exp. Med., 172:1243-1253 (1990); Lamarre et al., Science, 245:743-746 (1989)). Next examined was whether killing could be detected against cells bearing class II. 30 Fig. 6B shows that there is no specific cytolysis directed by CD4:5 against the Raji B cell line, which expresses abundant class II antigen. Although a modest ( $\approx$ 5%) cytolysis is observed, a class II-negative mutant of Raji, RJ2.2.5, (Accolla, R.S., J. Exp. Med., 157:1053-

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1058 (1983)) shows a similar susceptibility, as do Raji cells incubated with uninfected T cells.

#### EXAMPLE VIII

##### Sequence Requirements for Induction of Cytolysis by the T 5 Cell Antigen/Fc Receptor Zeta Chain

Although chimeras between CD4 and  $\zeta$  can arm cytotoxic T lymphocytes (CTL) to kill target cells expressing HIV gp120, an alternative to CD4 was sought in order to unambiguously compare the properties of zeta 10 chimeras introduced into human T cell lines. Such lines can express CD4, making it difficult to specifically define the relationship between the type or degree of calcium mobilization and the cytotoxic potential of the different chimeras. To circumvent this, chimeras were 15 created between  $\zeta$  and CD16 in which the extracellular domain of CD16 is attached to the transmembrane and intracellular sequences of  $\zeta$  (Fig. 7A). The gene fusions were introduced into a vaccinia virus expression plasmid bearing the E. coli gpt gene as a selectable marker and 20 inserted into the genome of the vaccinia WR strain by homologous recombination and selection for growth in mycophenolic acid (Falkner and Moss, J. Virol. 62:1849 (1988); Boyle and Coupar, Gene 65:123 (1988)).

T cell lines were infected with the vaccinia 25 recombinants and the relative cytoplasmic free calcium ion concentration was measured following crosslinking of the extracellular domains with antibodies. Both spectrofluorimetric (bulk population) and flow cytometric (single cell) measurements were performed, with cells 30 loaded with the dye Indo-1 (Grynkiewicz et al., J. Biol. Chem. 260:3440 (1985); Rabinovitch et al., J. Immunol. 137:952 (1986)). Figure 7B shows an analysis of data collected from cells of the Jurkat human T cell leukemia line infected with vaccinia recombinants expressing

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CD16:γ fusion protein. Crosslinking of the chimeras reproducibly increased intracellular calcium, while similar treatment of cells expressing nonchimeric CD16 had little or no effect. When the chimera was expressed 5 in mutant cell lines lacking antigen receptor, either REX33A (Breitmeyer et al. J. Immunol. **138**:726 (1987); Sancho et al. J. Biol. Chem. **264**:20760 (1989)), or Jurkat mutant JRT3.T3.5 (Weiss et al., J. Immunol. **135**:123 (1984)); or a strong response to CD16 antibody 10 crosslinking was seen. Similar data have been collected on the REX20A (Breitmeyer et al., supra, 1987; Blumberg et al., J. Biol. Chem. **265**:14036 (1990)) mutant cell line, and a CD3/Ti negative mutant of the Jurkat cell line established in this laboratory (data not shown). 15 Infection with recombinants expressing CD16:γ did not restore the response to anti-CD3 antibody, showing that the fusion protein did not act by rescuing intracellular CD3 complex chains (data not shown).

To evaluate the ability of the chimeras to 20 redirect cell-mediated immunity, CTLs were infected with vaccinia recombinants expressing CD16 chimeras and used to specifically lyse hybridoma cells expressing membrane-bound anti-CD16 antibodies (see below). This assay is an extension of a hybridoma cytotoxicity assay originally 25 developed to analyze effector mechanisms of cells bearing Fc receptors (Graziano and Fanger, J. Immunol. **138**:945, 1987; Graziano and Fanger, J. Immunol. **139**:35-36, 1987; Shen et al., Mol. Immunol. **26**:959, 1989; Fanger et al., Immunol. Today **10**: 92, 1989). Fig. 8B shows that 30 expression of CD16:γ in cytotoxic T lymphocytes allows the armed CTL to kill 3G8 (anti-CD16; Fleit et al., Proc. Natl. Acad. Sci. USA **79**:3275, 1982) hybridoma cells, whereas CTL expressing the phosphatidylinositol-linked form of CD16 are inactive. CTL armed with CD16:γ also do

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not kill hybridoma cells expressing an irrelevant antibody (data not shown).

To identify the minimal  $\zeta$  sequences necessary for cytotoxicity, a series of deletion mutants were prepared in which successively more of the  $\zeta$  intracellular domain was removed from the carboxyl terminus (Fig. 8A). Most of the intracellular domain of zeta could be removed with little consequence for cytolytic potential; the full length chimera CD16: $\zeta$  was essentially equal in efficacy to the chimera deleted to residue 65, CD16: $\zeta$ Asp66\* (Fig. 8B). A substantial decrease in cytotoxicity was observed on deletion to  $\zeta$  residue 59 (chimera CD16: $\zeta$ Glu60\*), and further deletion to residue 50 resulted in slightly less activity. However complete loss of activity was not observed even when the intracellular domain was reduced to a three residue transmembrane anchor (Fig. 8B).

Because  $\zeta$  is a disulfide linked dimer, one explanation for the retention of cytolytic activity was that endogenous  $\zeta$  was forming heterodimers with the chimeric  $\zeta$  deletion, thereby reconstituting activity. To test this idea,  $\zeta$  residues 11 and 15 were changed from Asp and Cys respectively to Gly (Cys11Gly/Asp15Gly), and immunoprecipitations were carried out as follows. Approximately  $2 \times 10^6$  CV1 cells were infected for one hour in serum free DME medium with recombinant vaccinia at a multiplicity of infection (moi) of at least ten. Six to eight hours post-infection, the cells were detached from the plates with PBS/1mM EDTA and surface labeled with 0.2 mCi  $^{125}\text{I}$  per  $2 \times 10^6$  cells using lactoperoxidase and  $\text{H}_2\text{O}_2$  by the method of Clark and Einfeld (*Leukocyte Typing II*, pp. 155-167, Springer=Verlag, NY, 1986). The labeled cells were collected by centrifugation and lysed in 1% NP-40, 0.1% SDS, 0.15M NaCl, 0.05M Tris, pH 8.0, 5mM  $\text{MgCl}_2$ , 5mM KCl, 0.2M iodoacetamide and 1mM PMSF. Nuclei were removed by

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centrifugation, and CD16 proteins were immunoprecipitated with antibody 3G8 (Flit et al., *supra*, 1982; Medarex) and anti-mouse IgG agarose (Cappel, Durham, NC). Samples were electrophoresed through an 8% polyacrylamide/SDS gel 5 under non-reducing conditions or through a 10% gel under reducing conditions. These immunoprecipitations confirmed that the CD16:; $\zeta$ Cys11Gly/Asp15Gly chimera did not associate in disulfide-linked dimer structures.

The cytolytic activity of the mutant receptors was 10 also tested. The mutated chimera deleted to residue 65 (CD16:; $\zeta$ Cys11Gly/Asp15Gly/Asp66\*) was, depending on the conditions of assay, two to eight fold less active in the cytolysis assay than the comparable unmutated chimera (CD16:; $\zeta$ Asp66\*), which was usually within a factor of two 15 of, or indistinguishable in activity from, CD16:; $\zeta$  (Fig. 9B). The reduction in activity of the mutant chimeras is comparable to the reduction seen with CD4 chimeras of similar structure (see above) and is most likely attributable to the lower efficiency of  $\zeta$  monomers 20 compared to dimers. In contrast, the Asp<sup>-</sup>, Cys<sup>-</sup> mutated chimera deleted to residue 59 had no cytolytic activity (Fig. 9B), supporting the hypothesis that association with other chains mediated by the transmembrane Cys and/or Asp residues was responsible for the weak 25 persistence of cytolytic activity in deletions more amino terminal than residue 65.

Flow cytometric studies showed that the deletion mutants lacking transmembrane Asp and Cys residues could still promote an increase in free intracellular calcium 30 ion in response to antibody crosslinking in a TCR<sup>-</sup> mutant Jurkat cell line (Fig. 9D). Similar results were obtained for chimeras expressed in the parental Jurkat line (not shown). In the case of CD16:; $\zeta$ Cys11Gly/Asp15Gly/Glu60\*, these data demonstrate 35 that the ability to mediate calcium responsiveness can be

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mutationally separated from the ability to support cytolysis.

To definitively eliminate the possible contribution of  $\zeta$  transmembrane residues, the 5 transmembrane and first 17 cytoplasmic residues of  $\zeta$  were replaced by sequences encoding the membrane spanning and first 14 or first 17 cytoplasmic residues of the CD5 or CD7 antigens, respectively (Fig. 9A). The resulting tripartite fusion proteins CD16:5: $\zeta$ (48-65) and 10 CD16:7: $\zeta$ (48-65) did not form disulfide-linked dimers as do the simpler CD16: $\zeta$  chimeras, because they lacked the cysteine residue in the  $\zeta$  transmembrane domain. Both tripartite chimeras were able to mobilize calcium in Jurkat and TCR negative cell lines (Fig. 9D) and to mount 15 a cytolytic response in CTL (Fig. 9C and data not shown). However truncation of the  $\zeta$  portion to residue 59 in chimera CD16:7: $\zeta$ (48-59) abrogates the ability of tripartite fusion to direct calcium responsiveness in TCR positive or negative Jurkat cells or cytolysis in mature 20 CTL (Fig. 9C and 9D and data not shown).

To examine the contributions of individual residues within the 18-residue motif, we prepared a number of mutant variants by site-directed mutagenesis, and evaluated their ability to mediate receptor-directed 25 killing under conditions of low (Figs. 10A and 10D) or high (Figs. 10B and 10E) expression of chimeric receptor. Fig. 10 shows that while a number of relatively conservative substitutions (i.e., replacing acidic residues with their cognate amides, or tyrosine with 30 phenylalanine) which spanned residues 59 to 63 yielded moderate compromise of cytolytic efficacy, in general the variants retained the ability to mobilize calcium. However collectively these residues comprise an important submotif inasmuch as their deletion eliminates cytolytic 35 activity. Conversion of Tyr 62 to either Phe or Ser

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eliminated both the cytotoxic and calcium responses. At the amino terminus of the 18 residue segment, replacement of Tyr 51 with Phe abolished both calcium mobilization and cytolytic activity, while substitution of Leu with 5 Ser at position 50 eliminated the calcium response while only partially impairing cytolysis. Without being bound to a particular hypothesis, it is suspected that the inability of the Leu50Ser mutant to mobilize calcium in short term flow cytometric assays does not fully reflect 10 its ability to mediate a substantial increase in free intracellular calcium ion over the longer time span of the cytolysis assay. However, calcium-insensitive cytolytic activity has been reported for some cytolytic T cell lines, and the possibility that a similar phenomenon 15 underlies the results described herein has not been ruled out. Replacement of Asn48 with Ser partially impaired cytotoxicity in some experiments while having little effect in others.

To investigate the potential role of redundant 20 sequence elements, the intracellular domain of  $\zeta$  was divided into three segments, spanning residues 33 to 65, 71 to 104, and 104 to 138. Each of these segments was attached to a CD16:CD7 chimera by means of a MluI site introduced just distal to the basic membrane anchoring 25 sequences of the intracellular domain of CD7 (see below; Fig. 11A). Comparison of the cytolytic efficacy of the three elements showed they were essentially equipotent (Fig. 11B). Sequence comparison (Fig. 11A) shows that the second motif bears eleven residues between tyrosines, 30 whereas the first and third motifs bear ten.

Although a precise accounting of the process of T cell activation has not been made, it is clear that aggregation of the antigen receptor, or of receptor chimeras which bear  $\zeta$  intracellular sequences, triggers 35 calcium mobilization, cytokine and granule release, and

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the appearance of cell surface markers of activation. The active site of  $\zeta$ , a short linear peptide sequence probably too small to have inherent enzymatic activity, likely interacts with one or at most a few proteins to 5 mediate cellular activation. It is also clear that mobilization of free calcium is not by itself sufficient for cellular activation, as the ability to mediate cytolysis can be mutationally separated from the ability to mediate calcium accumulation.

10 As shown herein, addition of 18 residues from the intracellular domain of  $\zeta$  to the transmembrane and intracellular domain of two unrelated proteins allows the resulting chimeras to redirect cytolytic activity against target cells which bind to the extracellular portion of 15 the fusion proteins. Although chimeras bearing the 18 residue motif are approximately eight-fold less active than chimeras based on full length  $\zeta$ , the reduced activity can be attributed to the loss of transmembrane interactions which normally allow wild-type  $\zeta$  to form 20 disulfide linked dimers. That is,  $\zeta$  deletion constructs which have the same carboxyl terminus as the motif and lack transmembrane Cys and Asp residues typically show slightly less activity than chimeras bearing only the 18 residue motif.

25 The cytolytic competency element on which we have focused has two tyrosines and no serines or threonines, restricting the possible contributions of phosphorylation to activity. Mutation of either tyrosine destroys activity, however, and although preliminary experiments 30 do not point to a substantial tyrosine phosphorylation following crosslinking of chimeric surface antigens bearing the 18 residue motif, the possible participation of such phosphorylation at a low level cannot be excluded. In addition to the effects noted at the two 35 tyrosine residues, a number of amino acid replacements at

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the amino and carboxyl terminus of the motif weaken activity under conditions of low receptor density.

Sequences similar to the  $\zeta$  active motif can be found in the cytoplasmic domains of several other 5 transmembrane proteins, including the CD3  $\delta$  and  $\gamma$  molecules, the surface IgM associated proteins mb1 and B29, and the  $\beta$  and  $\gamma$  chains of the high affinity IgE receptor, Fc $\epsilon$ RI (Reth, Nature 338:383, 1989). Although 10 the function of these sequences is uncertain, if efficiently expressed, each may be capable of autonomous T cell activation, and such activity may explain the residual TCR responsiveness seen in a zeta-negative 15 mutant cell line (Sussman et al., Cell 52:85, 1988).

$\zeta$  itself bears three such sequences, approximately 15 equally spaced, and a rough trisection of the intracellular domain shows that each is capable of initiating a cytolytic response.  $\eta$ , a splice isoform of  $\zeta$  (Jin et al., supra, 1990; Clayton et al., Proc. Natl. Acad. Sci. USA 88:5202, 1991), lacks the carboxyl half of 20 the third motif. Because removal of the carboxyl half of the first motif abolishes activity, it appears likely that the majority of the biological effectiveness of  $\eta$  can be attributed to the first two motifs. Although by different measures  $\eta$  is equally as active as  $\zeta$  in 25 promoting antigen-mediated cytokine release (Bauer et al., Proc. Natl. Acad. Sci. USA 88:3842, 1991) or redirected cytolysis (see above),  $\eta$  is not phosphorylated in response to receptor stimulation (Bauer et al., supra, 1991). Thus either the presence of all three motifs is 30 required for phosphorylation, or the third motif represents a favored substrate for an unidentified tyrosine kinase.

#### EXAMPLE IX

#### Cytolytic Signal Transduction by Human Fc Receptor

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To evaluate the actions of different human Fc receptor subtypes, chimeric molecules were created in which the extracellular domain of the human CD4, CD5 or CD16 antigens were joined to the transmembrane and 5 intracellular domains of the FcRII $\gamma$ A, B1, B2, and C subtypes (nomenclature of Ravetch and Kinet, Ann. Rev. Immunol. 9:457, 1991). Specifically, cDNA sequences corresponding to the transmembrane and cytoplasmic domains of the previously described FcRIIA, B1, and B2 10 isoforms were amplified from the preexisting clone PC23 or from a human tonsil cDNA library (constructed by standard techniques) using oligonucleotide primers:

15 CCC GGA TCC CAG CAT GGG CAG CTC TT (SEQ ID NO: 18;  
FcRIIA forward)  
CGC GGG GCG GCC GCT TTA GTT ATT ACT GTT GAC ATG  
GTC GTT (SEQ ID NO: 19; FcRIIA reverse);  
GCG GGG GGA TCC CAC TGT CCA AGC TCC CAG CTC TTC  
ACC G (SEQ ID NO: 20; FcRIIB1 and FcRIIB2 forward); and  
GCG GGG GCG GCC GCT TAA ATA CGG TTC TGG TC (SEQ ID  
20 NO: 21; FcRIIB1 and FcRIIB2 reverse).

These primers contained cleavage sites for the enzymes BamHI and NotI, respectively, indented 6 residues from the 5' end. The NotI site was immediately followed by an antisense stop codon, either CTA or TTA. All primers 25 contained 18 or more residues complementary to the 5' and 3' ends of the desired fragments. The cDNA fragment corresponding to the FcRII $\gamma$ C cytoplasmic domain, which differs from the IIA isoform in only one amino acid residue (L for P at residue 268) was generated by site 30 directed mutagenesis by overlap PCR using primers of sequence:

TCA GAA AGA GAC AAC CTG AAG AAA CCA ACA A (SEQ ID  
NO: 22) and  
TTG TTG GTT TCT TCA GGT TGT GTC TTT CTG A (SEQ ID  
35 NO: 23).

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The PCR fragments were inserted into vaccinia virus expression vectors which contained the CD16 or CD4 extracellular domains respectively and subsequently inserted into wild type vaccinia by recombination at the 5 thymidine kinase locus, using selection for cointegration of E. coli gpt to facilitate identification of the desired recombinants. The identities of all isoforms (shown in Fig. 12) were confirmed by dideoxy sequencing.

Production of the chimeric receptor proteins was 10 further confirmed by immunoprecipitation studies.

Approximately  $10^7$  JRT3.T3.5 cells were infected for one hour in serum free IMDM medium with recombinant vaccinia at a multiplicity of infection of at least ten. Twelve hours post-infection, the cells were harvested and 15 surface labeled with 0.5mCi  $^{125}\text{I}$  per  $10^7$  cells using the lactoperoxidase/glucose oxidase method (Clark and Einfeld, supra). The labeled cells were collected by centrifugation and lysed 1% NP-40, 0.1mM  $\text{MgCl}_2$ , 5mM KCl, 0.2M iodoacetamide and 1mM PMSF. Nuclei were removed by 20 centrifugation, and CD16 fusion proteins immunoprecipitated with antibody 4G8 and anti-mouse IgG agarose. Samples were electrophoresed under reducing conditions. All immunoprecipitated chimeric receptor molecules were of the expected molecular masses.

25 To test the ability of the chimeric receptors to mediate an increase in cytoplasmic free calcium ion, the recombinant viruses were used to infect the TCR $^{-}$  mutant Jurkat cell line JRT3.T3.5 (as described herein) and cytoplasmic free calcium was measured in the cells (as 30 described herein) following crosslinking of the receptor extracellular domains with monoclonal antibody 3G8 or Leu-3A (as described herein). These experiments revealed that the intracellular domains of FcR $\gamma$ II A and C were capable of mediating an increase in cytoplasmic free 35 calcium ion after crosslinking of the extracellular

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domains, whereas the intracellular domains of FcR $\gamma$ II B1 and B2 were inactive under comparable conditions (Fig. 13 A and 13B). The CD4, CD5 and CD16 hybrids of FcR $\gamma$ II A shared essentially equal capacity to promote the calcium 5 response (Fig. 13 and data not shown). Other cell lines, from both monocytic and lymphocytic lineages, were capable of responding to the signal initiated by crosslinking of the extracellular domains (data not shown).

10 To explore the involvement of the different FcR $\gamma$ II intracellular domains in cytolysis, human cytotoxic T lymphocytes (CTL) were infected with vaccinia recombinants expressing CD16:FcR $\gamma$ II A, B1, B2 and C chimeras. The infected cells were then cocultured with 15  $^{51}\text{Cr}$ -loaded hybridoma cells (i.e., 3G8 10-2 cells) which expressed cell surface antibody to CD16. In this assay CTLs bearing the CD16 chimera killed the hybridoma target cells (allowing release of free  $^{51}\text{Cr}$ ) if the CD16 extracellular domain of the chimera has been joined to an 20 intracellular segment capable of activating the lymphocyte effector program; this cytolysis assay is described in detail below. Fig. 14A shows that CTL armed with CD16:FcR $\gamma$ IIA and C, but not FcR $\gamma$ II B1 or B2, are capable of lysing target cells expressing cell surface 25 anti-CD16 antibody.

To eliminate the possibility that the specific cytolysis was in some way attributable to interaction with the CD16 moiety, cytolysis experiments were conducted in which the FcRII intracellular domains were 30 attached to a CD4 extracellular domain. In this case the target cells were HeLa cells expressing HIV envelope gp120/41 proteins (specifically, HeLa cells infected with the vaccinia vector vPE16 (available from the National Institute of Allergy and Infectious Disease AIDS 35 Depository, Bethesda, MD). As in the CD16 system, target

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cells expressing HIV envelope were susceptible to lysis by T cells expressing the CD4:FcR $\gamma$ II A chimera, but not FcR $\gamma$ II B1 or B2 (Fig. 14B).

The intracellular domains of FcR $\gamma$ II A and C share 5 no appreciable sequence homology with any other protein, including the members of the extended FcR $\gamma$ /TCR $\zeta$  family. To define the sequence elements responsible for induction of cytolysis, 5' and 3' deletions of the intracellular domain coding sequences (described below and shown in 10 Fig. 15A) were prepared and were evaluated for efficacy in calcium mobilization and cytolysis assays (as described herein). In the experiments in which the amino terminal portion of the intracellular domain was removed, the transmembrane domain of FcR $\gamma$ II was replaced with the 15 transmembrane domain of the unrelated CD7 antigen to eliminate the possible contribution of interactions mediated by the membrane-spanning domain.

Figs. 15B and 15C show that removal of the 14 carboxyl-terminal residues, including tyrosine 298, 20 resulted in a complete loss of cytolytic capacity and a substantial reduction in calcium mobilization potential. Further deletion to just before tyrosine 282 gave an identical phenotype (Figs. 15B and 15C). Deletion from the N-terminus of the intracellular domain to residue 268 25 had no substantial effect on either calcium profile or cytolytic potency, whereas deletion to residue 275 markedly impaired free calcium release but had little effect on cytolysis (Figs. 15D and 15E). Further deletion, to residue 282, gave FcR $\gamma$ II tails which lacked 30 the ability to either mobilize calcium or trigger cytolysis (Figs. 15D and 15E). The 'active element' defined by these crude measures is relatively large (36 amino acids) and contains two tyrosines separated by 16 residues.

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#### EXAMPLE X

Other intracellular and transmembrane signal transducing domains according to the invention may be derived from the T cell receptor proteins, CD3 delta and 5 T3 gamma, and the B cell receptor proteins, mb1 and B29. The amino acid sequences of these proteins is shown in Fig. 16 (CD3 delta; SEQ ID NO: 24), Fig. 17 (T3 gamma; SEQ ID NO: 25), Fig. 18 (mb1; SEQ ID NO: 26) and Fig. 19 (B29; SEQ ID NO: 27). The portions of the sequences 10 sufficient for cytolytic signal transduction (and therefore preferably included in a chimeric receptor of the invention) are shown in brackets. Chimeric receptors which include these protein domains are constructed and used in the therapeutic methods of the invention 15 generally as described above.

#### EXAMPLE XI

##### Experimental Methods

###### Vaccinia Infection and Radioimmunoprecipitation

Approximately  $5 \times 10^6$  CV1 cells were infected for 20 one hour in serum free DME medium with recombinant vaccinia at a multiplicity of infection (moi) of at least ten (titer measured on CV1 cells). The cells were placed in fresh medium after infection and labelled metabolically with  $200\mu\text{Ci}/\text{ml}$   $^{35}\text{S}$ -methionine plus cysteine 25 (Tran $^{35}\text{S}$ -label, ICN; Costa Mesa, CA) in methionine and cysteine free DMEM (Gibco; Grand Island, NY) for six hours. The labelled cells were detached with PBS containing 1mM EDTA, collected by centrifugation, and lysed in 1% NP-40, 0.1% SDS, 0.15 M NaCl, 0.05M Tris pH 30 8.0, 5mM EDTA, and 1mM PMSF. Nuclei were removed by centrifugation, and CD4 proteins immunoprecipitated with OKT4 antibody and anti-mouse IgG agarose (Cappel, Durham, NC). Samples were electrophoresed through 8% 35 polyacrylamide/SDS gels under non-reducing (NR) and reducing (R) conditions. Gels containing  $^{35}\text{S}$ -labelled

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samples were impregnated with En<sup>3</sup>Hance (New England Nucl ar, Boston, MA) prior to autoradiography. Facilitated expression of the transmembrane form of CD16, CD16<sub>TM</sub>, was measured by comparing its expression in CV1 5 cells singly infected with CD16<sub>TM</sub> with expression in cells coinfecte<sup>d</sup> with viruses encoding CD16<sub>TM</sub> and  $\zeta$  or  $\gamma$  chimeras. After infection and incubation for six hours or more, cells were detached from plates with PBS, 1mM EDTA and the expression of CD16TM or the chimeras was 10 measured by indirect immunofluorescence and flow cytometry.

#### Calcium Flux Assay

Jurkat subline E6 (Weiss et al., J. Immunol., 133:123-128 (1984)) cells were infected with recombinant 15 vaccinia viruses for one hour in serum free IMDM at an moi of 10 and incubated for three to nine hours in IMDM, 10% FBS. Cells were collected by centrifugation and resuspended at  $3 \times 10^6$  cells/ml in complete medium containing 1mM Indo-1 acetomethoxyester (Grynkiewicz et 20 al., J. Biol. Chem., 260:3340-3450 (1985)) (Molecular Probes) and incubated at 37°C for 45 minutes. The Indo-1 loaded cells were pelleted and resuspended at  $1 \times 10^6$ /ml in serum free IMDM and stored at room temperature in the dark. Cells were analyzed for free calcium ion by 25 simultaneous measurement of the violet and blue fluorescence emission by flow cytometry (Rabinovitch et al., J. Immunol., 137:952-961 (1986)). To initiate calcium flux, either phycoerythrin (PE)-conjugated Leu-3A (anti-CD4) (Becton Dickinson, Lincoln Park, NJ) at 1 30  $\mu\text{g}/\text{ml}$  was added to the cell suspension followed by 10 $\mu\text{g}/\text{ml}$  of unconjugated goat anti-mouse IgG at time 0 or unconjugated 3G8 (anti-CD16) monoclonal antibody was added to the cell suspension at 1  $\mu\text{g}/\text{ml}$  followed by 10  $\mu\text{g}/\text{ml}$  of PE-conjugated Fab<sub>2</sub>' goat anti-mouse IgG at time 35 0. Histograms of the violet/blue emission ratio were

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collected from the PE positive (infected) cell population, which typically represented 40-80% of all cells. The T cell antigen receptor response in uninfected cells was triggered by antibody OKT3, without crosslinking. For experiments involving CD16 chimeric receptors, samples showing baseline drift toward lower intracellular calcium (without antibody) were excluded from the analysis. Histogram data were subsequently analyzed by conversion of the binary data to ASCII using Write Hand Man (Cooper City, FL) software, followed by analysis with a collection of FORTRAN programs. The violet/blue emission ratio prior to the addition of the second antibody reagents was used to establish the normalized initial ratio, set equal to unity, and the resting threshold ratio, set so that 10% of the resting population would exceed threshold.

#### Cytolysis Assay

Human T cell line WH3, a  $CD8^+$   $CD4^-$  HLA B44 restricted cytolytic line was maintained in IMDM, 10% human serum with 100 U/ml of IL-2 and was periodically stimulated either nonspecifically with irradiated (3000 rad) HLA-unmatched peripheral blood lymphocytes and 1 $\mu$ g/ml of phytohemagglutinin, or specifically, with irradiated B44-bearing mononuclear cells. After one day of nonspecific stimulation, the PHA was diluted to 0.5  $\mu$ g/ml by addition of fresh medium, and after three days the medium was changed. Cells were grown for at least 10 days following stimulation before use in cytotoxicity assays. The cells were infected with recombinant vaccinia at a multiplicity of infection of at least 10 for one hour in serum free medium, followed by incubation in complete medium for three hours. Cells were harvested by centrifugation and resuspended at a density of  $1 \times 10^7$  cells/ml. 100 $\mu$ l were added to each well of a U-bottom 35 microtiter plate containing 100  $\mu$ l/well of complete

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medium. Cells were diluted in two-fold serial steps. Two wells for each sample did not contain lymphocytes, to allow spontaneous chromium release and total chromium uptake to be measured. The target cells, from HeLa 5 subline S3, were infected in 6.0 or 10.0 cm plates at an approximate moi of 10 for one hour in serum free medium, followed by incubation in complete medium for three hours. They were then detached from the dishes with PBS, 1mM EDTA and counted. An aliquot of  $10^6$  target cells 10 (HeLa, Raji, or RJ2.2.5 cells for the CD4 chimeric receptor experiments and 3G8 10-2 cells; Shen et al., Mol. Immunol. 26:959 (1989) for the CD16 chimeric receptor experiments) was centrifuged and resuspended in 50  $\mu$ l of sterile  $^{51}\text{Cr}$ -sodium chromate (1mCi/ml, Dupont 15 Wilmington, DE) for one hour at 37°C with intermittent mixing, then washed three times with PBS. 100  $\mu$ l of labelled cells resuspended in medium at  $10^5$  cells/ml were added to each well. Raji and RJ2.2.5 target cells were labelled in the same manner as HeLa cells. The 20 microtiter plate was spun at 750 x g for 1 minute and incubated for 4 hours at 37°C. At the end of the incubation period, the cells in each well were resuspended by gentle pipetting, a sample removed to determine the total counts incorporated, and the 25 microtiter plate spun at 750 x g for 1 minute. 100 $\mu$ l aliquots of supernatant were removed and counted in a gamma ray scintillation counter. The percent killing was corrected for the fraction of infected target cells (usually 50-90%) measured by flow cytometry. For 30 infected effector cells the effector:target ratio was corrected for the percent of cells infected (usually 20-50% for the CD4 chimeric receptor experiments and >70% for the CD16 chimeric receptor experiments).

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**In Vitro Mutagenesis of the  $\zeta$  Sequence**

To create point mutations in amino acid residues 11 and/or 15 of the  $\zeta$  sequence, synthetic oligonucleotide primers extending from the BamHI site upstream of the  $\zeta$  transmembrane domain, and converting native  $\zeta$  residue 11 from Cys to Gly (C11G) or residue 15 from Asp to Gly (D15G) or both (C11G/D15G) were prepared and used in PCR reactions to generate mutated fragments which were reinserted into the wild type CD4: $\zeta$  constructs.

10 To create  $\zeta$  deletions,  $\zeta$  cDNA sequences were amplified by PCR using synthetic oligonucleotide primers designed to create a stop codon (UAG) after residues 50, 59, or 65. The primers contained the cleavage site for the enzyme NotI indented five or six residues from the 5' 15 end, usually in a sequence of the form CGC GGG CGG CCG CTA (SEQ ID NO: 11), where the last three residues correspond to the stop anticodon. The NotI and stop anticodon sequences were followed by 18 or more residues complementary to the desired 3' end of the fragment.

20 The resulting chimeras were designated CD16: $\zeta$ Y51\*, CD16: $\zeta$ E60\* and CD16: $\zeta$ D66\* respectively. The BamHI site upstream of the transmembrane domain and the NotI site were used to generate fragments that were reintroduced into the wild type CD16: $\zeta$  construct. Monomeric  $\zeta$  25 chimeras were created by liberating the  $\zeta$  transmembrane and membrane proximal intracellular sequences by BamHI and SacI digestion of the Asp<sup>-</sup> and Cys<sup>-</sup> CD4: $\zeta$  construct described above and inserting the fragment into the CD16: $\zeta$ E60\* and CD16: $\zeta$ D66\* construct respectively.

30 CD16:7: $\zeta$ (48-65) and CD16:7 $\zeta$ (48-59) tripartite chimera construction.

To prepare the construct CD16: $\zeta$ D66\*, the  $\zeta$  cDNA sequence corresponding to the transmembrane domain and the 17 following residues of the cytoplasmic domain was 35 replaced by corresponding transmembrane and cytoplasmic

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domain obtained from the CD5 and CD7 cDNA. The CD5 and CD7 fragments were generated by a PCR reaction using forward oligonucleotides including a BamHI restriction cleavage site and corresponding to the region just 5 upstream of the transmembrane domain of CD5 and CD7 respectively and the following reverse oligonucleotides overlapping the CD5 and CD7 sequences respectively and the  $\zeta$  sequence which contained the SacI restriction cleavage site.

10 CD5: $\zeta$ : CGC GGG CTC GTT ATA GAG CTG GTT CTG GCG  
CTG CTT CTT CTG (SEQ ID NO: 12)

CD7: $\zeta$ : CGC GGG GAG CTC GTT ATA GAG CTG GTT TGC  
CGC CGA ATT CTT ATC CCG (SEQ ID NO: 13).

The CD5 and CD7 PCR products were digested with BamHI and 15 SacI and ligated to BamHI and SacI digested CD16: $\zeta$ E60\* and replacing the  $\zeta$  sequence from BamHI to SacI by the CD7 fragment. To make the constructs CD16:CD5 and CD16:CD7, CD5 and CD7 fragments were obtained by PCR using an oligonucleotide containing a NotI restriction 20 cleavage site and encoding a stop codon (UAA) after the residue Gln416 and Ala193 of CD5 and CD7 respectively. The CD5 and CD7 PCR fragment were digested with BamHI and NotI and inserted in the CD16: $\zeta$ Asp66\* construct.

25 In Vitro Mutagenesis of the N-terminal Residues within the  $\zeta$  Cytolytic Signal-Transducing Motif

Synthetic oligonucleotide primers extending from the SacI site inside the  $\zeta$  motif and converting native residue 48 from Asn to Ser (N48S), residue 50 from Leu to Ser (L50S) and residue 51 from Tyr to Phe (Y51F) were 30 synthesized and used in a PCR reaction to generate fragments that were reintroduced into the wild type CD16:7: $\zeta$ (48-65) construct.

In Vitro Mutagenesis of C-terminal Residues within the  $\zeta$  Cytolytic Signal-Transducing Motif

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Synthetic oligonucleotide primers extending from the NotI site 3' to the stop codon and converting native residue 60 from Glu to Gln (E60Q), residue 61 from Glu to Gln (E61Q), residue 62 from Tyr to Phe or Ser (Y62F or Y62S) and residue 63 from Asp to Asn (D63N) were synthesized and used in PCR to generate fragments that were subcloned into the wild type CD16:5D66\* construct from the BamHI site to the NotI site.

CD16:7:5(33-65), CD16:7:5(71-104), CD16:7:5(104-137)

#### 10 Chimera Constructions

A CD7 transmembrane fragment bearing MluI and NotI sites at the junction between the transmembrane and intracellular domains was obtained by PCR using an oligonucleotide with the following sequence: CGC GGG 15 GCG GCC ACG CGT CCT CGC CAG CAC ACA (SEQ ID NO: 14). The resulting PCR fragment was digested with BamHI and NotI and reinserted into the CD16:7:5(48-65) construct. 5 fragments encoding residues 33 to 65, 71 to 104, and 104 to 137 were obtained by PCR reaction using pairs of 20 primers containing MluI sites at the 5' end of the forward primers and stop codons followed by NotI sites at the 5' end of the reverse primers. In each case the restriction sites were indented six residues from the 5' terminus of the primer to insure restriction enzyme 25 cleavage.

5 33: CGC GGG ACG CGT TTC AGC CGT CCT CGC CAG CAC ACA (SEQ ID NO: 15);

5 71: CGC GGG ACG CGT GAC CCT GAG ATG GGG GGA AAG (SEQ ID NO: 16); and

30 5 104: CGC GGG ACG CGT ATT GGG ATG AAA GGC GAG CGC (SEQ ID NO: 17).

#### Construction of FcR $\gamma$ IIA Deletion Mutants

Carboxyl terminal FcRIIA deletion mutants were constructed by PCR in the same fashion as for the full 35 length constructs, converting the sequences encoding

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tyrosine at positions 282 and 298 into stop codons (TAA). The N-terminal deletions were generated by amplifying fragments encoding successively less of the intracellular domain by PCR, using oligonucleotides which allowed the 5 resulting fragments to be inserted between MluI and NotI restriction sites into a previously constructed expression plasmid encoding the CD16 extracellular domain fused to the CD7 transmembrane domain, the latter terminating in a MluI site at the junction between the 10 transmembrane and the intracellular domain.

OTHER EMBODIMENTS

The examples described above demonstrate that aggregation of  $\zeta$ ,  $\eta$ , or  $\gamma$  chimeras suffices to initiate the cytolytic effector cell response in T cells. The 15 known range of expression of  $\zeta$ ,  $\eta$ , and  $\gamma$ , which includes T lymphocytes, natural killer cells, basophilic granulocytes, macrophages and mast cells, suggests that conserved sequence motifs may interact with a sensory apparatus common to cells of hematopoietic origin and 20 that an important component of host defense in the immune system may be mediated by receptor aggregation events.

The potency of the cytolytic response and the absence of a response to target cells bearing MHC class II receptors demonstrates that chimeras based on  $\zeta$ ,  $\eta$ , or 25  $\gamma$  form the basis for a genetic intervention for AIDS through adoptive immunotherapy. The broad distribution of endogenous  $\zeta$  and  $\gamma$  and evidence that Fc receptors associated with  $\gamma$  mediate cytotoxicity in different cell types (Fanger et al., Immunol. Today, 10:92-99 (1989)) 30 allows a variety of cells to be considered for this purpose. For example, neutrophilic granulocytes, which have a very short lifespan ( $\approx 4h$ ) in circulation and are intensely cytolytic, are attractive target cells for expression of the chimeras. Infection of neutrophils 35 with HIV is not likely to result in virus release, and

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the abundance of these cells (the most prevalent of the leukocytes) should facilitate host defens . Anoth r attractive possibility for host cells are mature T cells, a population presently accessible to retroviral  
5 engineering (Rosenberg, S.A. Sci. Am., 262:62-69 (1990)). With the aid of recombinant IL-2, T cell populations can be expanded in culture with relative ease, and the expanded populations typically have a limited lifespan when reinfused (Rosenberg et al., N. Engl. J. Med.,  
10 323:570-578 (1990)).

Under the appropriate conditions, HIV recognition by cells expressing CD4 chimeras should also provide mitogenic stimuli, allowing the possibility that the armed cell population could respond dynamically to the  
15 viral burden. Although we have focused here on the behavior of the fusion proteins in cytolytic T lymphocytes, expression of the chimeras in helper lymphocytes might provide an HIV-mobilized source of cytokines which could counteract the collapse of the  
20 helper cell subset in AIDS. Recent description of several schemes for engineering resistance to infection at steps other than virus penetration (Friedman et al., Nature, 335:452-454 (1988); Green et al., Cell, 58:215-223 (1989); Malim et al., Cell, 58:205-214 (1989); Trono  
25 et al., Cell, 59:113-120 (1989); Buonocore et al., Nature, 345:625-628 (1990)) suggests that cells bearing CD4 chimeras could be designed to thwart virus production by expression of appropriate agents having an intracellular site of action.

30 The ability to transmit signals to T lymphocytes through autonomous chimeras also provides the ability for the regulation of retrovirally engineered lymphocytes in vivo. Crosslinking stimuli, mediated for example by specific IgM antibodies engineered to remove complement-  
35 binding domains, may allow such lymphocytes to increase

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in number *in situ*, while treatment with similar specific IgG antibodies (for example recognizing an amino acid variation engineered into the chimeric chain) could selectively deplete the engineered population.

5 Additionally, anti-CD4 IgM antibodies do not require additional crosslinking to mobilize calcium in Jurkat cells expressing CD4:5 chimeras (data not shown). The ability to regulate cell populations without recourse to repeated extracorporeal amplification may substantially 10 extend the range and efficacy of current uses proposed for genetically engineered T cells.

To create other chimeras consisting of  $\zeta$ ,  $\eta$  or  $\gamma$  intracellular sequences, cDNA or genomic sequences encoding an extracellular domain of the receptor can be 15 endowed with a restriction site introduced at a location just preceding the transmembrane domain of choice. The extracellular domain fragment terminating in the restriction site can then be joined to  $\zeta$ ,  $\eta$ , or  $\gamma$  sequences. Typical extracellular domains may be derived 20 from receptors which recognize complement, carbohydrates, viral proteins, bacteria, protozoan or metazoan parasites, or proteins induced by them. Similarly, ligands or receptors expressed by pathogens or tumor cells can be attached to  $\zeta$ ,  $\eta$ , or  $\gamma$  sequences, to direct 25 immune responses against cells bearing receptors recognizing those ligands.

While the invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications 30 and this application is intended to cover variations, uses, or adaptations of the invention and including such departures from the present disclosure as come within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth as 35 follows in the scope of the appended claims.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANTS: The General Hospital Corporation

(ii) TITLE OF INVENTION: Redirection of Cellular Immunity by Receptor Chimeras

(iii) NUMBER OF SEQUENCES: 27

(iv) CORRESPONDENCE ADDRESS:

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(v) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: 3.5" Diskette, 1.44 Mb
- (B) COMPUTER: IBM PS/2 Model 50Z or 55SX
- (C) OPERATING SYSTEM: IBM P.C. DOS (Version 3.30)
- (D) SOFTWARE: Wordperfect (Version 5.0)

(vi) CURRENT APPLICATION DATA:

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(viii) ATTORNEY/AGENT INFORMATION:

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(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1728 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

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(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTI N: SEQ ID NO:1:

ATGAACCGGG GAGTCCCTT TAGGCACTTG CTTCTGGTGC TGCAACTGGC	50
GCTCCTCCCA GCAGCCACTC AGGGAAACAA AGTGGTGCTG GGCAAAAAAG	100
GGGATACAGT GGAACGTGACC TGTACAGCTT CCCAGAAGAA GAGCATAACAA	150
TTCCACTGGA AAAACTCCAA CCAGATAAAG ATTCTGGAA ATCAGGGCTC	200
CTTCTTAACT AAAGGTCCAT CCAAGCTGAA TGATCGCGCT GACTCAAGAA	250
GAAGCCTTGT GGACCAAGGA AACCTCCCCC TGATCATCAA GAATCTTAAG	300
ATAGAAGACT CAGATACCTA CATCTGTGAA GTGGAGGACC AGAAGGAGGA	350
GGTGCAATTG CTAGTGTTCG GATTGACTGC CAACTCTGAC ACCCACCTGC	400
TTCAGGGGCA GAGCCTGACC CTGACCTTGG AGAGCCCCCC TGCTAGTAGC	450
CCCTCAGTGC AATGTAGGAG TCCAAGGGGT AAAAACATAC AGGGGGGGAA	500
GACCCCTCTCC GTGTCTCAGC TGGAGCTCCA GGATAGTGGC ACCTGGACAT	550
GCACTGTCTT GCAGAACCAAG AAGAAGGTGG AGTTCAAAAT AGACATCGTG	600
GTGCTAGCTT TCCAGAAGGC CTCCAGCATA GTCTATAAGA AAGAGGGGGAA	650
ACAGGTGGAG TTCTCTTCC CACTCGCTT TACAGTTGAA AAGCTGACGG	700
GCAGTGGCGA GCTGTGGTGG CAGGCCGAGA GGGCTTCCTC CTCCAAGTCT	750
TGGATCACCT TTGACCTGAA GAACARGGAA GTGTCTGTAA AACGGGTTAC	800
CCAGGACCT AAGCTCCAGA TGGGCAAGAA GCTCCCGCTC CACCTCACCC	850
TGCCCCAGGC CTTGCCTCAG TATGCTGGCT CTGGAAACCT CACCCCTGGCC	900
CTTGAAGCGA AAACAGGAAA GTTGCATCAG GAAGTGAACC TGGTGGTGAT	950
GAGAGCCACT CAGCTCCAGA AAAATTGAC CTGTGAGGTG TGGGGACCCCA	1000
CCTCCCCCTAA GCTGATGCTG AGCTTGAAAC TGGAGAACAA GGAGGGAAAG	1050
GTCTCGAACG GGGAGAAGCC GGTGTGGGTG CTGAACCCCTG AGGCCGGGAT	1100
GTGGCAGTGT CTGCTGAGTG ACTCGGGACA GGTCCCTGCTG GAATCCAACA	1150
TCAAGGTTCT GCCCACATGG TCCACCCCCGG TGCACCGCGGA TCCCRAACTC	1200
TGCTACTTGC TAGATGGAAT CCTCTTCATC TACGGAGTCA TCATCACAGC	1250
CCTGTACCTG AGAGCAAAAT TCAGCAGGAG TGCAGAGACT GCTGCCAAC	1300

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TGCAGGACCC	CAACCAGCTC	TACAATGAGC	TCAATCTAGG	GCGAAGAGAG	1350
GAATATGACG	TCTTGGAGAA	GAAGCGGGCT	CGGGATCCA	AGATGGGAGG	1400
CRAACAGCAG	AGGAGGGAGGA	ACCCCCAGGA	AGGCGTATAAC	AATGCACTGC	1450
AGAAAGACAA	GATGCCAGAA	GCCTACAGTG	AGATCGGCAC	AAAAGGCGAG	1500
AGGCAGGAGAG	GCAAGGGGCA	CGATGGCCTT	TACCAGGACA	GCCACTTCCA	1550
AGCAGTGCAG	TTCGGGAACA	GAAGAGAGAG	AGAAGGTTCA	GAACTCACAA	1600
GGACCCCTTGG	TTAAAGAGCC	CGCCCCAAAG	GTGAAAGCAC	CCAGCAGACT	1650
AGCCAATCCT	GTGCCAGCGT	CTTCAGCATT	CCCACTCTGT	GGAGTCCATG	1700
GCCACCCAGT	AGCAGCTCCC	AGCTCTAA			1728

## (2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1389 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

ATGAACCGGG	GACTCCCTTT	TAGGCACITG	CTTCTGGTGC	TGCAACTGGC	50
GCTCCTCCCA	GCAGCCACTC	AGGGAAACAA	AGTGGTGTG	GGCAAAAAAG	100
GGGATACAGT	GGAACGTGACC	TGTACAGCTT	CCCAGAAGAA	GAGCATACAA	150
TTCCACTGGA	AAAACCTCAA	CCAGATAAAAG	ATTCTGGAA	ATCAGGGCTC	200
CTTCTTAACT	AAAGGTCCAT	CCAAGCTGAA	TGATCGCGCT	GACTCAAGAA	250
GAAGCCTTTG	GGACCAAGGA	AACTTCCCCC	TGATCATCAA	GAATCTTAAG	300
ATAGAAGACT	CAGATACTTA	CATCTGTGAA	GTGGAGGACC	AGAAGGAGGA	350
GGTGCAATTG	CTAGTGTTCG	GATTGACTGC	CAACTCTGAC	ACCCACCTGC	400
TTCAGGGGCA	GAGCCTGACC	CTGACCTTGG	AGAGCCCCCC	TGGTAGTAGC	450
CCCTCAGTGC	AATGTAGGAG	TCCAAGGGGT	AAAAACATAC	AGGGGGGGAA	500
GACCCCTCTCC	GTGTCTCAGC	TGGAGCTCCA	GGATAGTGGC	ACCTGGACAT	550
GCACTGTCTT	GCAGAACCGAG	AAGAAGGTGG	AGTTCAAAAT	AGACATCGTG	600

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GTCCTAGCTT TCCAGAAGGC CTCCAGCATA GTCTATAAGA AAGAGGGGGA	650
ACAGGTGGAG TTCTCCCTCC CACTCGCCTT TACAGTTGAA AAGCTGACGG	700
GCAGTGGCGA GCTGTGGTGG CAGGGGGAGA GGGCTTCCTC CTCCAAGTCT	750
TGGATCACCT TTGACCTGAA GAACAAGGAA GTGTCTGTAA AACGGGTTAC	800
CCAGGACCT AAGCTCCAGA TGGGCAAGAA GCTCCGCTC CACCTCACCC	850
TGCCCCAGGC CTTGCCTCAG TATGCTGGCT CTGGAAACCT CACCCCTGGCC	900
CITGAAGCGA AAACAGGAAA GTTGCATCAG GAAGTGAACC TGGTGGGTGAT	950
GAGAGCCACT CAGCTCCAGA AAAATTGAC CTGTGAGGTG TGGGGACCCA	1000
CCTCCCCCTAA GCTGATGCTG AGCTTGAAAC TGGAGAACAA GGAGGCAAAG	1050
GTCTCGAAGC GGGAGAAGCC GGTGTGGGTG CTGAACCCCTG AGGCAGGGAT	1100
GTGGCAGTGT CTGCTGAGTG ACTCGGGACA GGTCTGCTG GAATCCAACA	1150
TCAAGGTCT TCCCACATGG TCCACCCCCG TGCACCGGGA TCCGCAGCTC	1200
TGCTATATCC TGGATGCCAT CCTGTTTTTG TATGGTATTG TCCTTACCCCT	1250
GCTCTACTGT CGACTCAAGA TCCAGGTCCG AAAGGCAGAC ATAGCCAGCC	1300
GTGAGAAATC AGATGCTGTC TACACGGGCC TGAACACCCG GAACCAGGAG	1350
ACATATGAGA CTCTGAAACCA TGAGAAACCA CCCCCAATAG	1389

## (2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1599 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATGAACCGGG GAGTCCCTTT TAGGCACCTG CTTCTGGTGC TGCAACTGGC	50
GCTCCTCCCA CGAGCCACTC AGGGAAACAA AGTGGTGCTG GGCAAAAAAG	100
GGGATACAGT GGAACGTGACC TGTACAGCTT CCCAGAAGAA GAGCATAACAA	150
TTCCACTGGA AAAACTCCAA CCAGATAAAG ATTCTGGGAA ATCAGGGCTC	200
CITCTTAACT AAAGGTCCAT CCAAGCTGAA TGATCGCGCT GACTCAAGAA	250

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GAAGCCTTTC	GGACCAAGGA	AACTTCCCCC	TGATCATCAA	GAATCTTAAG	300
ATAGAAGACT	CAGATACTTA	CATCTGTGAA	GTGGAGGACC	AGAAGGAGGA	350
GCTGCAATTG	CTAGTGTTCG	GATTGACTGC	CAACTCTGAC	ACCCACCTGC	400
TTCAGGGGCA	GAGCCTGACC	CTGACCTTGG	AGAGCCCCCC	TGGTAGTAGC	450
CCCTCAGTGC	AAATGAGGAG	TCCAAAGGGT	AAAAACATAC	ACGGGGGGAA	500
GACCCCTCTCC	GTGTCTCAGC	TGGACCTCCA	GGATAGTGGC	ACCTGGACAT	550
GCACGTGCTT	GCAGAACCAAG	AAAAGGTGG	AGITCAAAAT	AGACATCGTG	600
GTGCTAGCTT	TCCAGAAGGC	CTCCAGCATA	GTCTATAAGA	AAAGAGGGGA	650
ACAGGTGGAG	TTCTCCTTCC	CACTCGCCTT	TACAGTTGAA	AAGCTGACGG	700
GCAGTGGCGA	GCTGTGGTGG	CAGGCCGAGA	GGGCTTCCTC	CTCCAAGTCT	750
TGGATCACCT	TTGACCTGAA	GAACAAGGAA	GTGCTGTAA	AACGGGTTAC	800
CCAGGACCC	AAGCTCCAGA	TGGGCCAGAA	GCTCCCGCTC	CACCTCACCC	850
TGCCCCAGGC	CTTGCCTCAG	TATGCTGGCT	CTGGAAACCT	CACCTGGCC	900
CTTGAAGGCA	AAACAGGAAA	GTTGCATCAG	GAAGTGAACC	TGGTGGTGAT	950
GAGAGCCACT	CAGCTCCAGA	AAAATTTGAC	CTGTGAGGTG	TGGGGACCCA	1000
CCTCCCCCTAA	GCTGATGCTG	AGCTTGAAAC	TGGAGAACAA	GGAGGCAAAG	1050
GTCTCGAAC	GGGAGAAGCC	GGTGTGGTG	CTGAACCTG	AGGGGGGGAT	1100
GTGGCAGTGT	CTGCTGAGTG	ACTCGGGACA	GGTCCTGCTG	GAATCCAACA	1150
TCAAGGTTCT	GCCCACATGG	TCCACCCCGG	TGCACGCGGA	TCCCAAACTC	1200
TGCTACCTGC	TGGATGGAAT	CCTCTTCATC	TATGGTGTCA	TTCTCACTGC	1250
CTTGTTCCTG	AGAGTGAAGT	TCAGCAGGAG	CGCAGAGCCC	CCCGCGTACC	1300
AGCAGGGCCA	GAACCAAGCTC	TATAACGAGC	TCAATCTAGG	ACGAAGAGAG	1350
GAGTACGATG	TTTTGGACAA	GAGACGTGGC	CGGGACCCCTG	AGATGGGGGG	1400
AAAGCCGAGA	AGGAAGAAC	CTCAGGAAGG	CCTGTACAAT	GAACCTGCAGA	1450
AAGATAAGAT	GGCGGAGGCC	TACAGTGAGA	TTGGGATGAA	AGGCGAGGCC	1500
CGGAGGGGCA	AGGGGCACGA	TGGCCTTAC	CAGGGTCTCA	GTACAGCCAC	1550
CAAGGACACC	TACGACGCC	TTCACATGCA	GGCCCTGCC	CCTCGCTAA	1599

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 575 amino acids  
 (B) TYPE: amin acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Asn Arg Gly Val Pro Phe Arg His Leu Leu Leu Val Leu Gln Leu  
 1 5 10 15  
 Ala Leu Leu Pro Ala Ala Thr Gln Gly Asn Lys Val Val Leu Gly Lys  
 20 25 30  
 Lys Gly Asp Thr Val Glu Leu Thr Cys Thr Ala Ser Gln Lys Lys Ser  
 35 40 45  
 Ile Gln Phe His Trp Lys Asn Ser Asn Gln Ile Lys Ile Leu Gly Asn  
 50 55 60  
 Gln Gln Ser Phe Leu Thr Lys Gly Pro Ser Lys Leu Asn Asp Arg Ala  
 65 70 75 80  
 Asp Ser Arg Arg Ser Leu Trp Asp Gln Gly Asn Phe Pro Leu Ile Ile  
 85 90 95  
 Lys Asn Leu Lys Ile Glu Asp Ser Asp Thr Tyr Ile Cys Glu Val Glu  
 100 105 110  
 Asp Gln Lys Glu Glu Val Gln Leu Leu Val Phe Gly Leu Thr Ala Asn  
 115 120 125  
 Ser Asp Thr His Leu Leu Gln Gly Gln Ser Leu Thr Leu Thr Leu Glu  
 130 135 140  
 Ser Pro Pro Gly Ser Ser Pro Ser Val Gln Cys Arg Ser Pro Arg Gly  
 145 150 155 160  
 Lys Asn Ile Gln Gly Gly Lys Thr Leu Ser Val Ser Gln Leu Glu Leu  
 165 170 175  
 Gln Asp Ser Gly Thr Trp Thr Cys Thr Val Leu Gln Asn Gln Lys Lys  
 180 185 190  
 Val Glu Phe Lys Ile Asp Ile Val Val Leu Ala Phe Gln Lys Ala Ser  
 195 200 205  
 Ser Ile Val Tyr Lys Lys Glu Gly Glu Gln Val Glu Phe Ser Phe Pro  
 210 215 220  
 Leu Ala Phe Thr Val Glu Lys Leu Thr Gly Ser Gly Glu Leu Trp Trp  
 225 230 235 240  
 Gln Ala Glu Arg Ala Ser Ser Ser Lys Ser Trp Ile Thr Phe Asp Leu  
 245 250 255  
 Lys Asn Lys Glu Val Ser Val Lys Arg Val Thr Gln Asp Pro Lys Leu  
 260 265 270  
 Gln Met Gly Lys Lys Leu Pro Leu His Leu Thr Leu Pro Gln Ala Leu  
 275 280 285  
 Pro Gln Tyr Ala Gly Ser Gly Asn Leu Thr Leu Ala Leu Glu Ala Lys  
 290 295 300  
 Thr Gly Lys Leu His Gln Glu Val Asn Leu Val Val Met Arg Ala Thr  
 305 310 315 320  
 Gln Leu Gln Lys Asn Leu Thr Cys Glu Val Trp Gly Pro Thr Ser Pro  
 325 330 335  
 Lys Leu Met Leu Ser Leu Lys Leu Glu Asn Lys Glu Ala Lys Val Ser  
 340 345 350  
 Lys Arg Glu Lys Pro Val Trp Val Leu Asn Pro Glu Ala Ily Met Trp  
 355 360 365

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Gln Cys Leu Leu Ser Asp Ser Gly Gln Val Leu Leu Glu Ser Asn Ile  
 370 375 380  
 Lys Val Leu Pro Thr Trp Ser Thr Pro Val His Ala Asp Pro Lys Leu  
 385 390 395 400  
 Cys Tyr Leu Leu Asp Gly Ile Leu Phe Ile Tyr Gly Val Ile Ile Thr  
 405 410 415  
 Ala Leu Tyr Leu Arg Ala Lys Phe Ser Arg Ser Ala Glu Thr Ala Ala  
 420 425 430  
 Asn Leu Gln Asp Pro Asn Gln Leu Tyr Asn Glu Leu Asn Leu Gly Arg  
 435 440 445  
 Arg Glu Glu Tyr Asp Val Leu Glu Lys Lys Arg Ala Arg Asp Pro Glu  
 450 455 460  
 Met Gly Gly Lys Gln Gln Arg Arg Arg Asn Pro Gln Glu Gly Val Tyr  
 465 470 475 480  
 Asn Ala Leu Gln Lys Asp Lys Met Pro Glu Ala Tyr Ser Glu Ile Gly  
 485 490 495  
 Thr Lys Gly Glu Arg Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln  
 500 505 510  
 Asp Ser His Phe Gln Ala Val Gln Phe Gly Asn Arg Arg Glu Arg Glu  
 515 520 525  
 Gly Ser Glu Leu Thr Arg Thr Leu Gly Leu Arg Ala Arg Pro Lys Gly  
 530 535 540  
 Glu Ser Thr Gln Gln Ser Ser Gln Ser Cys Ala Ser Val Phe Ser Ile  
 555 550 565 560  
 Pro Thr Leu Trp Ser Pro Trp Pro Pro Ser Ser Ser Ser Gln Leu  
 565 570 575

## (2) INFORMATION FOR SEQ ID NO:5:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 462 amino acids
- (B) TYPE: amino acid
- (C) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Asn Arg Gly Val Pro Phe Arg His Leu Leu Leu Val Leu Gln Leu  
 1 5 10 15  
 Ala Leu Leu Pro Ala Ala Thr Gln Gly Asn Lys Val Val Leu Gly Lys  
 20 25 30  
 Lys Gly Asp Thr Val Glu Leu Thr Cys Thr Ala Ser Gln Lys Lys Ser  
 35 40 45  
 Ile Gln Phe His Trp Lys Asn Ser Asn Gln Ile Lys Ile Leu Gly Asn  
 50 55 60  
 Gln Gly Ser Phe Leu Thr Lys Gly Pro Ser Lys Leu Asn Asp Arg Ala  
 65 70 75 80  
 Asp Ser Arg Arg Ser Leu Trp Asp Gln Gly Asn Phe Pro Leu Ile Ile  
 85 90 95  
 Lys Asn Leu Lys Ile Glu Asp Ser Asp Thr Tyr Ile Cys Glu Val Glu  
 100 105 110  
 Asp Gln Lys Glu Glu Val Gln Leu Leu Val Phe Gly Leu Thr Ala Asn  
 115 120 125  
 Ser Asp Thr His Leu Leu Gln Gly Gln Ser Leu Thr Leu Thr Leu Glu

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130	135	140
Ser Pro Pro Gly Ser Ser Pr	Ser Val Gln Cys Arg Ser Pro Arg Gly	
145	150	155
Lys Asn Ile Gln Gly Gly Lys Thr Leu Ser Val Gln Leu Glu Leu		160
165	170	175
Gln Asp Ser Gly Thr Trp Thr Cys Thr Val Leu Gln Asn Gln Lys Lys		
180	185	190
Val Glu Phe Lys Ile Asp Ile Val Val Leu Ala Phe Gln Lys Ala Ser		
195	200	205
Ser Ile Val Tyr Lys Lys Glu Gly Glu Gln Val Glu Phe Ser Phe Pro		
210	215	220
Leu Ala Phe Thr Val Glu Lys Leu Thr Gly Ser Gly Glu Leu Trp Trp		
225	230	235
Gln Ala Glu Arg Ala Ser Ser Ser Lys Ser Trp Ile Thr Phe Asp Leu		
245	250	255
Lys Asn Lys Glu Val Ser Val Lys Arg Val Thr Gln Asp Pro Lys Leu		
260	265	270
Gln Met Gly Lys Lys Leu Pro Leu His Leu Thr Leu Pro Gln Ala Leu		
275	280	285
Pro Gln Tyr Ala Gly Ser Gly Asn Leu Thr Leu Ala Leu Glu Ala Lys		
290	295	300
Thr Gly Lys Leu His Gln Glu Val Asn Leu Val Val Met Arg Ala Thr		
305	310	315
Gln Leu Gln Lys Asn Leu Thr Cys Glu Val Trp Gly Pro Thr Ser Pro		
325	330	335
Lys Leu Met Leu Ser Leu Lys Leu Glu Asn Lys Glu Ala Lys Val Ser		
340	345	350
Lys Arg Glu Lys Pro Val Trp Val Leu Asn Pro Glu Ala Gly Met Trp		
355	360	365
Gln Cys Leu Leu Ser Asp Ser Gly Gln Val Leu Leu Glu Ser Asn Ile		
370	375	380
Lys Val Leu Pro Thr Trp Ser Thr Pro Val His Ala Asp Pro Gln Leu		
385	390	395
Cys Tyr Ile Leu Asp Ala Ile Leu Phe Leu Tyr Gly Ile Val Leu Thr		
405	410	415
Leu Leu Tyr Cys Arg Leu Lys Ile Gln Val Arg Lys Ala Asp Ile Ala		
420	425	430
Ser Arg Glu Lys Ser Asp Ala Val Tyr Thr Gly Leu Asn Thr Arg Asn		
435	440	445
Gln Glu Thr Tyr Glu Thr Leu Lys His Glu Lys Pro Pro Gln		
450	455	460 462

## (2) INFORMATION FOR SEQ ID NO:6:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 532 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Asn Arg Gly Val Pro Phe Arg His Leu Leu Leu Val Leu Gln Leu		
1	5	10
		15

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Ala Leu Leu Pro Ala Ala Thr Gln Gly Asn Lys Val Val Leu Gly Lys  
 20 25 30  
 Lys ly Asp Thr Val Glu Leu Thr Cys Thr Ala Ser Gln Lys Lys Ser  
 35 40 45  
 Ile Gln Phe His Trp Lys Asn Ser Asn Gln Ile Lys Ile Leu Gly Asn  
 50 55 60  
 Gln Gly Ser Phe Leu Thr Lys Gly Pro Ser Lys Leu Asn Asp Arg Ala  
 65 70 75 80  
 Asp Ser Arg Arg Ser Leu Trp Asp Gln Gly Asn Phe Pro Leu Ile Ile  
 85 90 95  
 Lys Asn Leu Lys Ile Glu Asp Ser Asp Thr Tyr Ile Cys Glu Val Glu  
 100 105 110  
 Asp Gln Lys Glu Glu Val Gln Leu Leu Val Phe Gly Leu Thr Ala Asn  
 115 120 125  
 Ser Asp Thr His Leu Leu Gln Gly Gln Ser Leu Thr Leu Thr Leu Glu  
 130 135 140  
 Ser Pro Pro Gly Ser Ser Pro Ser Val Gln Cys Arg Ser Pro Arg Gly  
 145 150 155 160  
 Lys Asn Ile Gln Gly Lys Thr Leu Ser Val Ser Gln Leu Glu Leu  
 165 170 175  
 Gln Asp Ser Gly Thr Trp Thr Cys Thr Val Leu Gln Asn Gln Lys Lys  
 180 185 190  
 Val Glu Phe Lys Ile Asp Ile Val Val Leu Ala Phe Gln Lys Ala Ser  
 195 200 205  
 Ser Ile Val Tyr Lys Lys Glu Gly Glu Gln Val Glu Phe Ser Phe Pro  
 210 215 220  
 Leu Ala Phe Thr Val Glu Lys Leu Thr Gly Ser Gly Glu Leu Trp Trp  
 225 230 235 240  
 Gln Ala Glu Arg Ala Ser Ser Ser Lys Ser Trp Ile Thr Phe Asp Leu  
 245 250 255  
 Lys Asn Lys Glu Val Ser Val Lys Arg Val Thr Gln Asp Pro Lys Leu  
 260 265 270  
 Gln Met Gly Lys Lys Leu Pro Leu His Leu Thr Leu Pro Gln Ala Leu  
 275 280 285  
 Pro Gln Tyr Ala Gly Ser Gly Asn Leu Thr Leu Ala Leu Glu Ala Lys  
 290 295 300  
 Thr Gly Lys Leu His Gln Glu Val Asn Leu Val Val Met Arg Ala Thr  
 305 310 315 320  
 Gln Leu Gln Lys Asn Leu Thr Cys Glu Val Trp Gly Pro Thr Ser Pro  
 325 330 335  
 Lys Leu Met Leu Ser Leu Lys Leu Glu Asn Lys Glu Ala Lys Val Ser  
 340 345 350  
 Lys Arg Glu Lys Pro Val Trp Val Leu Asn Pro Glu Ala Gly Met Trp  
 355 360 365  
 Gln Cys Leu Leu Ser Asp Ser Gly Gln Val Leu Leu Glu Ser Asn Ile  
 370 375 380  
 Lys Val Leu Pro Thr Trp Ser Thr Pro Val His Ala Asp Pro Lys Leu  
 385 390 395 400  
 Cys Tyr Leu Leu Asp Gly Ile Leu Phe Ile Tyr Gly Val Ile Leu Thr  
 405 410 415  
 Ala Leu Phe Leu Arg Val Lys Phe Ser Arg Ser Ala Glu Pro Pro Ala  
 420 425 430  
 Tyr Gln Gln Gly Gln Asn Gln Leu Tyr Asn Glu Leu Asn Leu Gly Arg  
 435 440 445  
 Arg Glu Glu Tyr Asp Val Leu Asp Lys Arg Arg Gly Arg Asp Pro Glu  
 450 455 460

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Met Gly Gly Lys Pro Arg Arg Lys Asn Pro Gln Iu Gly Leu Tyr Asn  
465 470 475 480  
Glu Leu Gln Lys Asp Lys Met Ala Glu Ala Tyr Ser Iu Ile Gly Met  
485 490 495  
Lys Iu Glu Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln Gly  
500 505 510  
Leu Ser Thr Ala Thr Lys Asp Thr Tyr Asp Ala Leu His Met Gln Ala  
515 520 525  
Leu Pro Pro Arg  
530

**(2) INFORMATION FOR SEQ ID NO:7:**

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 33 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CGGGGGGTGA CCGTGCCCTC CAGCAGCTTG GGC

33

**(2) INFORMATION FOR SEQ ID NO:8:**

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 50 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

CGCGGGGATC CGTCGTCCAG AGCCCGTCCA GCTCCCCGTC CTGGGCCTCA

50

**(2) INFORMATION FOR SEQ ID NO:9:**

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 33 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CGCGGGCGGC CGCGACGCCG GCCAAGACAG CAC

33

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CGCGTTGACG AGCAGCCAGT TGGGCAGCAG CAG

33

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

CGCGGGCGGC CGCTA

15

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 42 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CGCGGGCTCG TTATAGAGCT GGTTCTGGCG CTGCTTCITC TG

42

(2) INFORMATION FOR SEQ ID NO:13:

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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 48 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CGCGGGGAGC TCGTTATAGA GCTGGTTTGC CGCCGAATTC TTATCCCG

48

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

CGCGGGGCGG CCACGCGTCC TCGCCAGCAC ACA

33

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

CGCGGGACGC GTTTCAGCCG TCCTCGCCAG CACACA

36

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

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(xi) SEQUENCE DESCRIPTION: SEQ ID N :16:

CGCGGGACGC GTGACCCCTGA GATGGGGGGA AA

33

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

CGCGGGACGC GTATTGGGAT GAAAGGGCAG CGC

33

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CCCGGATCCCC AGCATGGGCA GCTCTT

26

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 42 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CGCGGGGGCGG CCCCTTTAGT TATTACTGTT GACATGGTCG TT

42

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

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- (A) LENGTH: 30 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

GGGGGGGGAT CCCACTGTCC AAGCTCCAG

30

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 32 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

GGGGGGGGCGG CCCCTTAAAT ACGGTTCTGG TC

32

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 31 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

TCAGAAAGAG ACAACCTGAA GAAACCAACA A

31

(2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 31 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: nucleic acid

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(xi) SEQUENCE DESCRIPTI N: SEQ ID N :23:

TTGTTGGTTT CCTCAGGTG TGTCTTTCTG A

31

**(2) INFORMATION FOR SEQ ID NO:24:**

**(1) SEQUENCE CHARACTERISTICS:**

(A) LENGTH: 171 amino acids

(B) TYPE: amino acid

• 11 •

Met	Glu	His	Ser	Thr	Phe	Leu	Ser	Gly	Leu	Val	Leu	Ala	Thr	Leu	Leu	
					5					10				15		
Ser	Gln	Val	Val	Ser	Pro	Phe	Lys	Ile	Pro	Ile	Glu	Glu	Leu	Glu	Asp	Arg
					20					25				30		
Val	Phe	Val	Asn	Cys	Asn	Thr	Ser	Ile	Thr	Trp	Val	Glu	Gly	Thr	Val	
					35					40				45		
Gly	Thr	Leu	Leu	Ser	Asp	Ile	Thr	Arg	Leu	Asp	Leu	Gly	Lys	Arg	Ile	
					50					55				60		
Leu	Asp	Pro	Arg	Gly	Ile	Tyr	Arg	Cys	Asn	Gly	Thr	Asp	Ile	Tyr	Lys	
					65					70				75		80
Asp	Lys	Glu	Ser	Thr	Val	Gln	Val	His	Tyr	Arg	Met	Cys	Gln	Ser	Cys	
					85					90				95		
Val	Glu	Leu	Asp	Pro	Ala	Thr	Val	Ala	Gly	Ile	Ile	Val	Thr	Asp	Val	
					100					105				110		
Ala	Ile	Thr	Leu	Leu	Leu	Ala	Leu	Gly	Val	Phe	Cys	Phe	Ala	Gly	His	
					115					120				125		
Glu	Thr	Gly	Arg	Leu	Ser	Gly	Ala	Ala	Asp	Thr	Gln	Ala	Leu	Leu	Arg	
					130					135				140		
Asn	Asp	Gln	Val	Tyr	Gln	Pro	Leu	Arg	Asp	Arg	Asp	Asp	Ala	Gln	Tyr	
					145					150				155		160
Ser	His	Leu	Gly	Gly	Asn	Trp	Ala	Arg	Asn	Lys						
					165					170						

**(2) INFORMATION FOR SEQ ID NO:25:**

**(i) SEQUENCE CHARACTERISTICS:**

(A) LENGTH: 182 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: amino acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Met Glu Gln Gly Lys Gly Leu Ala Val Leu Ile Leu Ala Ile Ile Leu  
5 10 15

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Leu Gln Gly Thr Leu Ala Gln Ser Ile Lys Gly Asn His Leu Val Lys  
 20 25 30  
 Val Tyr Asp Tyr Gln Glu Asp Gly Ser Val Leu Leu Thr Cys Asp Ala  
 35 40 45  
 Glu Ala Lys Asn Ile Thr Trp Phe Lys Asp Gly Lys Met Ile Gly Phe  
 50 55 60  
 Leu Thr Glu Asp Lys Lys Lys Trp Asn Leu Gly Ser Asn Ala Lys Asp  
 65 70 75 80  
 Pro Arg Gly Met Tyr Gln Cys Lys Gly Ser Gln Asn Lys Ser Lys Pro  
 85 90 95  
 Leu Gln Val Tyr Tyr Arg Met Cys Gln Asn Cys Ile Glu Leu Asn Ala  
 100 105 110  
 Ala Thr Ile Ser Gly Phe Leu Phe Ala Glu Ile Val Ser Ile Phe Val  
 115 120 125  
 Leu Ala Val Gly Val Tyr Phe Ile Ala Gly Gln Asp Gly Val Arg Gln  
 130 135 140  
 Ser Arg Ala Ser Asp Lys Gln Thr Leu Leu Pro Asn Asp Gln Leu Tyr  
 145 150 155 160  
 Gln Pro Leu Lys Asp Arg Glu Asp Asp Gln Tyr Ser His Leu Gln Gly  
 165 170 175  
 Asn Gln Leu Arg Arg Asn  
 180

## (2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 220 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: amino acids

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met Pro Gly Gly Leu Glu Ala Leu Arg Ala Leu Pro Leu Leu Phe  
 5 10 15  
 Leu Ser Tyr Ala Cys Leu Gly Pro Gly Cys Gln Ala Leu Arg Val Glu  
 20 25 30  
 Gly Gly Pro Pro Ser Leu Thr Val Asn Leu Gly Glu Glu Ala Arg Leu  
 35 40 45  
 Thr Cys Glu Asn Asn Gly Arg Asn Pro Asn Ile Thr Trp Trp Phe Ser  
 50 55 60  
 Leu Gln Ser Asn Ile Thr Trp Pro Pro Val Pro Leu Gly Pro Gly Gln  
 65 70 75 80  
 Gly Thr Thr Gly Gln Leu Phe Phe Pro Glu Val Asn Lys Asn Thr Gly  
 85 90 95  
 Ala Cys Thr Gly Cys Gln Val Ile Glu Asn Asn Ile Leu Lys Arg Ser  
 100 105 110  
 Cys Gly Thr Tyr Leu Arg Val Arg Asn Pro Val Pro Arg Pro Phe Leu  
 115 120 125  
 Asp Met Gly Glu Gly Thr Lys Asn Arg Ile Ile Thr Ala Glu Gly Ile  
 130 135 140  
 Ile Leu Leu Phe Cys Ala Val Val Pro Gly Thr Leu Leu Leu Phe Arg  
 145 150 155 160

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Lys	Arg	Trp	Gln	Asn	Glu	Lys	Phe	Gly	Val	Asp	Met	Pro	Asp	Asp	Tyr
					165				170						175
lu	Asp	Glu	Asn	Leu	Tyr	Glu	Gly	Leu	Asn	Leu	Asp	Asp	Cys	Ser	Met
					180				185						190
Tyr	Glu	Asp	Il	Ser	Arg	Gly	Leu	In	Gly	Thr	Tyr	In	Asp	Val	Gly
					195			200							205
Asn	Leu	His	Ile	Gly	Asp	Ala	Gln	Leu	Glu	Lys	Pro				
					210			215			220				

**(2) INFORMATION FOR SEQ ID NO:27:**

### **(i) SEQUENCE CHARACTERISTICS:**

- (A) LENGTH: 228 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: amino acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

Met Ala Thr Leu Val Leu Ser Ser Met Pro Cys His Trp Leu Leu Phe			
5	10	15	
Leu Leu Leu Leu Phe Ser Gly Glu Pro Val Pro Ala Met Thr Ser Ser			
20	25	30	
Asp Leu Pro Leu Asn Phe Gln Gly Ser Pro Cys Ser Gln Ile Trp Gln			
35	40	45	
His Pro Arg Phe Ala Ala Lys Lys Arg Ser Ser Met Val Lys Phe His			
50	55	60	
Cys Tyr Thr Asn His Ser Gly Ala Leu Thr Trp Phe Arg Lys Arg Gly			
65	70	75	80
Ser Gln Gln Pro Gln Glu Leu Val Ser Glu Glu Gly Arg Ile Val Gln			
85	90	95	
Thr Gln Asn Gly Ser Val Tyr Thr Leu Thr Ile Gln Asn Ile Gln Tyr			
100	105	110	
Glu Asp Asn Gly Ile Tyr Phe Cys Lys Gln Lys Cys Asp Ser Ala Asn			
115	120	125	
His Asn Val Thr Asp Ser Cys Gly Thr Glu Leu Leu Val Leu Gly Phe			
130	135	140	
Ser Thr Leu Asp Gln Leu Lys Arg Arg Asn Thr Leu Lys Asp Gly Ile			
145	150	155	160
Ile Leu Ile Gln Thr Leu Leu Ile Ile Leu Phe Ile Ile Val Pro Ile			
165	170	175	
Phe Leu Leu Leu Asp Lys Asp Asp Gly Lys Ala Gly Met Glu Glu Asp			
180	185	190	
His Thr Tyr Glu Gly Leu Asn Ile Asp Gln Thr Ala Thr Tyr Glu Asp			
195	200	205	
Ile Val Thr Leu Arg Thr Gly Glu Val Lys Trp Ser Val Gly Glu His			
210	215	220	
Pro Gly Gln Glu			
225			

claims

1        1. A method of directing a cellular response to an  
2        infective agent, to a cell infected with said agent, to a  
3        tumor or cancerous cell, or to an autoimmune-generated cell in  
4        a mammal, said method comprising administering to said mammal  
5        an effective amount of therapeutic cells, said cells being  
6        capable of specifically recognizing and destroying said agent  
7        or said cell.

1        2. The method of claim 1, wherein said cellular response  
2        is MHC-independent.

1        3. The method of claim 1, wherein said therapeutic cells  
2        express a membrane-bound, proteinaceous chimeric receptor,  
3        comprising (a) an extracellular portion which is capable of  
4        specifically recognizing and binding said agent or said cell,  
5        and (b) an intracellular or transmembrane portion which is  
6        capable of signalling said therapeutic cell to destroy a  
7        receptor-bound agent or a receptor-bound cell.

1        4. The method of claim 3, wherein said intracellular  
2        portion or said transmembrane portion is a signal-transducing  
3        portion of a T cell receptor protein, an Fc receptor protein,  
4        or a B cell receptor protein, or a functional derivative  
5        thereof.

1        5. The method of claim 4, wherein said receptor protein  
2        is  $\zeta$ ,  $\eta$ ,  $\gamma$ , CD3 delta, T3 gamma, mb1, or B29.

1        6. The method of claim 5, wherein said chimeric receptor  
2        comprises:

- 3        (a) amino acids 421-532 of SEQ ID NO: 6;
- 4        (b) amino acids 423-455, 438-455, 461-494 or 494-528 of  
5        SEQ ID NO:6;
- 6        (c) amino acids 400-420 of SEQ ID NO:6;
- 7        (d) amino acids 421-575 of SEQ ID NO: 4;

8                    ( ) amino acids 423-455, 438-455, 461-494 or 494-528 of  
9                    SEQ ID NO:4;  
10                  (f) amino acids 400-420 of SEQ ID NO: 4;  
11                  (g) amino acids 421-462 of SEQ ID NO: 5;  
12                  (h) amino acids 402-419 of SEQ ID NO:5;  
13                  (i) amino acids Tyr282-Tyr298 inclusive of Fig. 15A;  
14                  (j) amino acids 132-171 of Fig. 16 (SEQ ID NO: 24);  
15                  (k) amino acids 140-182 of Fig. 17 (SEQ ID NO: 25);  
16                  (l) amino acids 162-220 of Fig. 18 (SEQ ID NO: 26);  
17                  (m) amino acids 183-228 of Fig. 19 (SEQ ID NO: 27).

1                  7. The method of claim 1, wherein said therapeutic cells  
2                  are selected from the group consisting of:

3                  (a) T lymphocytes;  
4                  (b) cytotoxic T lymphocytes;  
5                  (c) natural killer cells;  
6                  (d) neutrophils;  
7                  (e) granulocytes;  
8                  (f) macrophages;  
9                  (g) mast cells;  
10                 (h) HeLa cells; and  
11                 (i) embryonic stem cells (ES).

1                  8. The method of claim 1, wherein said infective agent  
2                  is an immunodeficiency virus.

1                  9. The method of claim 8, wherein said extracellular  
2                  portion comprises an HIV envelope-binding portion of CD4, or a  
3                  functional HIV envelope-binding derivative thereof.

1                  10. The method of claim 9, wherein said cytotoxic T  
2                  lymphocyte is a CD8<sup>+</sup> cytotoxic T lymphocytes.

1                  11. A cell which expresses a proteinaceous membrane-  
2                  bound chimeric receptor, said receptor comprising (a) an  
3                  extracellular portion which is capable of specifically  
4                  recognizing and binding an infective agent, a cell infected

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5 with an infective agent, a tumor or cancerous cell, or an  
6 autoimmune-generated cell, and (b) an intracellular portion or  
7 transmembrane portion derived from a T cell receptor, an Fc  
8 receptor, or a B cell receptor which is capable of signalling  
9 said cell to destroy a receptor-bound agent or receptor-bound  
10 cell.

1 12. The cell of claim 11, wherein said binding is MHC-  
2 independent.

1 13. The cell of claim 11, wherein said receptor protein  
2 is  $\zeta$ ,  $\eta$ ,  $\gamma$ , CD3 delta, T3 gamma, mbl, or B29.

1 14. The cell of claim 13, wherein said chimeric receptor  
2 comprises amino acids:.

3 (a) amino acids 421-532 of SEQ ID NO: 6;  
4 (b) amino acids 423-455, 438-455, 461-494 or 494-528 of  
5 SEQ ID NO:6;  
6 (c) amino acids 400-420 of SEQ ID NO:6;  
7 (d) amino acids 421-575 of SEQ ID NO: 4;  
8 (e) amino acids 423-455, 438-455, 461-494 or 494-528 of  
9 SEQ ID NO:4;  
10 (f) amino acids 400-420 of SEQ ID NO: 4;  
11 (g) amino acids 421-462 of SEQ ID NO: 5;  
12 (h) amino acids 402-419 of SEQ ID NO:5;  
13 (i) amino acids Tyr282-Tyr298 inclusive of Fig. 15A;  
14 (j) amino acids 132-171 of Fig. 16 (SEQ ID NO: 24);  
15 (k) amino acids 140-182 of Fig. 17 (SEQ ID NO: 25);  
16 (l) amino acids 162-220 of Fig. 18 (SEQ ID NO: 26);  
17 (m) amino acids 183-228 of Fig. 19 (SEQ ID NO: 27).

1 15. The cell of claim 11, wherein said therapeutic cells  
2 are selected from the group consisting of:

3 (a) T lymphocytes;  
4 (b) cytotoxic T lymphocytes;  
5 (c) natural killer cells;  
6 (d) neutrophils;

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7                    ( ) granulocytes;  
8                    (f) macrophages;  
9                    (g) mast cells;  
10                   (h) HeLa cells; and  
11                   (i) embryonic stem cells (ES).

1                    16. The cell of claim 11, wherein said extracellular  
2                    portion comprises an HIV envelope-binding portion of CD4, or a  
3                    functional HIV envelope-binding derivative thereof.

1                    17. The cell of claim 16, wherein said cytotoxic T  
2                    lymphocyte is a CD8<sup>+</sup> cytotoxic T lymphocytes.

1                    18. The cell of claim 11, wherein said extracellular  
2                    portion is an immunoglobulin molecule, or an antigen-binding  
3                    fragment thereof.

1                    19. DNA encoding a chimeric receptor of claim 11.

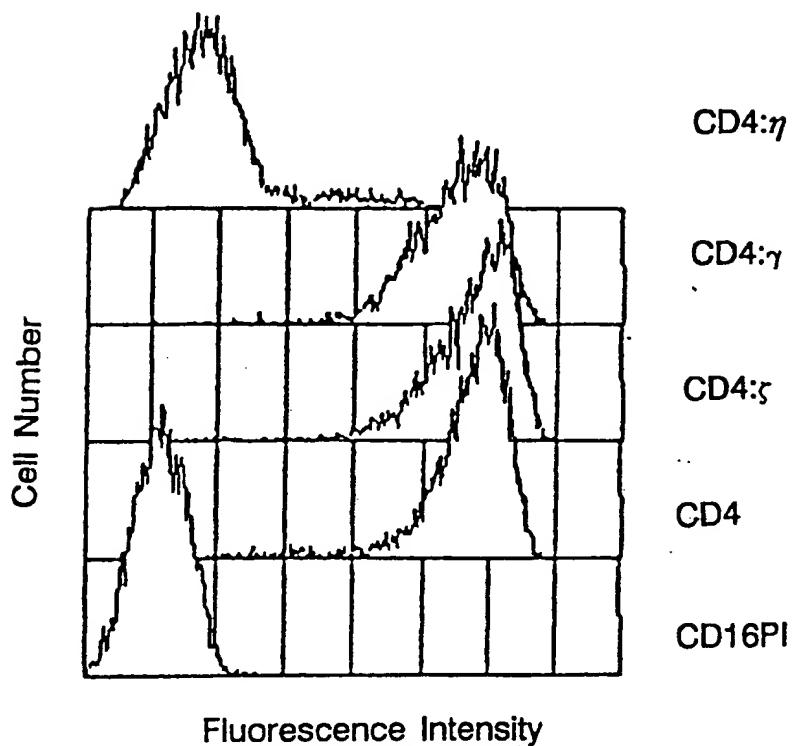
1                    20. A vector comprising the chimeric receptor DNA of  
2                    claim 19.

1                    21. A method of treating an immunodeficiency virus  
2                    infection, comprising administering the host cell of claim 16  
3                    to a mammal.

1                    22. A substantially pure antibody which specifically  
2                    recognizes and binds a chimeric receptor of claim 11.

	1	11	15
5	QSFG <b>LLDPK</b>	LCYLLDG--	11
	369		
CD4: $\zeta$	--PTWSTPV <b>HADPK</b>	LCYLLDG--	12
	1		
$\gamma$	L <b>GEPQ</b>	LCYILDA--	17
	369		
CD4: $\gamma$	--PTWSTPV <b>HADPQ</b>	LCYILDA--	14

**FIG. 1a**



**FIG. 1b**  
**SUBSTITUTE SHEET**

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FIG. 1C

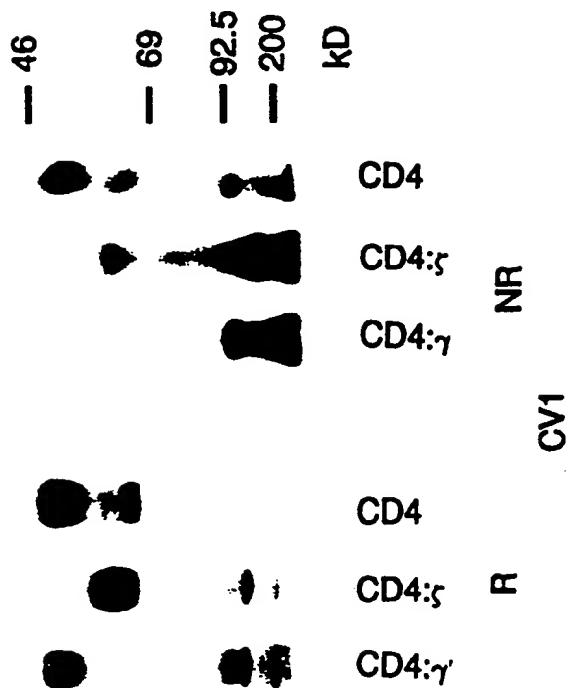
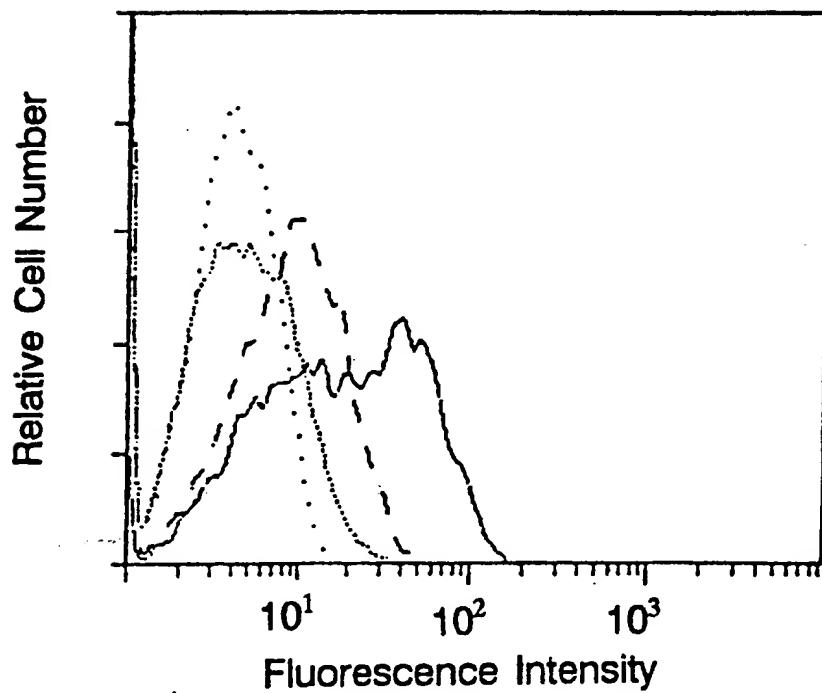
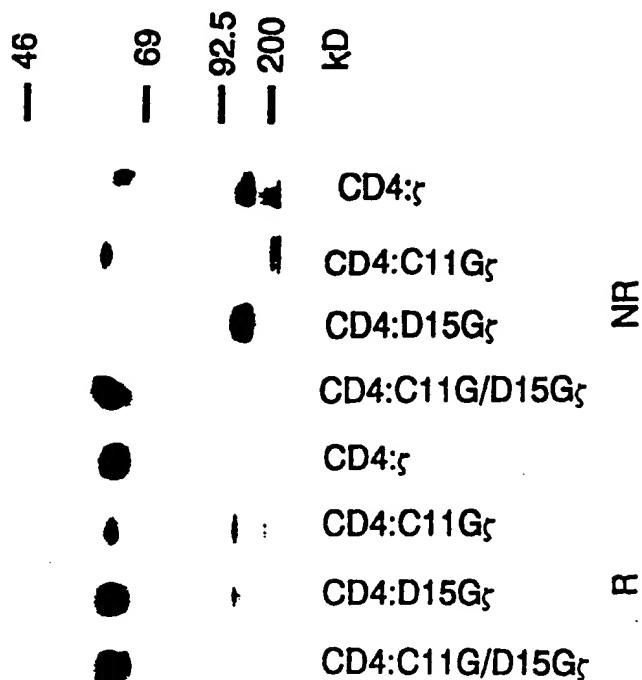
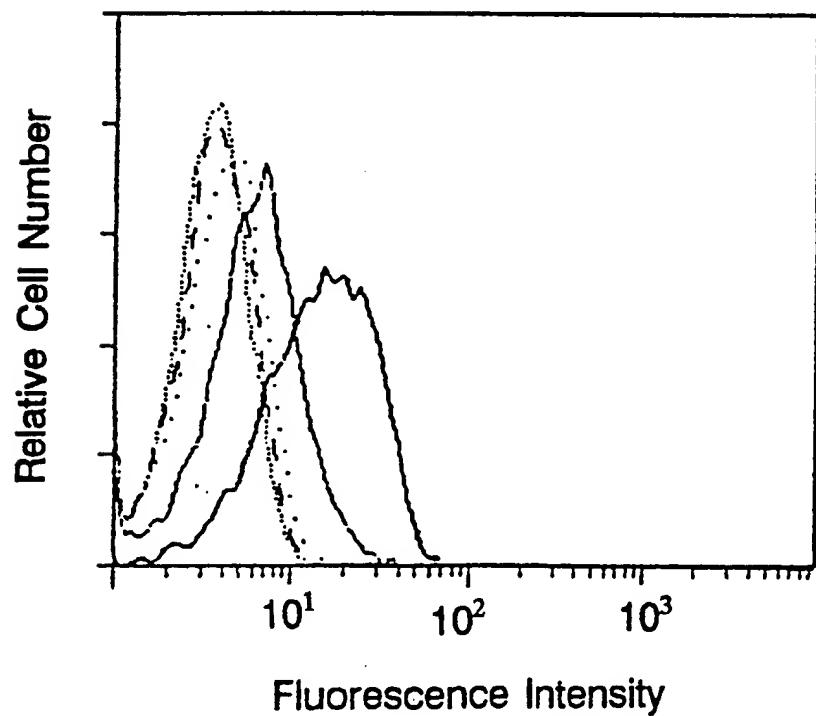


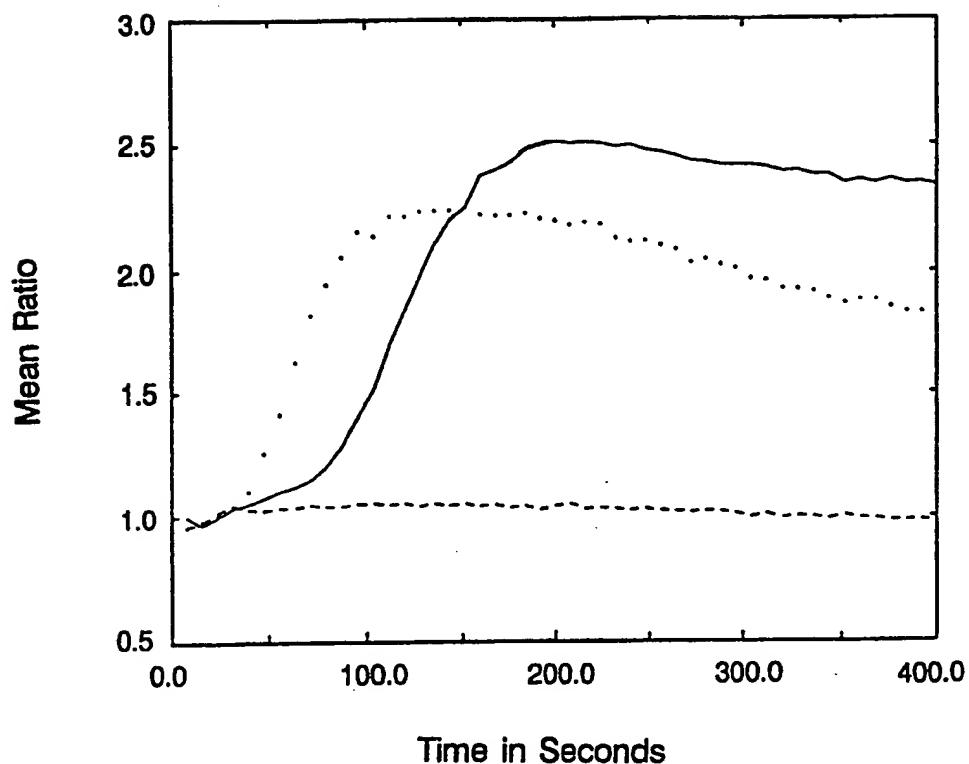
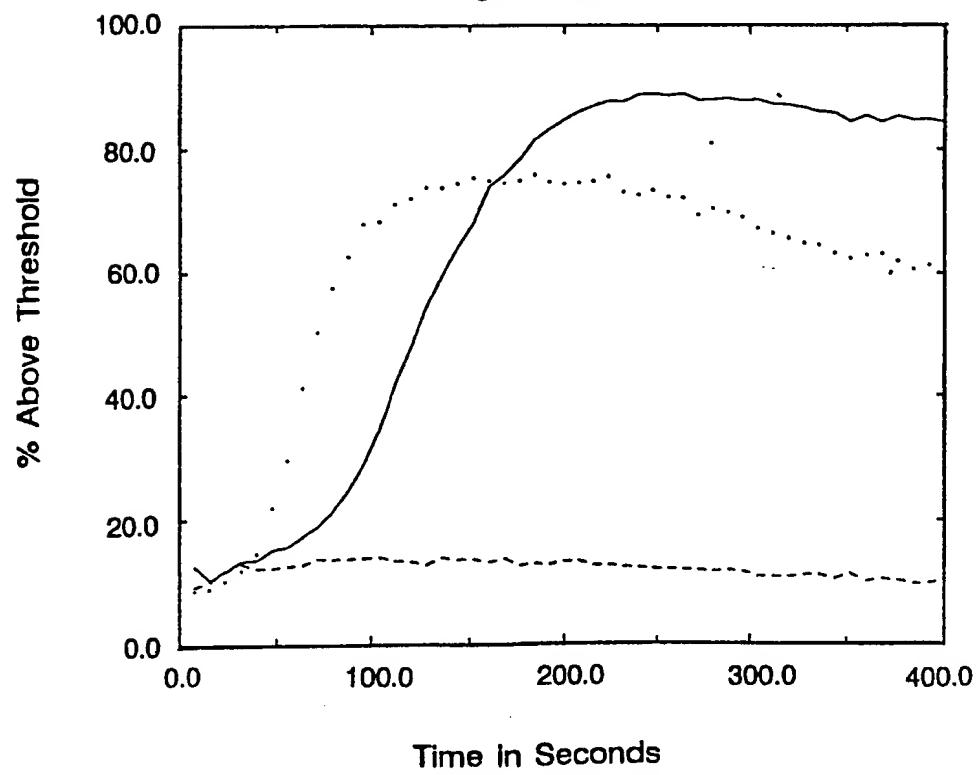
FIG. 2



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**FIG. 3a****FIG. 3b****SUBSTITUTE SHEET**

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**FIG. 4a****FIG. 4b****SUBSTITUTE SHEET**

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FIG. 4c

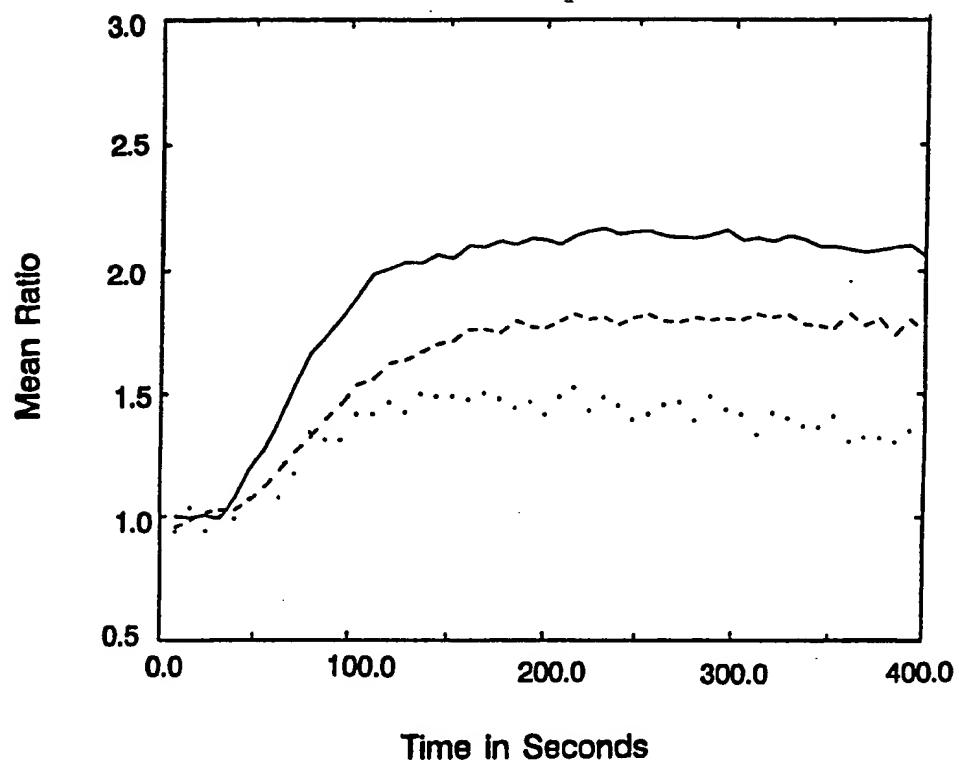
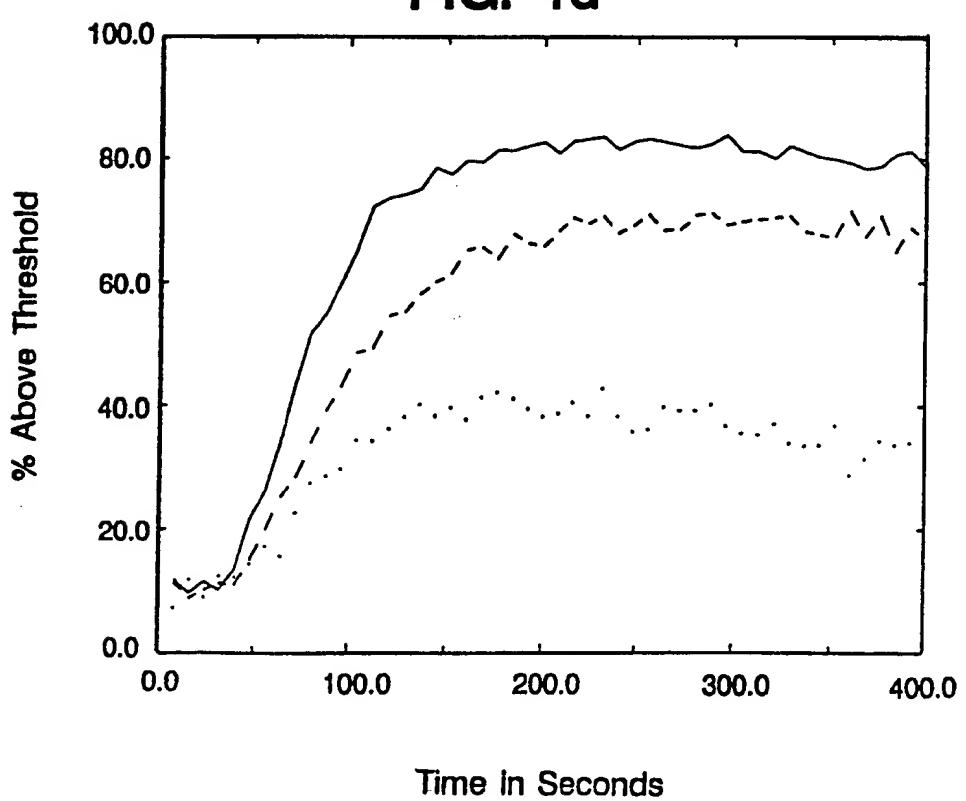
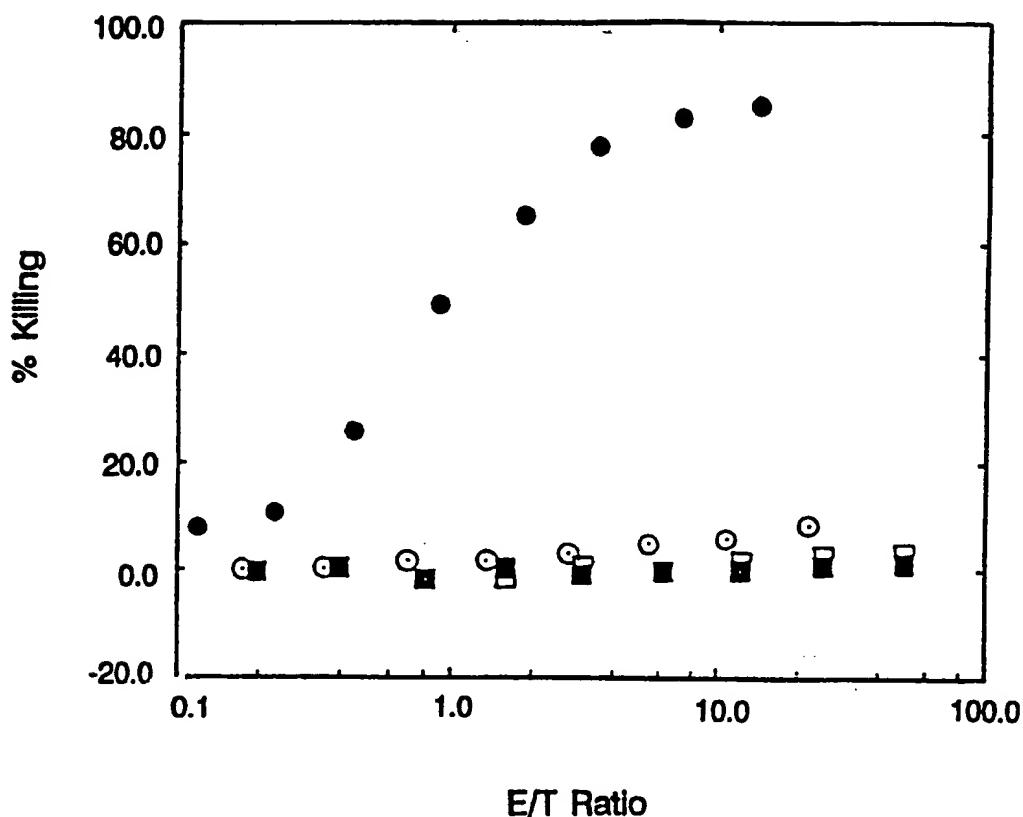
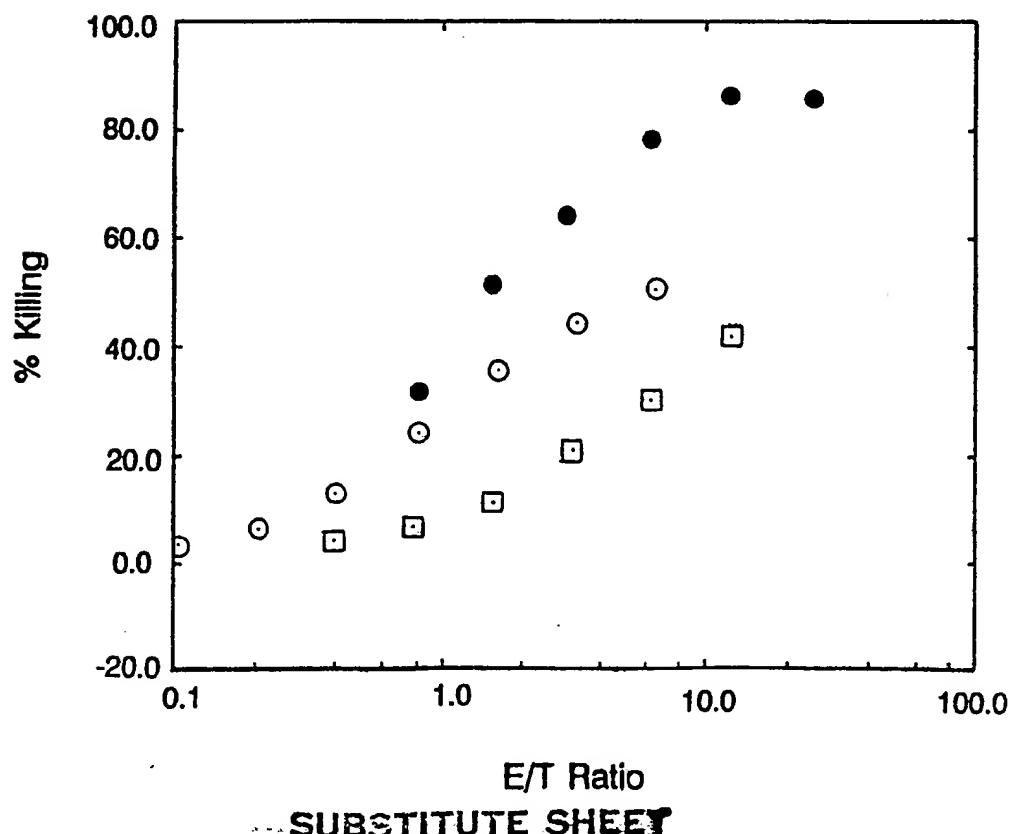
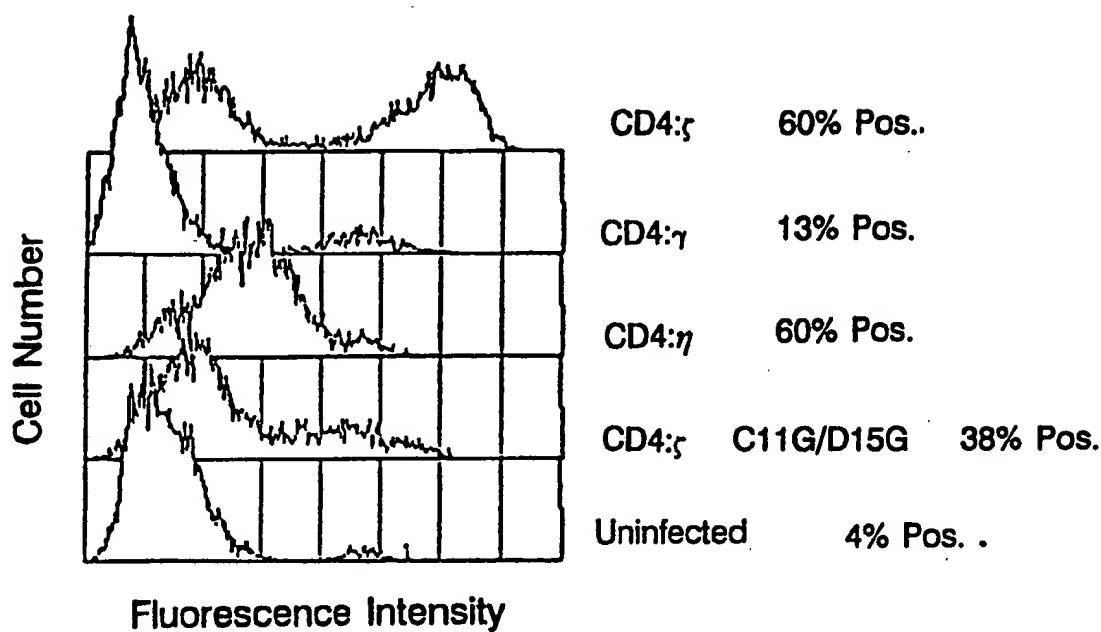


FIG. 4d

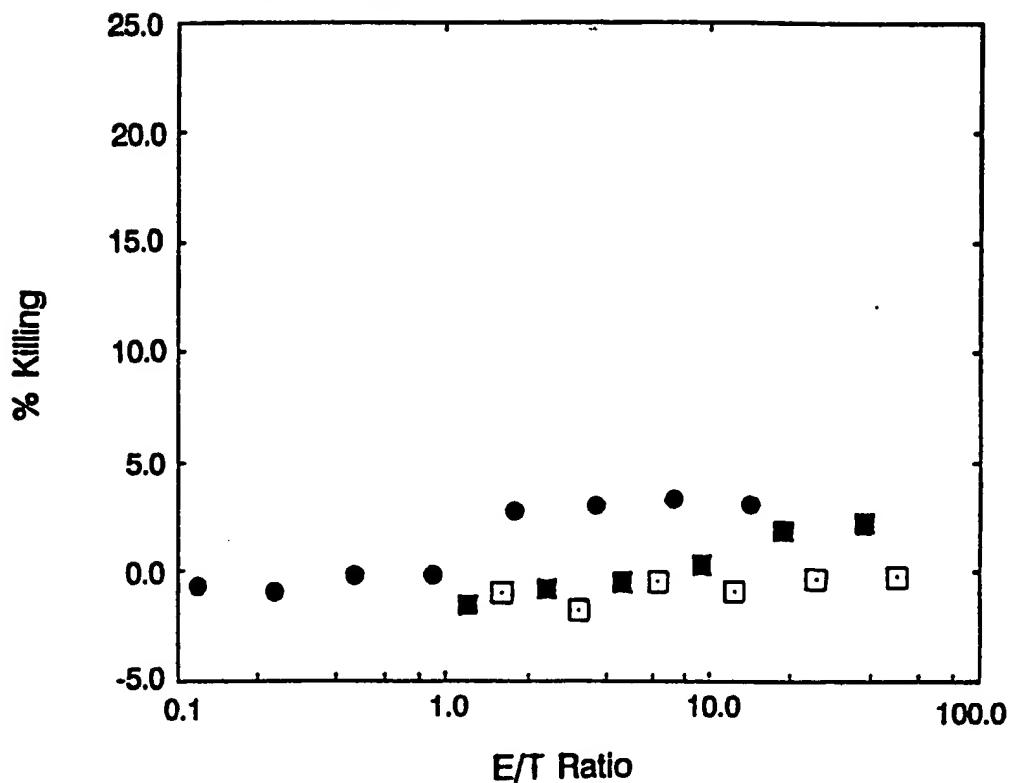
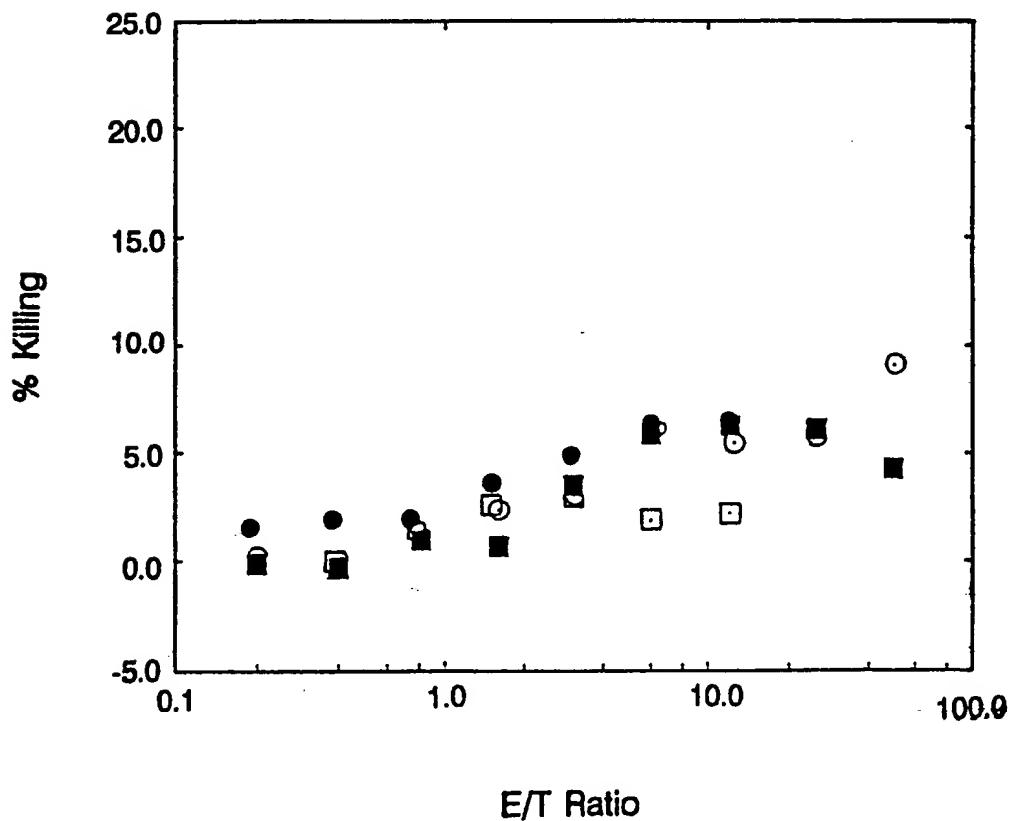


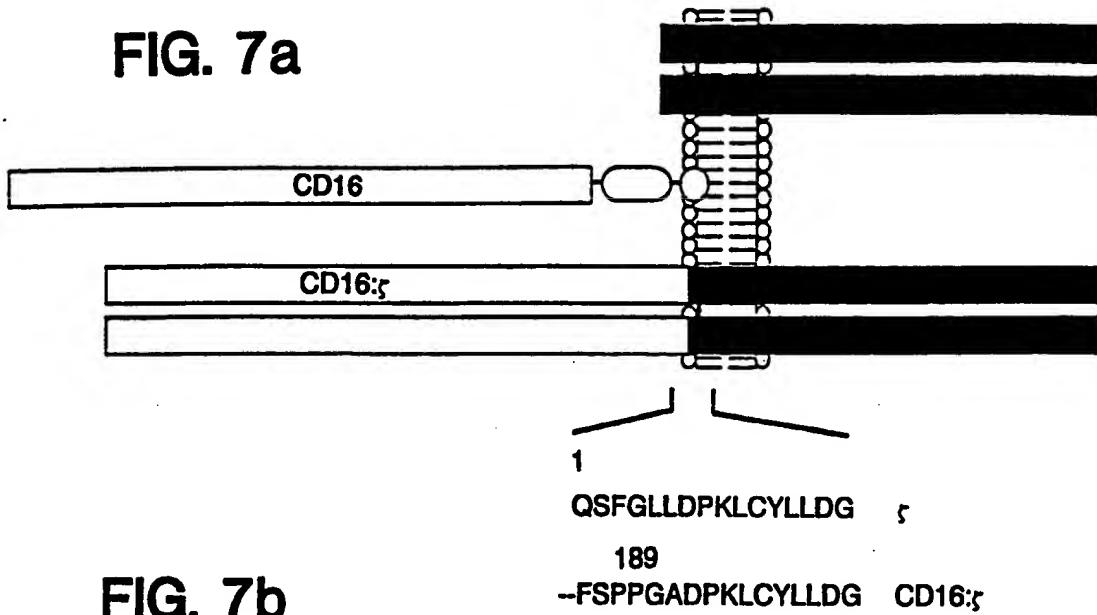
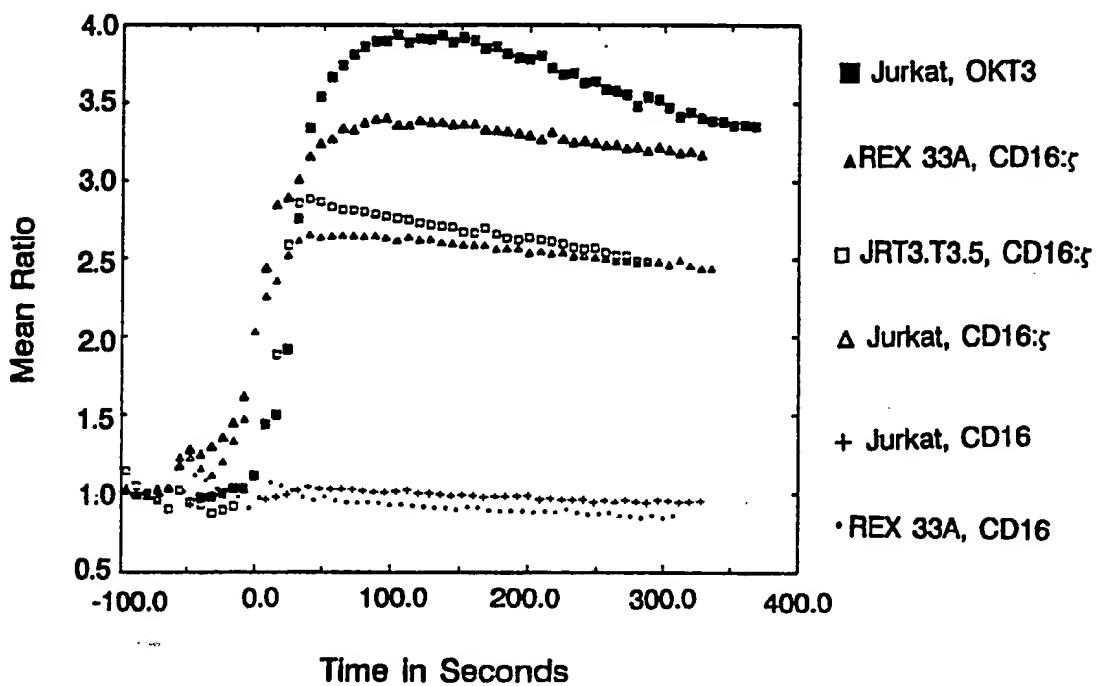
SUBSTITUTE SHEET

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**FIG. 5a****FIG. 5b****SUBSTITUTE SHEET**

**FIG. 5c****SUBSTITUTE SHEET**

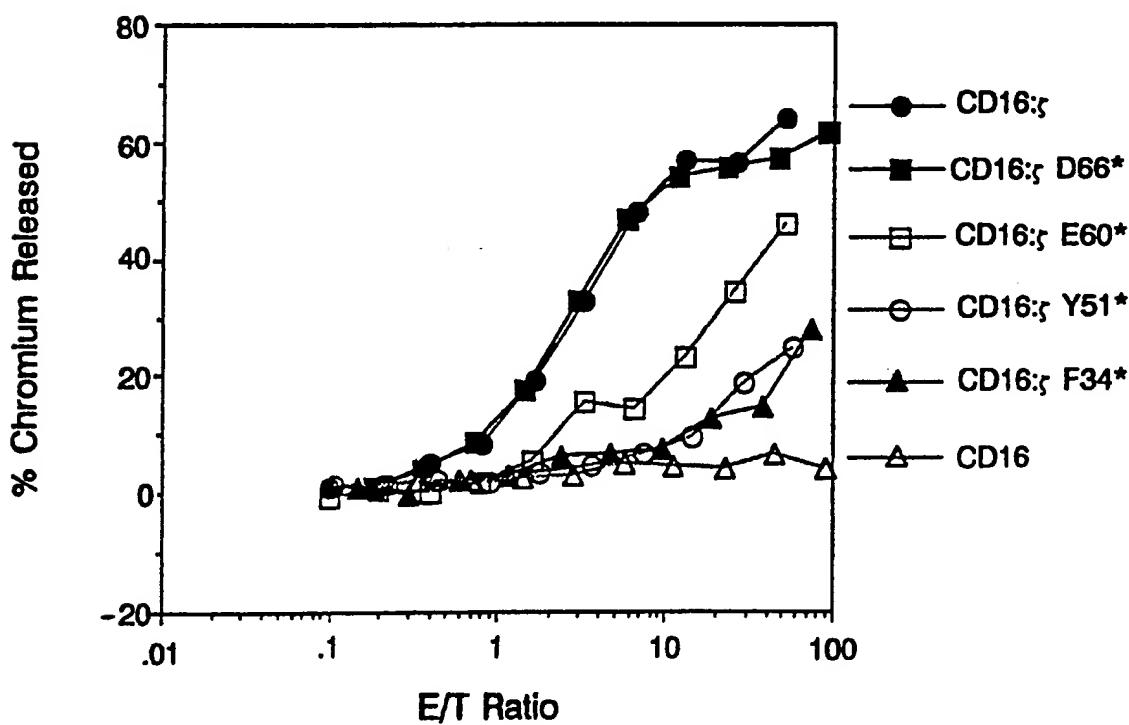
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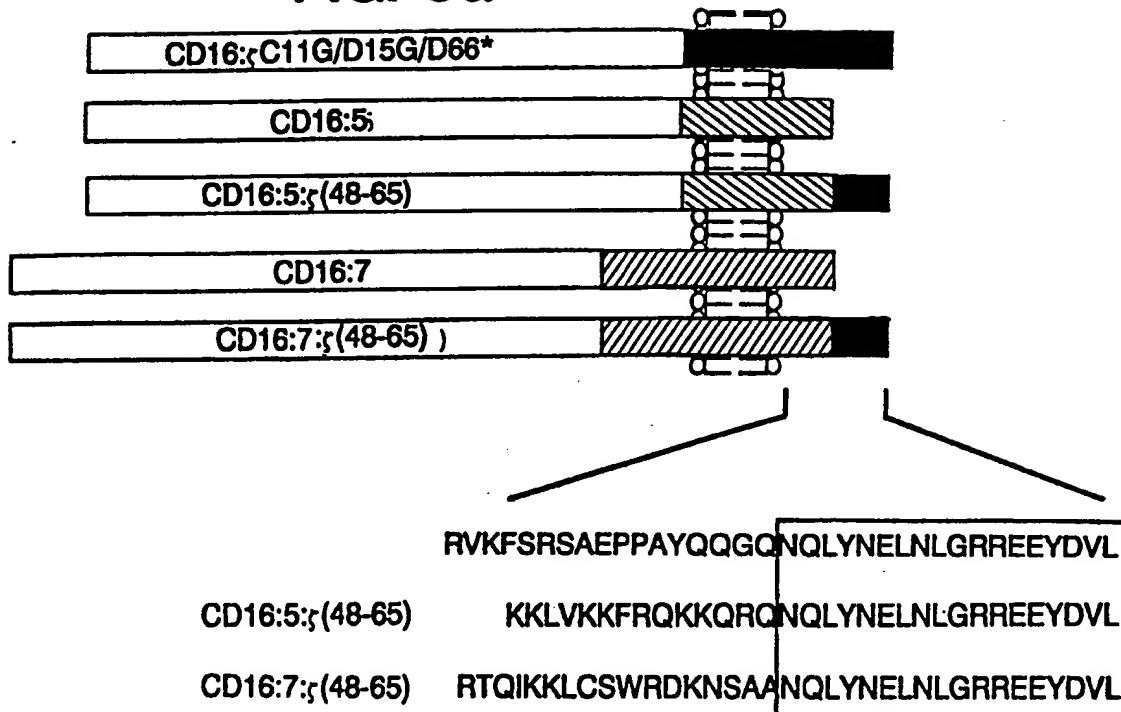
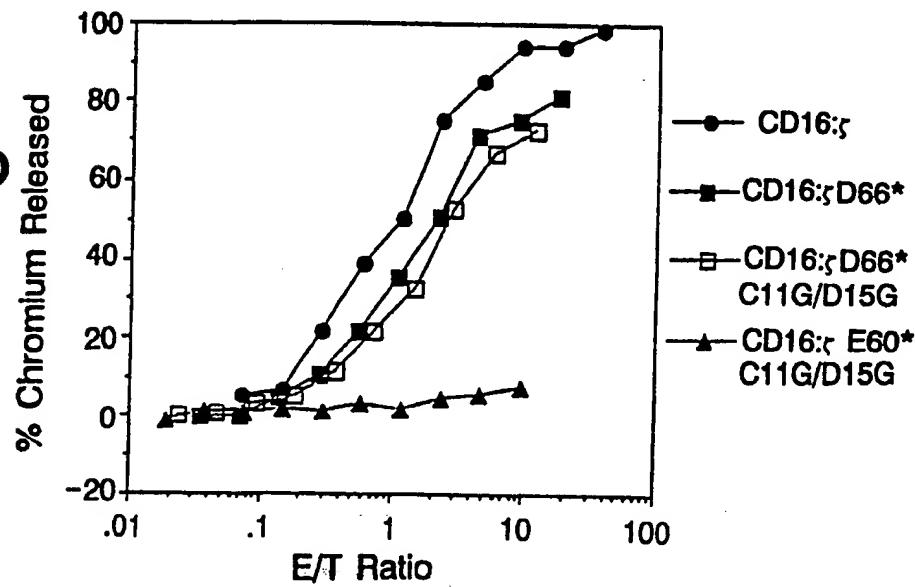
**FIG. 6a****FIG. 6b****SUBSTITUTE SHEET**

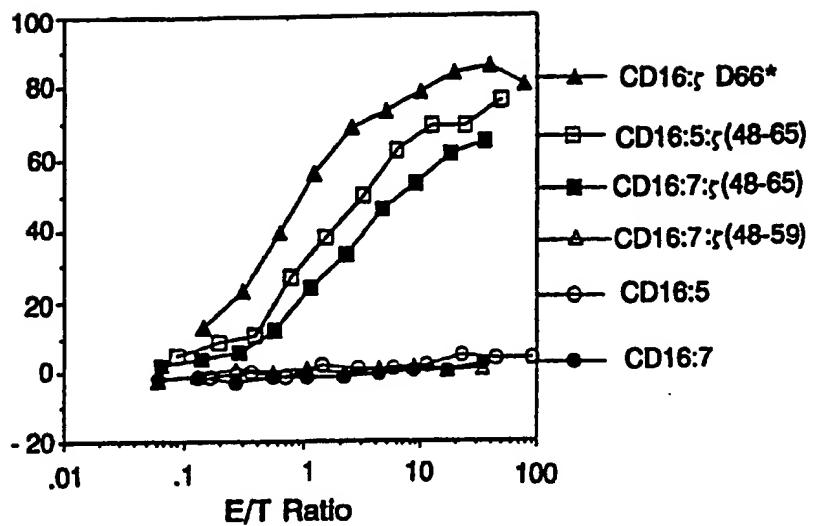
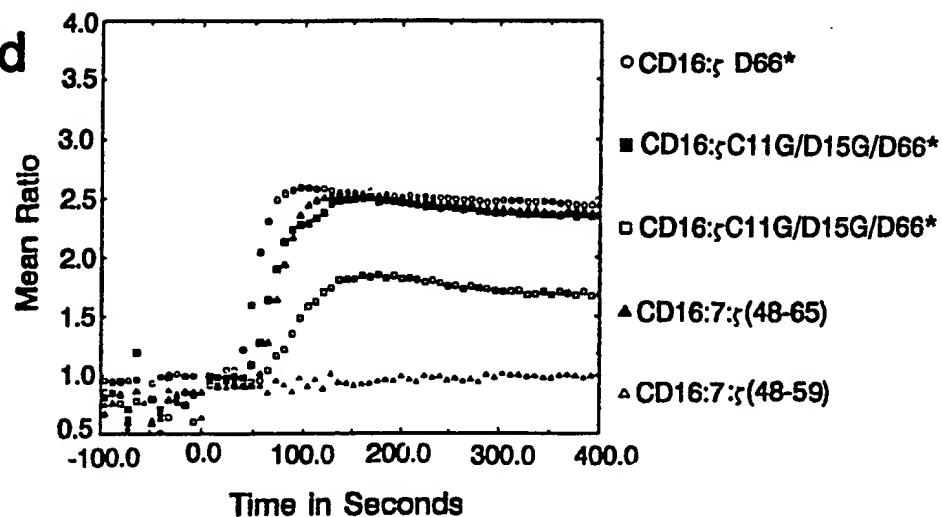
**FIG. 7a****FIG. 7b****SUBSTITUTE SHEET**

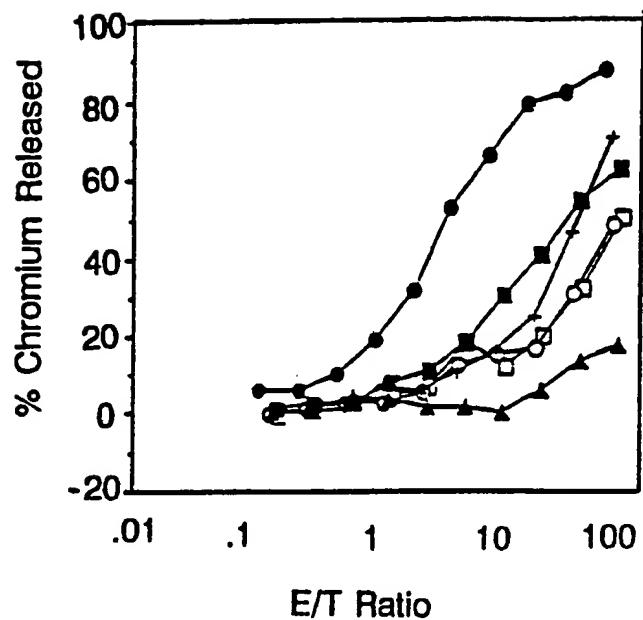
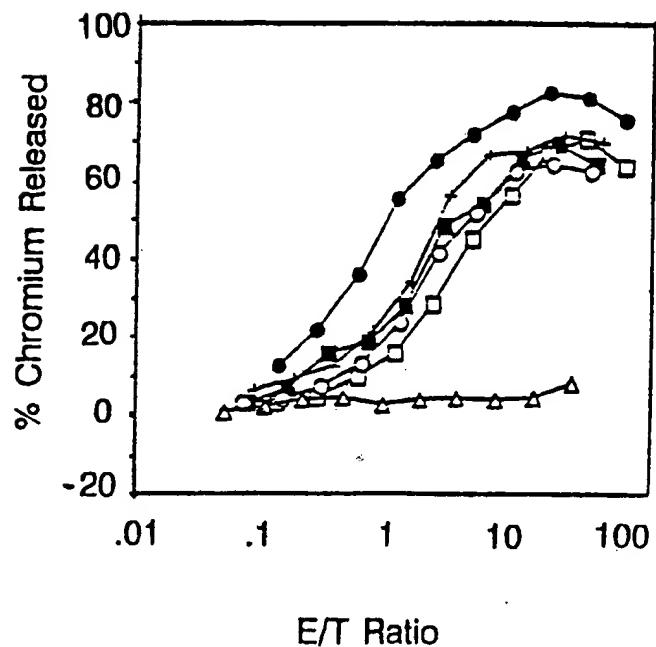
**FIG. 8a**

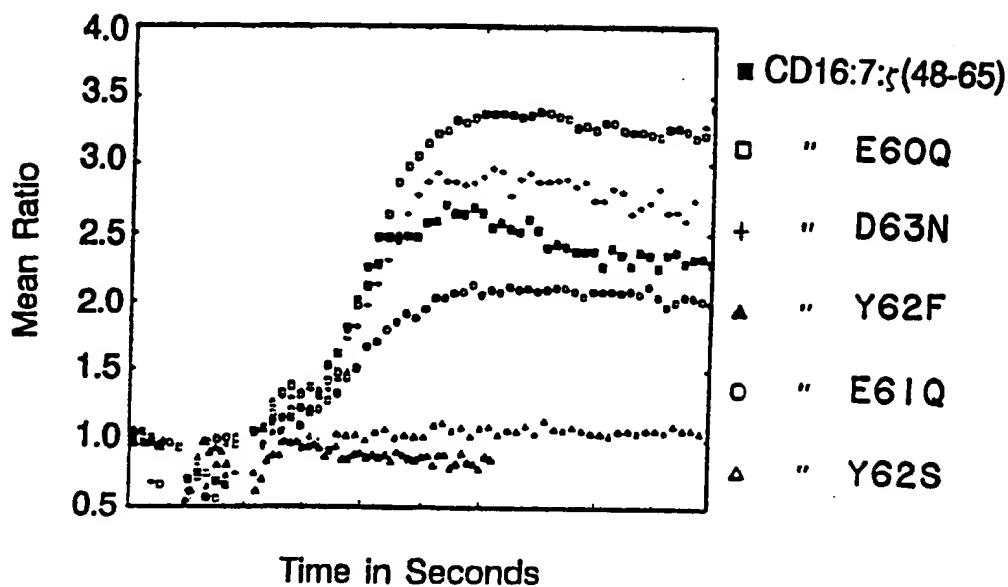
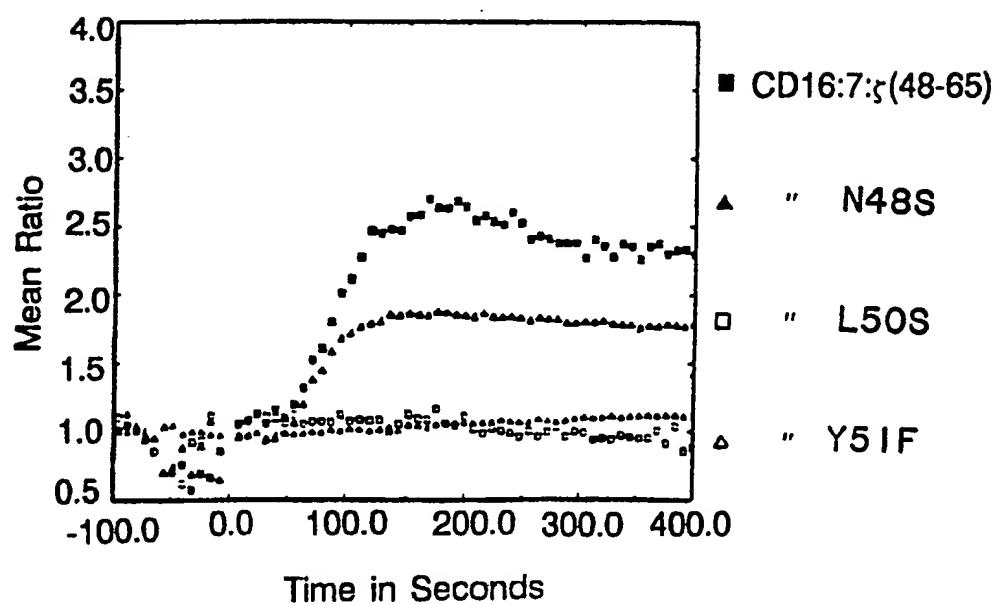
1 QSFGILLDPKL CYLLDGILFI YGVILTAFL RVKFSRSAEP PAYQQGQNQL  
 ↓      ↓  
 E60\* D66\*      F34\*      Y51\*  
 51 YNELNLGRRE EYDVLDKRRG RDPEMGGKPRRKNPQEGLYN ELQKDKMAEA  
 ↓      ↓  
 G122\*      A133\*      L139\*  
 101 YSEIGMKGER RRGKGHDGLY QGLSTATKDT YDALHMQALP PR  
 ↓      ↓      ↓

**FIG. 8b**

**FIG. 9a****FIG. 9b****SUBSTITUTE SHEET**

**FIG. 9c****FIG. 9d****SUBSTITUTE SHEET**

**FIG. 10a****FIG. 10b****SUBSTITUTE SHEET**

**FIG. 10c****FIG. 10f****SUBSTITUTE SHEET**

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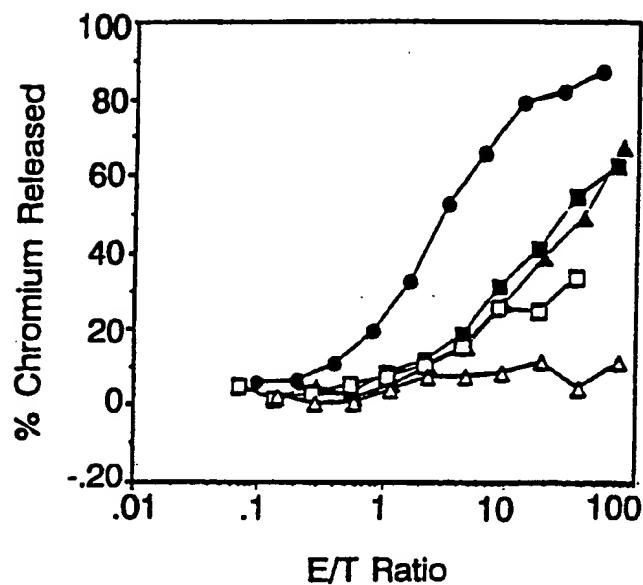
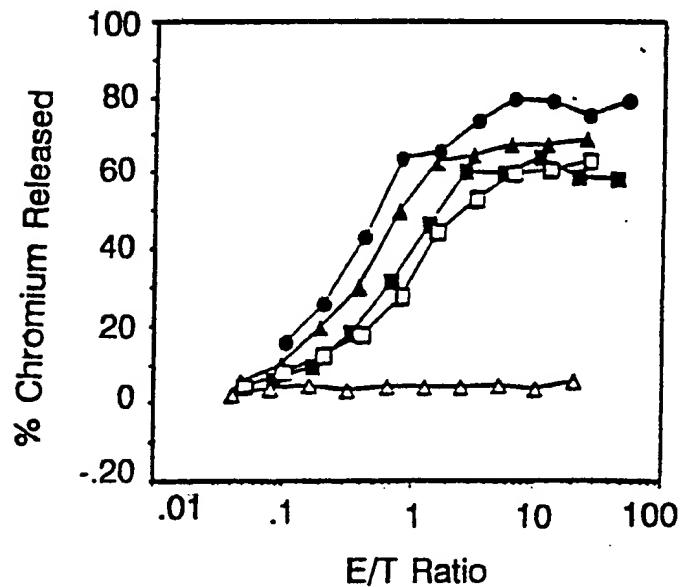
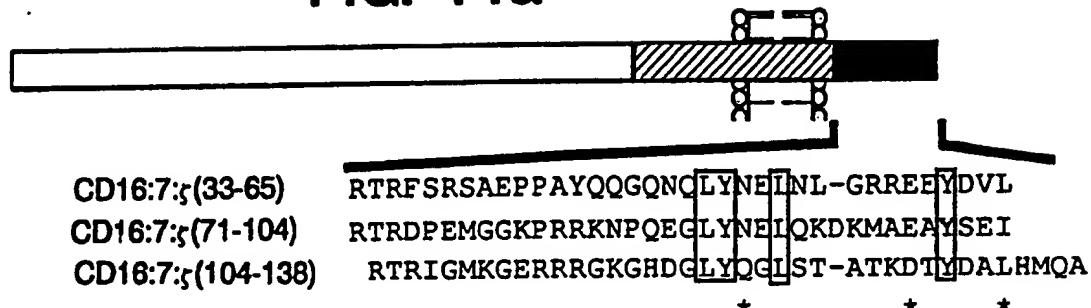
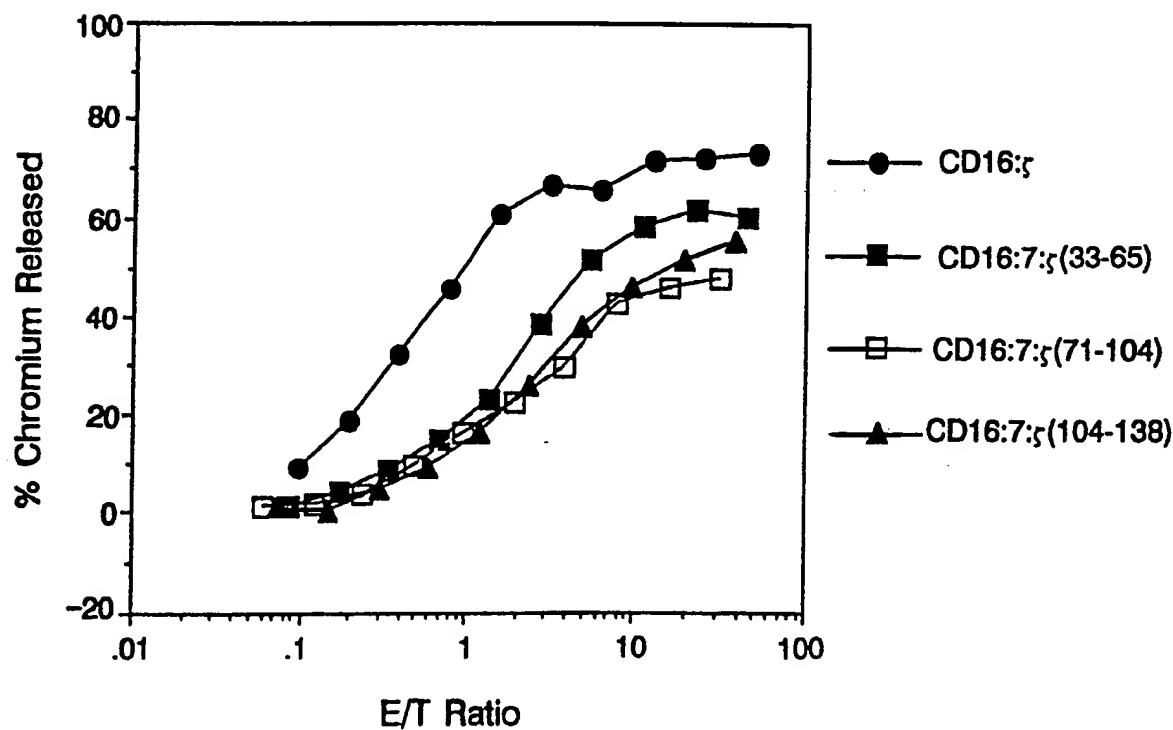
**FIG. 10d****FIG. 10e****SUBSTITUTE SHEET**

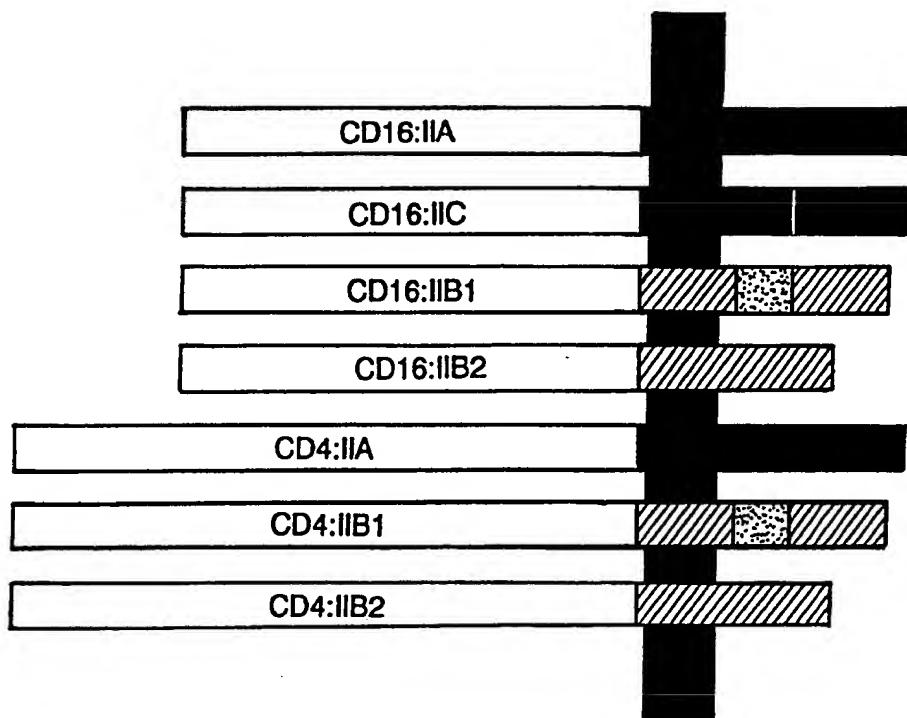
FIG. 11a

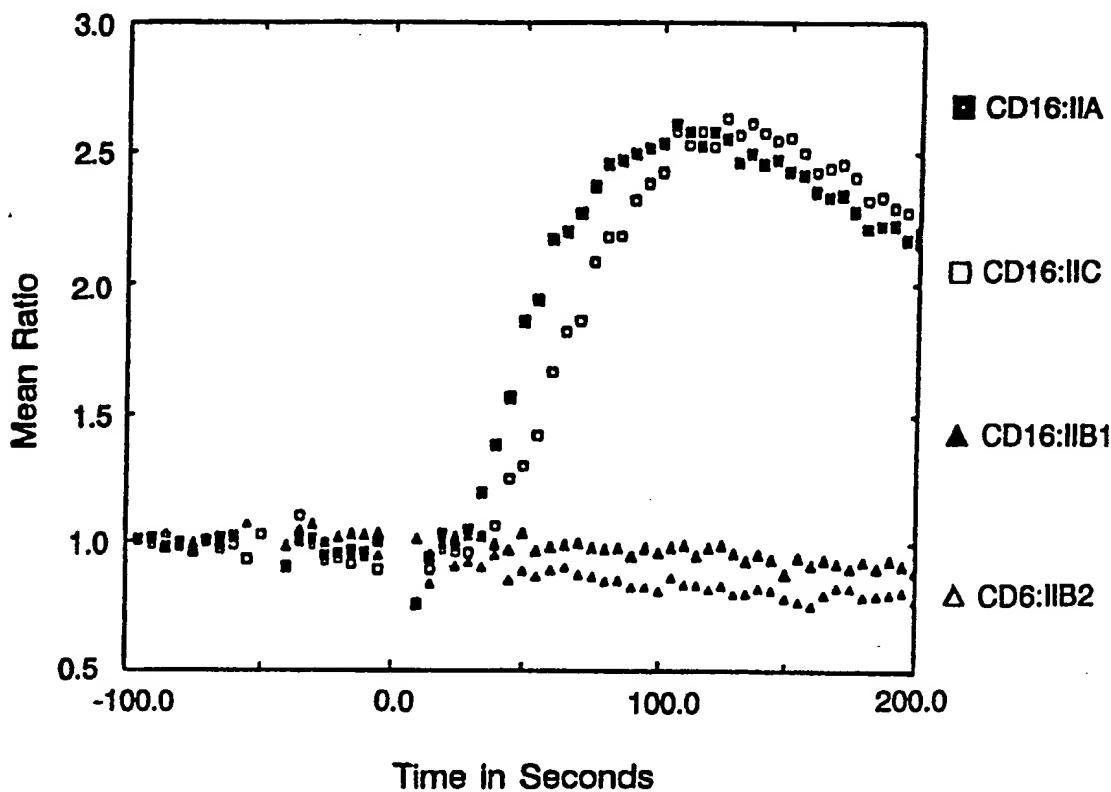
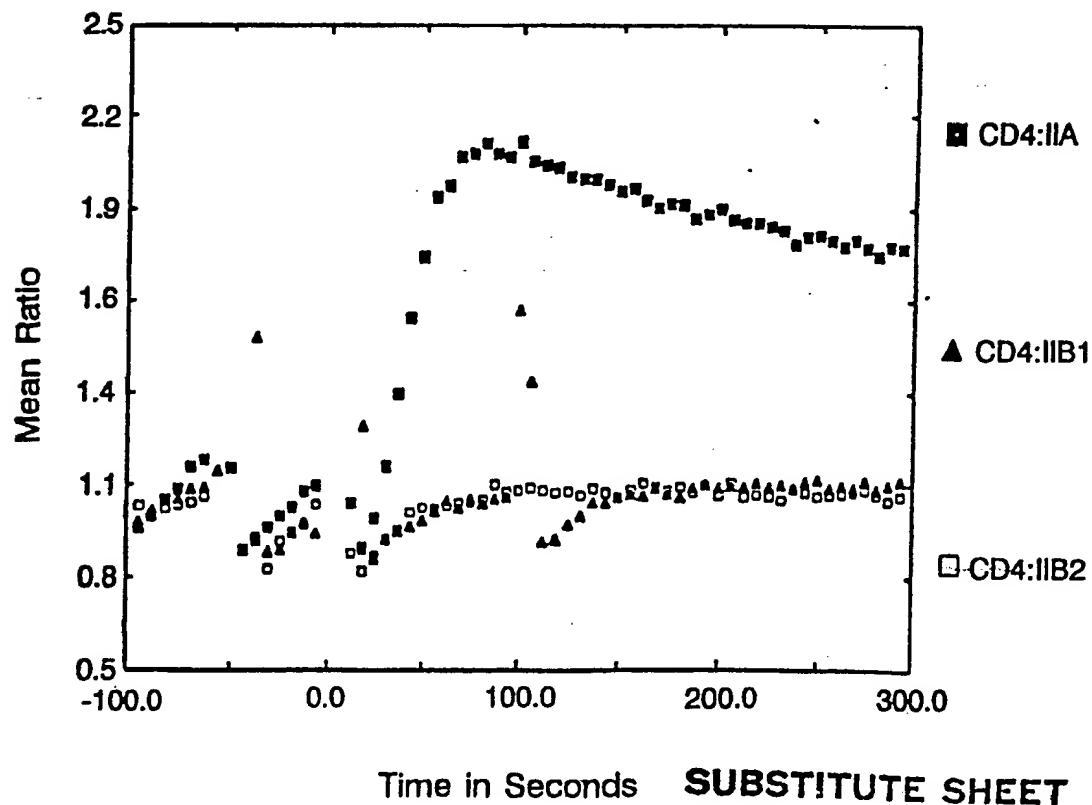


**FIG. 11b**



## **SUBSTITUTE SHEET**

**FIG. 12**

**FIG. 13a****FIG. 13b**

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FIG. 14a

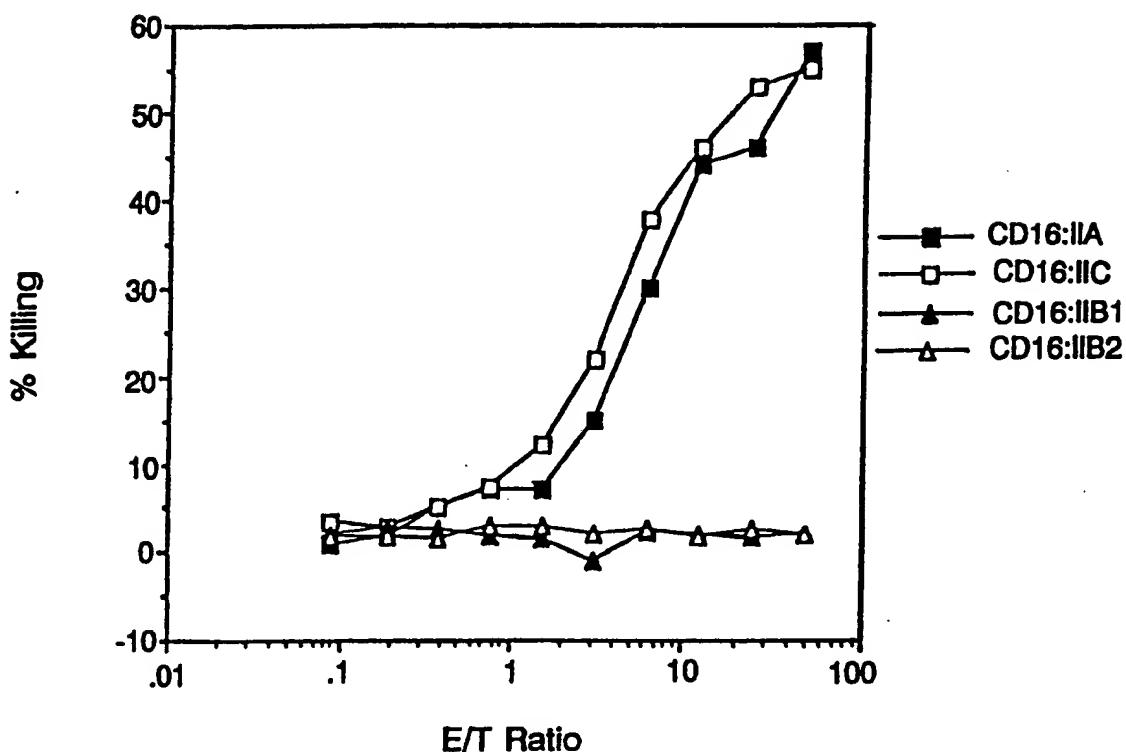
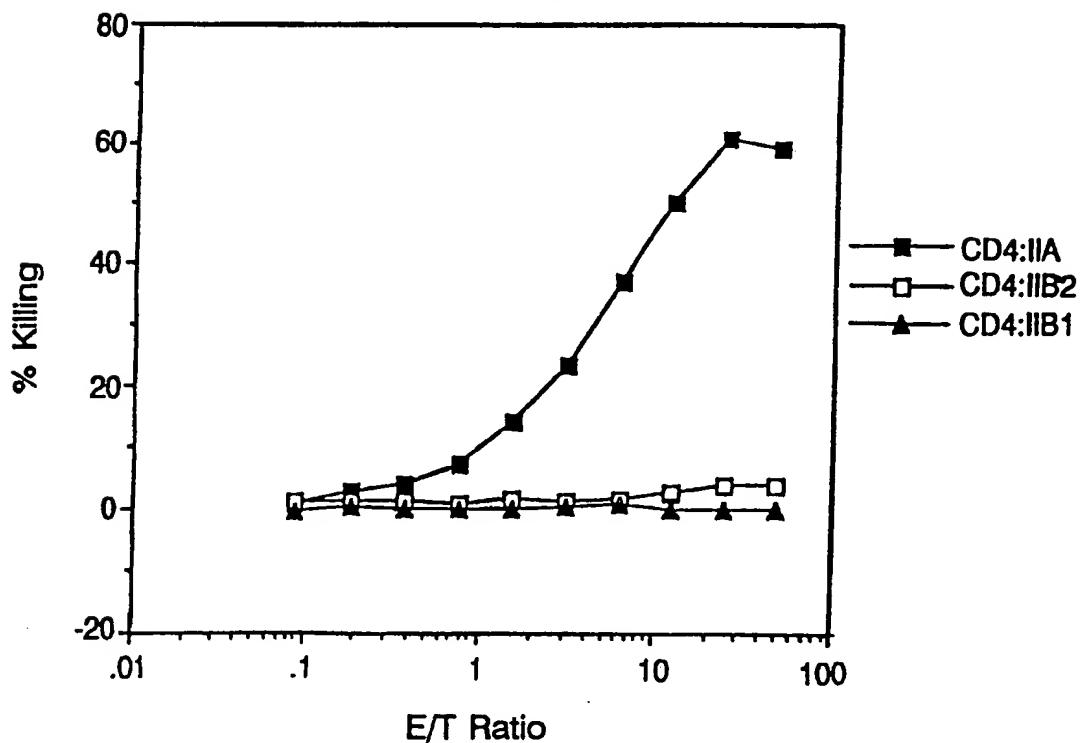


FIG. 14b



SUBSTITUTE SHEET

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FIG. 15a

(269-311)

236 RKKRISANST DPVKAAQFEP PGRQMIAIRK RQLEETNNNDY

(276-311) (282-311) (299-311)

276 ETADGGYMTL NPRAPTDDDK NIYLTLPND HVNSNN

Y282\* Y298\*

FIG. 15b

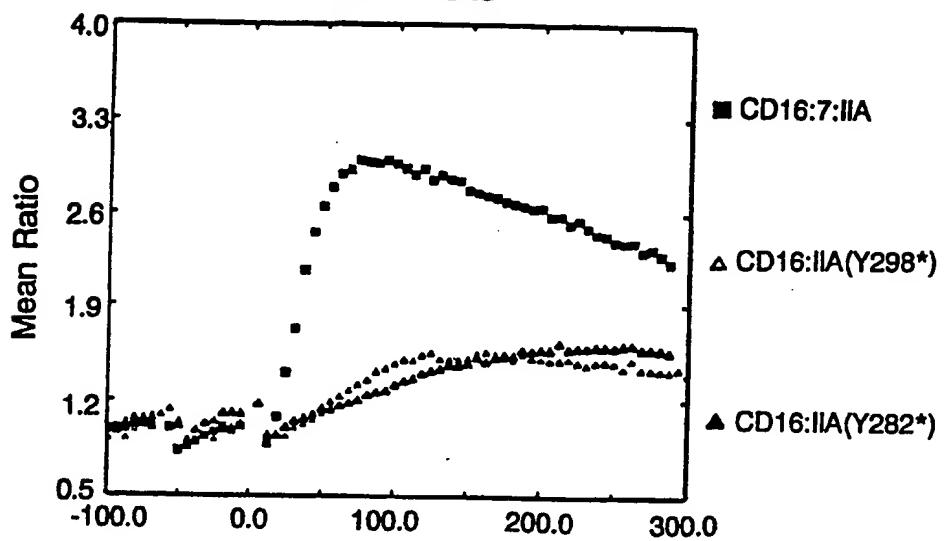
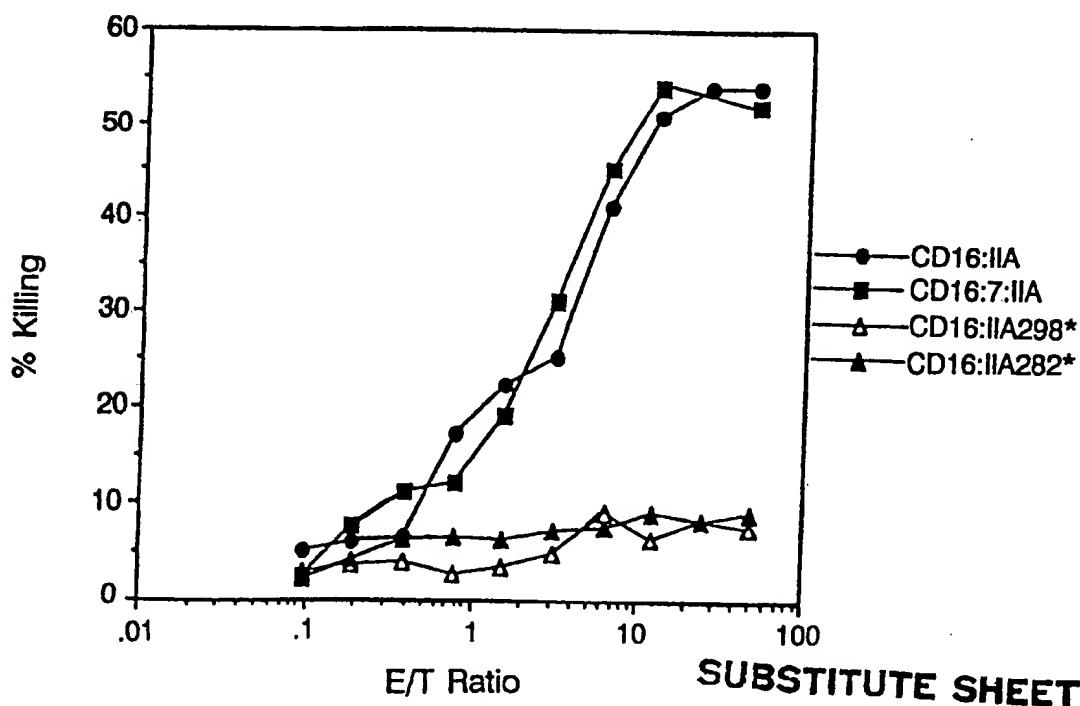


FIG. 15c



SUBSTITUTE SHEET

FIG. 15d

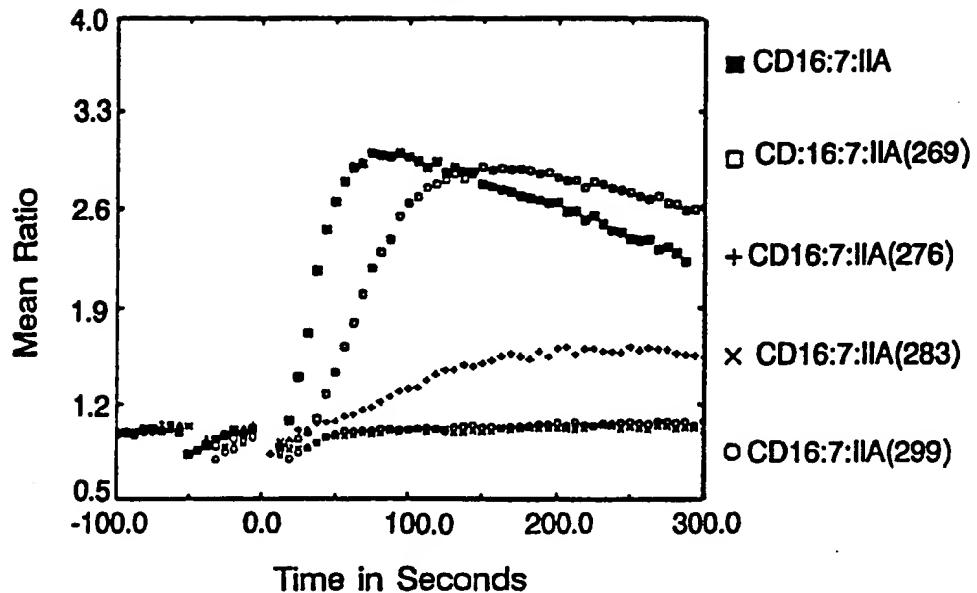
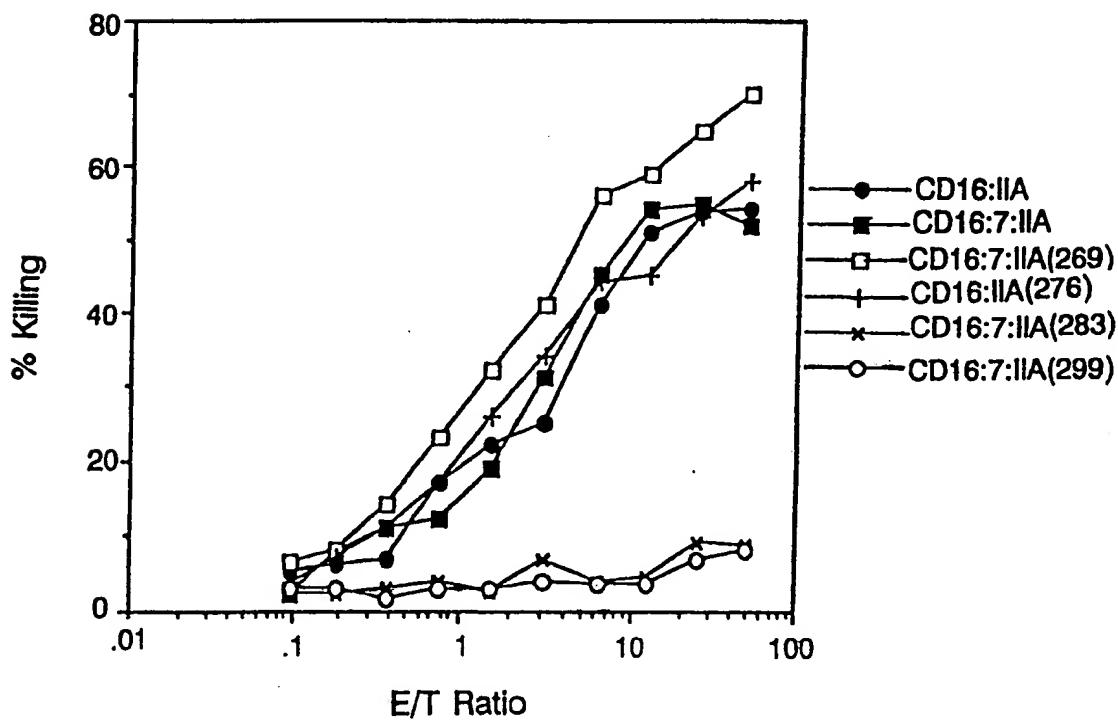


FIG. 15e



SUBSTITUTE SHEET

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**FIG.16 (Seq. ID No: 24)**

1	MEHSTFLSGL	VLATLLSQVS	PKIPIEEL	DRVFVNCNTS	ITWVEGTVGT
51	LLSDITRLDL	GKRILDPRGI	YRCNGTDIYK	DKESTVQVHY	RMCQSCVELD
101	PATVAGIIVT	DVIATLLLAL	GVFCFAGHET	<u>GRLSGAADTQ</u>	<u>ALLRNDQVYQ</u>
151	<u>PLRDRDDAQY</u>	<u>SHLGGNWARN</u>	<u>K*</u>		

**FIG.17 (Seq ID No: 25)**

1	MEQGKGLAVL	ILAIILLQGT	LAQSIKGHNHL	VKVYDYQEDG	SVLLTCDAEA
51	KNITWFKDGK	MIGFLTEDKK	KWNLGSNAKD	PRGMYQCKGS	QNKSPLQVY
101	YRMCQNCIEL	NAATISGFLF	AEIVSIFVLA	VGVYFIAGQD	<u>GVRQSRASDK</u>
151	<u>QTLLPNDQLY</u>	<u>QPLKDREDDQ</u>	<u>YSHLQGNQLR</u>	<u>RN*</u>	

**FIG.18 (Seq ID No: 26)**

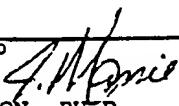
1	MPGGLALEALRA	LPLLLFLSYA	CLGPGCQALR	VEGGPPSLTV	NLGEEARLTC
51	ENNGRNPNIT	WWFSLQSNIT	WPPVPLGPGQ	GTTGQLFFPE	VNKNTGACTG
101	CQVIENNILK	RSCGTYLVRV	NPVPRPFLDM	GEGTKNRIIT	AEGIILLFCA
151	VVPGTLLLFR	<u>KRWQNEKFGV</u>	DMPDDYEDEN	LYEGLNLDDC	<u>SMYEDISRGL</u>
201	<u>QGTYQDVGNL</u>	<u>HIGDAQLEKP</u>	*		

**FIG.19 (Seq ID No: 27)**

1	MATLVLSSMP	CHWLLFLLLL	FSGEPVPAMT	SSDLPLNFQG	SPCSQIWQHP
51	RFAAKKRSSM	VKFHCYTNHS	GALTWFRKRG	SQQPQELVSE	EGRIVQTQNG
101	SVYTLTIQNI	QYEDNGIYFC	KQKCDSANHN	VTDSCGTELL	VLGFSTLDQL
151	KRRNTLKDGI	ILIQTLLIIL	FIIVPIFLLL	<u>DKDDGKAGME</u>	<u>EDHTYEGLNI</u>
201	<u>DQTATYEDIV</u>	<u>TLRTGEVKWS</u>	<u>VGEHPGQE*</u>		

# INTERNATIONAL SEARCH REPORT

International Application No. PCT/US92/01785

<b>I. CLASSIFICATION OF SUBJECT MATTER</b> (If several classification symbols apply, indicate all) <sup>3</sup>		
According to International Patent Classification (IPC) or to both National Classification and IPC		
IPC (5) : A61K 37/12; C07K 3/00, 13/00, 15/00, 17/00		
US CL : 530/387		
<b>II. FIELDS SEARCHED</b>		
<b>Minimum Documentation Searched<sup>4</sup></b>		
Classification System	Classification Symbols	
U.S.	530/387	
<b>Documentation Searched other than Minimum Documentation to the extent that such Documents are included in the Fields Searched<sup>5</sup></b>		
APS, Dialog, Intelligenetics		
<b>III. DOCUMENTS CONSIDERED TO BE RELEVANT<sup>14</sup></b>		
Category <sup>6</sup>	Citation of Document <sup>15</sup> with indication, where appropriate, of the relevant passages <sup>17</sup>	Relevant to Claim No. <sup>18</sup>
X, P	Cell, Vol. 64, issued 08 March 1991, C.Romeo & B. Seed, "Cellular immunity to HIV activated by CD4 fused to T cell or Fc receptor polypeptides", pages 1037-1046, see entire document.	1-22
Y	Proc. Natl. Acad. Sci. USA, Vol. 85, issued December 1988, A.M. Weissman et al., "Molecular cloning and chromosomal localization of the human T-cell receptor zeta chain: Distinction from the molecular CD3 complex", pages 9709-9713, see entire document.	1-22
Y	J. Biol. Chem., Vol 264, No. 35, issued 15 December 1989, S.A. Carr et al., "Protein and carbohydrate structural analysis of a recombinant soluble CD4 receptor by mass spectrometry", pages 21286-21295, see entire document.	1-22
Y	Proc. Natl. Acad. Sci. USA, Vol. 83, issued May 1986, P. van den Elsen et al., "Exon/intron organization of the genes coding for the $\delta$ chains of the human and murine T-cell receptor/T3 complex", pages 2944-2948, see entire document.	1-22
<p><b>* Special categories of cited documents:<sup>16</sup></b></p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</p> <p>"&amp;" document member of the same patent family</p>		
<b>IV. CERTIFICATION</b>		
Date of the Actual Completion of the International Search <sup>2</sup>	Date of Mailing of this International Search Report <sup>2</sup>	
03 June 1992	12 JUN 1992	
International Searching Authority <sup>19</sup>	Signature of Authorized Officer <sup>20</sup>	
ISA/US	 KAREN COCHRANE CARLSON, PH.D.	

## FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

Y	DNA and Cell Biology, Vol. 9, No. 5, issued 1990, G. Zettlmeissl et al., "Expression and characterization of human CD4: Immunoglobulin fusion proteins", pages 347-353, see entire document.	1-22
Y	EMBO J, Vol. 6, No. 10, issued 1987, A. Tunnacliffe et al., "Physical linkage of three CD3 genes on human chromosome 11", pages 2953-2957, see entire document.	1-22

V.  OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE<sup>1</sup>

1.  Claim numbers \_\_, because they relate to subject matter (1) not required to be searched by this Authority, namely:

2.  Claim numbers \_\_, because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out (1), specifically:

3.  Claim numbers \_\_, because they are dependent claims not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI.  OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING<sup>2</sup>

This International Searching Authority found multiple inventions in this international application as follows:

1.  As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.

2.  As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:

3.  No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:

4.  As all searchable claims could be searched without effort justifying an additional fee, the International Search Authority did not invite payment of any additional fee.

Remark on protest

- The additional search fees were accompanied by applicant's protest.
- No protest accompanied the payment of additional search fees.

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category*	Citation of Document, <sup>16</sup> with indication, where appropriate, of the relevant passages <sup>17</sup>	Relevant to Claim No. <sup>18</sup>
Y	EMBO J, vol. 5, No. 8, issued 1986, G.W. Krissansen et al., "Primary structure of the T3 gamma subunit of the T3/T cell antigen receptor complex deduced from cDNA sequences: evolution of the T3 gamma and delta subunits", pages 1799-1808, see entire document.	1-22
Y	EMBO J, Vol. 7, No. 11, issued 1988, N. Sakaguchi et al., "B lymphocyte lineage-restricted expression of MB-1, a gene with CD3-like structural properties", pages 3457-3464, see entire document.	1-22
Y	Proc. Natl. Acad. Sci. USA, Vol. 85, issued September 1988, G.G. Hermanson et al., "B29: A member of the immunoglobulin gene superfamily exclusively expressed on B-lineage cells", pages 6890-6894, see entire document.	1-22
Y	Proc. Natl. Acad. Sci. USA, Vol. 87, issued May 1990, Y. Jin et al., "Molecular cloning of the CD3 eta subunit identifies a CD3 zeta-related product in thymus-derived cells", pages 3319-3323, see entire document.	1-22
Y	J. Biol. Chem., Vol. 265, No. 11, issued 15 April 1990, H. Kuster et al., "Characterization and expression of the gene for the human Fc receptor gamma subunit", pages 6448-6452, see entire document.	1-22